Clinical Significance

- The accurate assessment of fetal lung maturity can be extremely important in the treatment of complicated pregnancies.
- Respiratory distress syndrome (RDS) is a dire consequence of premature delivery.
- Laboratory analysis of amniotic fluid can predict susceptibility to RDS.
- Lamellar bodies are concentrically layered, phospholipids-filled vesicles which represent the storage form of surfactant.
- They are released by mature fetal respiratory epithelium into the amniotic fluid that is respired *in utero*.

Principle

- Lamellar body diameter $(1 5 \mu m)$ is similar to that of small platelets. Lamellar body counts (LBCs) can be obtained rapidly with the use of the platelet channel of Sysmex pocH-100i analyzer.
- The Sysmex pocH-100i is a quantitative automated hematology analyzer for in vitro diagnostic use.
- The *pocH-100i* counts the platelets (PLT) using electronic resistance detection and hydrodynamically focused technology.
- Sample is aspirated, measured to a predetermined volume, diluted at the specified ratio, then fed into each transducer.
- The transducer chamber has a minute hole called the aperture.
- Inside the detector, the sample nozzle is positioned in front of the aperture and in line with the center.
- After diluted sample is forced from the sample nozzle into the conical chamber, it is surrounded by front sheath reagent and passes through the aperture center.
- On both side of the aperture, there are the electrodes between which flows direct current.
- PLT suspended in the diluted sample pass through the aperture, causing direct current resistance to change between the electrodes.
- As direct current resistance changes, PLT volume is detected as electrical pulses.
- PLT count is calculated by counting the pulses, and a histogram is plotted by determining the pulse heights.

Work place safety

All laboratory employees are expected to maintain a safe working environment and an injury-free workplace. Laboratory employees are responsible for their own safety, the safety of others and adhering to all departmental and medical center safety policies and procedures.

- For standard precautions and safety practices in the laboratory; see LGM 8000, specifically, but not limited to, equipment safety, proper body mechanics, sharps exposure and proper use of personal protective equipment (PPE).
- For Universal Body Substance precautions, see LGM 8005, specifically, but not limited to, exposure to body fluids.
- For proper hand-washing, see LGM 8010, specifically, but not limited to, proper hand-washing.
- For proper infection control, see LGM 8004, specifically, but not limited to, proper use of gloves.
- For proper handling of regular and infectious waste, see LGM 8006, specifically, but not limited to proper disposal of regular and biohazardous waste.
- For proper cleaning of work area, see LGM 8007 Cleaning Work Areas.
- For proper handling of chemicals and reagents, see the Chemical Hygiene Plan.

Clinical Decision Point

- CDP = $65,000 / \mu L$.
- All LBC <65,000 / μ L to be reflexed for confirmatory phospholipid analysis (LS & PG) by TLC (Thin Layer Chromatography).

Specimen Requirement

The following are the specimen requirements for the procedure.

Specimen	Required Volume	Minimum Volume	
 Amniotic fluid Vaginal pool without obvious mucus 	10 mL	3.0 mL	

Unacceptable Specimen:

The following amniotic fluid samples are unacceptable for Lamellar Body testing:

- Bloody (with >5% RBC)
- Mucoid & Gelled specimen
- Contaminated with Meconium
- Vaginal Pool containing obvious mucus

Materials Needed

Equipment	Supplies
 Sysmex pocH-100i Analyzer Vortex Tube Mixer 	 Thermal Paper Transducer brush Philips Screwdriver 12 X 75 mm glass test tubes Disposable transfer pipettes Kimwipes Beige adapter (for sample tubes) Green adapter (for QC vials)

Controls and Calibrator

The following tables list the controls and calibrator used in this procedure, their preparation, storage, and stability.

Product	Preparation	Storage	Expiration
SYSMEX SCS-1000 Calibrator Kit	Ready to Use	Store at 2-8 °C before and after opening.	Unopened calibrators are stable until the expiration date shown on the vial. Once opened, the product is stable for 4 hours if returned to the refrigerator promptly after use.
Eightcheck 3WP X-TRA Control Set	Ready to Use	Store at 2-8 °C before and after opening.	Unopened controls are stable until the expiration date shown on the vial. Once opened, the product is stable for 14 days if returned to the refrigerator promptly after use.
Amniotic Pool Positive Control	Pool previously tested amniotic fluid (AF) samples. Run the pool on pocH-100i. Use negative AF samples from Genetics Lab. + previously tested LBC samples and adjust the concentration to ~ 70,000 /μL. Aliquot 2 mL each into a screw- capped plastic tube.	Store Refrigerate d At 2-10 C	Stable 6 months
Amniotic Pool Negative Control	Pool negative AF samples from Genetics Lab to obtain a concentration of < ~ 3,000 /μL. Aliquot 2 mL each into a screw- capped plastic tube.	<u>Store</u> <u>frozen</u>	Stable 6 months

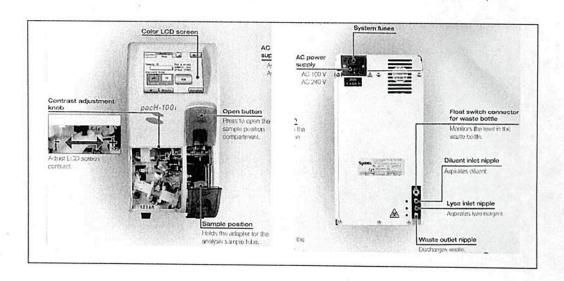
Reagents

The following tables list the reagents used in this method, their preparation, storage, and stability.

Reagent	Preparation	Storage	Expiration
pocH-pack D (Diluent)	Ready to Use	Room Temp.	 Unopened packs are stable for 1 year. Once opened, reagent is stable for 60 days.
pocH-pack L (Lysing Reagent)	Ready to Use	Room Temp.	 Unopened packs are stable for 1 year. Once opened, reagent is stable for 90 days.
Bleach (5%)	Ready to Use	Room Temp.	• Use fresh daily.

System Overview

Parts of Sysmex pocH-100i Analyzer



Startup Procedures

Step	Action
1	• Verify the following before powering on:
	• Check that the power cable is connected.
	A C III
2	• Check that there is sufficient printer paper, replace if needed.
	1. Open the paper holder by pushing the knob.
	2. Lift the printer cover.
	P P P P P P P P P P P P P P P P P P P
	Correct Incorrect
3	Check waste fluid in waste container, discard if needed.
4	Check and replace reagents if needed.

Power On/ Self Check

Action		
 right side of the instru Three automatic rinse of background check. Should any values be of two extra background 	out of the acceptable limit, a maximum and checks will automatically be	
PLT count is 10 x 10 ³ /μL or less	• print & record the result on the daily PM chart. • proceed to running controls	
PLT count is higher than 10 x 10 ³ /μL	 perform another cycle of Auto Rinse. if still outside the limits, refer to Section 6 page 71 of the Instruction Manual for more troubleshooting guidance once background reading is within limits, proceed to running controls. 	
Blank check), • press "Menu" • press "Str. Data • then select the f • results can first → arrow. • then press "Prin	ile "Blank check" be viewed on screen by using the ← or	
	right side of the instrue • Three automatic rinse of background check. • Should any values be of two extra background performed by instrume • Review results of back If PLT count is 10 x 10³/μL or less PLT count is higher than 10 x 10³/μL • To print results of back Blank check), • press "Menu" • press "Str. Data then select the fearesults can first → arrow. • then press "Print I arrow. • then press "Print I arrow.	

Performing Quality Control

Step	Action
1	• Run 1 set of EIGHTCHECK Control (Low, Normal, and High) on each day of instrument use.
	Remove control vials from the refrigerator at least 15 minutes before use.
	• Roll 10 times, Turn upside down 10 times, then Roll again for
	another 10 times.
	• Examine vial bottom. If there is red cell pellet still formed on the
	bottom, then repeat the whole mixing procedure!
	Include Amniotic Pool Positive & Pool Negative controls with every
	run of patient sample.
	• Negative Pool control is stored frozen. Thaw at <u>least 30 minutes</u> before use to bring to room temperature.
	Positive pool control is stored in the refrigerator, bring to room
	temperature before use.
	Vortex well before running.
i.	4860
4	• To run QC → from the main screen
	 Verify that "WB" mode is selected, if not, press "WB"
	1 2 3 3
	Main 1 1 1 1 1 1 1 1 1
	Selection for page (CM).
	4 (4.00.000)
	5. (Finalt) (Austria) (6. 63 (43/6)(2003)
	Press "QC". Select analyzing File. Open sample position.
	A countries
	Press "QC"
	Select the QC file (filename is the
	lot # for Low, Normal, or High
	control). Onen the sample door (also known as sample position) and
	 Open the sample door (also known as sample position) and insert the green adapter
	moore the green adapter
	The state of the s

Performing
Quality Control

Step	Action		
5		file (1) Pice control Nord in take observe and	
		wuit" button will appear on screen and to go back to main screen.	
	If	Then	
	PLT count is within acceptable performance for Low, Normal, and High controls	Proceed to running patient samples.	
	Control results are not acceptable	 Verify that the right QC file was selected. Check that there is no cell pellet on the bottom, mix the vial, then re-run control. If still outside the limits, refer to Section 6 page 71 of the Instruction Manual for more troubleshooting guidance. Document all problems and corrective action(s) taken on the QC/QI book. STOP! Do not proceed to patient testing until QC is 	

Maintaining Patient and Sample Identity

Step	Action
1	 Check that all patient's information on the worksheet or requisition match those on sample tube(s) (Name, Accession #, MR #, gender, etc).
2	Always use barcode labels when labeling secondary tubes.
3	Use barcode scanner to scan the Accession # for sample ID when running pocH 100i.

Performing Sample Analysis

tep	Action		
1	 To run pool or patient sam Verify that "WB" n Press "Sample ID" the alphanumeric keep 	▲	
	If	Then	
	running Pool controls	manually type in the ID (Pool).press enter.	
	running patient	• scan the barcode ID.	
	sample	• press enter.	
à	or by	the ID manually bar code reader press "Ent.". Dutton (WB) is selected.	
2	Vortex the pool controls a well mixed sample.	and patient samples for 5-10 sec fo	
3	Transfer at least 2 mL to a	a 12 X 75 mm test tube.	
4		ne patient sample, a Pool Negativ tient to verify no carryover exhibit	

	Step		Action	
Performing	5	The following sampling sequence must be followed exactly		
Sample Analysis, continued	4	 Pool Positive Control (PPC). Pool Negative Control (PNC). 		
		3. Sample 1 (patient)	mor (rrve).	
	41	4. Pool Negative Con	ntrol (PNC)	
		5. Sample 2 (patient)	The second second	
		6. Pool Negative Cor	ntrol (PNC)	
		7. Sample 3, (patient)	, etc.	
			ntive sampling sequence is to ensure no from previous patient or control sampl	
	6	If	Then	
		PNC result is "0"	Proceed to running sample.	
		PNC is giving any	• STOP!	
		value	• Do not run sample, perform a "Clean Transducer" cycle.	
			• See page 12 for stepwise procedure on How to clean the transducer .	
	7		(also known as sample position)	
		• Insert the beige sample	adapter, load sample and close door.	
oush the top		4	5	
part to open the door	7.			
		Insert the correct adapter.	Insert sample tube and close the door.	
	8	Press "Run"		

Document No. LHM299.11

QSE: Procedure

How to Perform Clean Transducer

Step	Action 1 • From Main screen at the "Ready" status, 1. press "Menu". 2. press "Maint." 3. select "Clean Transducer"	
1		
2	 Fill 12 x 75 mm test tube with concentrated bleach. Push the sample door to open the sample position. Place the tube with bleach on the beige adapter. Press "Execute" 	

How to fix Error Messages

Step	Action		
1	 When an error occurs, Alarm will sound Alarm button will flash. 		
2	 Error message(s) will appear on screen. In case of multiple errors, they will be displayed on screen ranked by importance. Press the "OK" button on error message screen to stop the alarm and to close error dialog. 	Sample 10 Set a maked sharely and press (RUN). Realized Rose "Printer Error" occurred.	
3	 Press the HELP button (also called alarm button). This will show on screen the kind of error that had occurred. Press "Detail" to see instructions on how to fix the error. 	Sample ID Sample ID Set a mixed state and press (RIM). Analysis Hole ME PO RIN 20 Result Disserve Statdown	

How to fix Error Messages, continued

St	ер	Action		
		• Refer to Page 67 (Chapter 6) of the Instructions Manual for more troubleshooting guidance.		
	5	• If unable to solve the problem, call 1(866)-8SYSMEX for technical assistance.		
		• <i>NOTE</i> : In case of power failure during operation, turn the main power switch OFF.		

How to Replace Reagents

Step	Action		
1	 Changing reagent from Main screen at the "Ready" status, press "Menu". 		
	(1 0)		
	• pr	ess "Chg. Reag"	
	Section Sect	The state of the s	Continue S. Dari
2	• The reagent screen will appear, (for example):		
		Lot. No.	Expiration
	pocH-pack D	A8073	07/14/2010
	pocH-pack L	A8014	12/18/2009
		agent to replace	7 [
	pocH-pack D	pocH-pack L	Reag. Log
3	• Press the appropriate button for the reagent pack to replace.		

How to Replace Reagents, continued

Step	Action		
4	The screen below opens up.		
	COR. Request mock-mark D town Serial Prock-mark D town Serial Brooks Barcole.		
	WHEN THE STREET STREET		
	• Reagent lot & exp. date can be entered manually or by scanning the reagent barcode.		
	• Scan the reagent barcode, the lot # & exp. date will automatically register on screen.		
5	Press "OK" to confirm the Lot # & exp. date. Occ. (2 ANAPOSCILIPSOSILARS) FORWARD CONFIRM TO SHAPE		
6	Press "OK". Insert the container spout kit into the new bottle.		
	• Tighten the cap then press "Execute".		
	S TO THE FORM THE STATE OF THE		
7	If the reagent runs out in the <u>middle of run</u> , an error message will appear on screen prompting the operator to replace the depleted reagent.		
	reagent. • Press "Execute" • Follow steps 4-6 above. Traction production for the first of t		
	Press "Execute".		

Performing Daily Shutdown Procedure

Step	Action		
1	 It is very important to <u>perform shutdown</u> procedure at the end of the day to remove deposits in the tubing. Deposits in the instrument tubing can cause incorrect results or it may damage the instrument. 		
2	Run concentrated bleach as sample to clean the transducer of residual protein.		
3	Perform shutdown cycle, from Main screen at the "Ready" status, press "Shutdown". press "Execute"		
4	• After the shutdown cycle (2 minutes), turn OFF the instrument.		

Reporting/ Results Interpretation

Step	Action	
1	Lamellar Body Cou	ints are read as PLT on pocH-100i
	If	Then
	PLT count is $\geq 65 \times 10^3 / \mu L$	• Release in LMS, it will be reported as MATURE .
	PLT count is $< 65 \times 10^3 / \mu L$	• Release in LMS, it will reflex to L/S & PG confirmation by TLC.

How to Calibrate

Step	Action		
1	• Calibration interval on <i>pocH-100i</i> is every 6 months.		
2	 The calibration is always performed in the "WB" mode. The type of calibration performed is "Calibrator calibration for whole blood (WB) mode" page 65, instructions manual. 		
3	 A precision check is performed prior to calibration. When performing this calibration, the first analysis data is not included in the calculation. It is overwritten by the second analysis data. 		
4	 If an error occurs during precision check analysis or calibrator analysis, an alarm sounds and an error dialog appears The results of the analysis where the error occurred are displayed in reverse video, as "". 		
5	 Press "OK" to close the error dialog, then press "Run" again. Take out the calibrator from refrigerator at least 30 minutes before use to warm the vial to room temperature. Mix each vial by gentle end-to-end inversion until the cell button in the bottom of the vial is completely suspended. Allow the vial to rest on flat surface 15 seconds prior to analysis to allow for the dispersion of micro-bubbles. 		
6	 From Main screen, press "Menu" press "Calib" select "Calibrator (WB)" 		
7	• Insert the green adapter into the sample position then load the calibrator vial.		
8	 Press "Run", let the analysis finish; then repeat this step 10 more times. This step should be performed a total of 11 times for precision check. 		
9	 After the completion of precision check, press "Next" press "Quit" 		

How to Calibrate, continued

Step	• The calibration screen opens up. • Manually enter the target values listed on calibrator insert and verify that they are correct • press "Enter" • press "Next" • press "OK"		
10			
11	 Press "Run", let the analysis finish; then repeat this step 5 more times. This step should be performed a total of 6 times to obtain required calibration data. 		
12	 After analysis, press "Next". The calibration results show on screen. 		
13	• Press "Quit" to update calibration value and calibration history.		
14	Document calibration on calibration log, CLS initials & date.		

Reviewing Calibration Results

Step	Action		
1	Image below shows an example result of calibration run		
	Top I Cab (WD) 1		
	WBC RBC HGR HCT PLT Range V. 7 20 03 13 20 Markhangn 7 19 0.4 16 35 Meant V. 69 482 15 1 40.9 21.2 Octurs 5.26 0.08 5.09 5.00 5.01 SERV LMT 14 00 5.00 5.00 5.00 15.00 Carront 99.8 102.4 100.8 52.9 97.4 New 102.4 103.4 100.8 52.9 98.3		
	Press "Quit" to update calibration value and calibration history.		

Reviewing Calibration Results, continued

	Range V.	The highest value minus the lowest value within five consecutive analysis.
	MaxRange	The maximum Range Value allowed
	Mean V.	Mean value of five consecutive analysis results.
	Delta %	Assay Target – Mean Value X 100 Mean Value (this is automatically calculated and displayed).
	ACPT LMT	Maximum Upper Limit of Delta Percent (%).
<i>20</i>	SERV LMT	• If the Delta Percent exceeds this limit, technical assistance may be necessary and instrument system needs service.
	Current	 Current calibration value which was obtained in the previous calibration procedure.
	New	New calibration value calculated from the calibrator analysis.

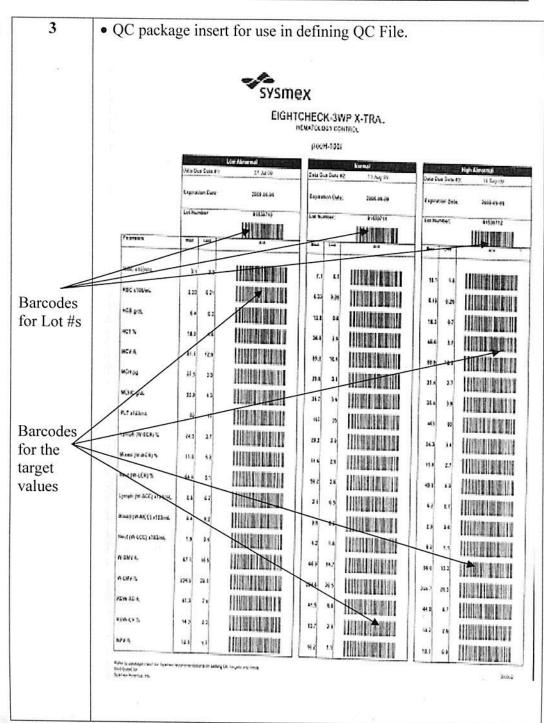
Reviewing Calibration Results, continued

Step	Action		
3	Table below shows how the calibration data is used by the system		
1950	If	Then	
	Delta % is within ACPT LMT	• Calibration factor will not be changed, i.e. instrument will continue using the "Current" calibration value.	
	Delta % <= ACPT LMT	• The "New" calibration value will become the "Current" calibration value.	
	Range V. > MaxRange	• The "New" calibration value will become the "Current" calibration value.	
	Delta % > SERV LMT	 Calibration will not be allowed. Instrument needs service. Call 1(866)-8SYSMEX for technical assistance. 	
4	"NEW" calibration value (automatically calculated)	e = (Target/Mean V.) x Current d & stored by instrument)	

How to Setup QC File

Step	Action			
1	 To setup QC file, from Main screen, press "Menu". press "Chg. Reag" press "Setup" 			
	Service Servic			
2	 Scan the barcode label for the Lot #. (the first set of barcodes on top of the QC package insert-refer to step 3). Expiration date has to be manually entered. 			

How to Setup QC File, continued



How to Setup QC File, continued

Step	Action		
4	 Press → to go to the next 5 QC file setting screens. 		
	• Scan all 19 barcode labels for each level (Low, Normal, and High). 5		
5	Once all the parameter have been entered/scanned, press "Save press "Ok Once all the parameter have been entered/scanned, press "Ok Once all the parameter have been entered/scanned, press "Ok Once all the parameter have been entered/scanned, press "Ok Once all the parameter have been entered/scanned, once all the parameter have been entered/scanned,		
	The completion The		

Quality Control Evaluation

Step	Action
1	 When calculating ranges for new lot of whole blood controls or new batch of amniotic pool positive control, use the same historical SD & CV as the Regional Reference Laboratory.

Notes and Limitations

Step	Action		
1	 For sub-optimal samples that are unacceptable for Lamellar Body Count obtain L/S& PG order and send specimen to the Regional reference laboratory for L/S & PG detection by TLC. 		
2	 If the AF hematocrit (HCT) exceeds 1%, reflex the specimen for L/S & PG detection by TLC. 		

Alternate Method

Step	Action
1	• In the event of instrument malfunction and the problem could no be resolved, samples should be sent straight for L/S & PG
	Detection by TLC.

Documents

Non-Controlled The following non-controlled documents support this procedure.

Reference	
SYSMEX pocH-100i Analyzer, Instructions 2005-2006	Non-controlled
SYSMEX package inserts (reagents, calibrators, controls)	Non-controlled
Preventive Maintenance for pocH-100i Analyzer	Non-controlled
Lamellar Body Count Inventory	Non-controlled
Calibration Log	Non-controlled
Operator Training Checklist	Non-controlled

Controlled documents

	Title	
LGM8000	Standard precautions and safety practices in the laboratory	
LGM8005	Universal Body Substance precautions	
LGM8010	Proper hand-washing	
LGM8006	Infection Control	
LGM8007	Cleaning Work Areas	
LGM8012	Proper storage and disposal of chemical hazardous waste	

Author

M Toprakci

Reviewed and approved by:

DATE
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12/10/2012
TANKS OF THE PARTY

Continued on next page

Kaiser Permanente Medical Care Program California Division-South SCPMG Laboratory System
Process Control
Procedure

Procedure for Lamellar cody Counts in Amniotic Fluid on SYSMEX pocH-100i Analyzer, Continued

Document History Page

Effective Date:

Change	Changes Made to	Signature	Med. Dir.	Lab	Date change
type: New,	SOP – describe	responsible	Reviewed/	Manager	implemented
Major,		person/date	Date	reviewed/	
Minor etc.		11349	/	date	
New		10/29/12	1	Milologi	
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