

***Kaiser Permanente Medical Center, San Francisco  
Northern California Region***

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|  | **Standard Operating Procedure** |

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| **Title:** | **IC: Laboratory Spills** | **Procedure Number** SFOSOP-0078  **Revision:** 8 |

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| **Department:**  Infection Control  **Area:**  2200 Geary Blvd SFO Clinical Lab 2238 Geary Blvd SFO Clinical Lab 2425 Geary Blvd SFO Hospital Lab 4131 Geary Blvd French Campus Lab | ***Approved & Released Standard Operating Procedure*** | **Implementation Date:**  11/22/2017 |

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| **Type of Document:**  General Lab | **Review Period -** 340 **Days** |

**PURPOSE**

To provide guidelines for proper disinfection of surface and non-critical equipment and cleaning of biological spills in the laboratory

**POLICY**

I. Use hospital-approved disinfectant when cleaning surfaces and biological spills.

**PROCEDURE**

A. Routine Cleanup of Work Area

1. Work areas must be cleaned and disinfected with hospital-approved disinfectant at the beginning and end of each shift and after spills occur.

2. Practice Standard/Universal Precautions at all times.

3. Use the hospital-approved disinfectants Sani-Cloth by PDI (Professional Disposables International, Inc.).

a. Working solution

1) Super Sani-Cloth is a germicidal disposal wipe for use on pre-cleaned environmental surfaces/objects previously soiled with blood/bloody fluids.

a) Wipe pre-cleaned surfaces with Sani-Cloth for 2 minutes at room temperature. It has the following activities:

i. **bactericidal** - *Staphylococcus aureus, Methicillin resistant Staphylococcus aureus (MRSA), Vancomycin resistant Enterococcus faecalis (VRE), Pseudomonas aeruginosa, Salmonella enterica, Burkholderia cepacia, Escherichia coli, Escherichi coli 0157:H7, ESBL producing Escherichia coli, Klebsiella pneumoniae*

ii. **tuberculocidal** - *Mycobacterium bovis BCG (tuberculosis)*

iii. **fungocidal -** *Candida albicans*

iv. **virucidal -** *HIV-1, Hepatitis B (HBV), Hepatitis C (BCV), Vaccinia virus, rotavirus strain WA, Rhinovirus, Respiratory Syncytial Virus (RSV), Herpes Simplex Type 2, Adenovirus, Human coronavirus*

b) Storage

Store in cool dry place. Do not store near heat or open flame.

c) Disposal

i. Triple rinse with water.

ii. Recycle container or toss in regular trash container.

4. Personal Protection

a. **Wear appropriate barrier protection such as gloves, mask, goggle, face shield or eye protection, and gowns when cleaning and disinfecting equipment soiled with blood or other potentially infectious materials**.

b. Wear closed-toe shoes that are impermeable to fluids.

c. Dispose of contaminated or bloody gloves, mask or eye protection, or paper towels in red biohazardous containers.

d Place dirty lab gowns in specific bins for collection by Environmental Services.

5. Precleaning surfaces or non-critical equipment surface prior to disinfection

a. Blood and other body fluids must be thoroughly cleaned from surfaces and objects before disinfection with Sani-Cloth. This is to remove excess organic debris that may prevent effective disinfection.

b. Rinse.

c. Rough dry.

d. Apply Sani-Cloth directly to surface.

e. Allow to remain wet for 5 minutes.

f. Wipe surfaces using a paper towel.

g. Discard towel.

6. Disinfecting work surfaces

a. Use Sani-Cloth to wipe precleaned surface, thoroughly wetting area to be disinfected.

b. Allow surface to remain wet for 5 minutes at room temperature.

c. Wipe dry with paper towel prior to use.

d. Discard towel.

B. Disinfecting non-critical equipment *surface*or small metals

1. Practice Standard/Universal Precautions.

2. Preclean surface; follow A.5 above.

3. Use Sani-Cloth cide to wipe precleaned surface, thoroughly wetting area to be disinfected.

4. Allow surface to remain wet for 3 minutes at room temperature.

5. Wipe dry with paper towel prior to use.

C. Disinfecting non-critical *instruments*or small metals

1. Practice Standard/Universal Precautions.

2. Preclean to remove excess organic debris, rinsed and rough dried; follow A.5 above.

3. Clean and rinse lumens of hollow instruments before filling with solution or before immersion.

4. Using a soaking tray or an ultrasonic unit, immerse instruments into 10% bleach solution for at least3 minutes.

5. Remove and rinse instruments.

6. Wipe dry with paper towel prior to use.

D. Disinfection of spills of blood/body fluids, other clinical specimens, or microbiological specimens

1. Practice Standard/Universal Precautions. Protect hands with gloves. Wear goggles and/or face masks if splashes into the face may occur.

2. Cover spill with disposable towels to absorb liquid and prevent the formation of aerosols.

3. Disinfect by liberally swabbing with Sani-Cloth.

4. Dispose of towels in red bags.

5. Dispose of sharps in rigid, leak-proof sharps container.

E. Disinfection of tube carriers when specimen containers break or leak

1. If the spill or breakage is contained within a biohazard bag, dispose of the specimen according to procedure. See Infection Control procedure: "Disposal of Infectious Wastes".

2. If other intact specimens are in the same bag with the broken specimen,

a. Prepare a chuck or gauze on which to decontaminate them.

b. Wearing gloves, remove the intact specimens with tongs or forceps. **Do not put your hand in the bag.**

c. Place the specimens in the gauze or chuck and proceed with decontamination with Sani-Cloth as in section D.

3. If the biohazard bag is punctured or otherwise damaged or leaking:

a. Discard the liner along with the broken specimen according to procedure. See Infection Control procedure "Disposal of Infectious Waste".

b. Decontaminate the outer carrier with hospital-approved disinfectant.

F. While the HIV-1 virus is inactivated in 2 minutes, use the recommended contact time for disinfection of 5 minutes.

G. For tuberculocidal activity, allow 5 minutes after wiping with Sani-Cloth at room temperature. Wipe surface using a paper or cloth towel, allow to air dry, then discard towel.

I. Use Sani-Cloth to wipe surfaces.

J. For larger spills, refer to SDS for any chemicals involved. If over 1 liter, or if spill involves chemicals of high risk, review Rainbow Chart, then contact Facility Safety Officer for clean up (do not contact EVS for larger chemical spills).

H. Attached below is instructions for alternative way to clean up spill. Use this kit to clean up spill up to 1 liter.



**REFERENCES**

1. Website: http://www.cdc.gov/ohs/biosfty/bleachiv.itm

2. College of American Pathologists, Lab General GEN.73500, 74500, 74600 *Current Edition*

**Associated Documents:**

External Documents:

Associated Documents:

SFOSOP-0125 -- SFTY - Chemical Hygiene Plan  
SFOSOP-0080 -- IC: Laboratory Wastes (CAP GEN.70600 70650 73000 73100 70700)

Click to Open an Associated Document

**Documents Generated:**

**Document Revision History:**

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| --- | --- | --- |
| **Revision:** 8 | **Date Created:** 04/27/2007  **Date of Last Revision:** 11/22/2017 | **Last Approval Date:** 11/22/2017 |

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| **Document Author:**  Richard Chui |  |

**Reason for Change:**

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| **Revision:** | **Sec/Para Changed** | **Change Made:** | **Date** |
| 1 | N/A | Import SOP to QSI database | 09/02/2008 |
| 2 | Procedure A 3 | replaced Caviwipe with Sani-cloth | 04/26/2010 |
| 3 | Select Area, Asso'd docs | Added 2200 & 2425 morgue. Added SFOSOP-0080 | 01/12/2011 |
| 4 | Title, Policy, Procedure, References, Approvals | Updated Title, added CAP citations to Policy, updated disinfection times for Sani-Cloth, added CAP reference, updated CLIA Med Dir/Approvals | 11/06/2011 |
| 5 | N/A | Change of Lab Director | 8/6/2013 |
| 6 | Title, Policy | Removed CAP Gen numbers | 4/6/2015 |
| 7 | N/A | Change of Director | 6/29/2016 |
| 8 | Procedure | Removed "Cavicide". Added Spill kit instructions. | 11/22/2017 |

**Approvals:**

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| **Name:** Eric Suba/CA/KAIPERM  **Title:** Chief of Pathology; CLIA Director | Nov 22, 2017 04:14:05 PM PST - Approved by: Eric Suba/CA/KAIPERM |