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| PRINCIPLE | The purpose of the blood smear is to demonstrate all cellular elements in the same proportions in which they exist in the blood. With the glass slide method large, heavy cells will be concentrated at the distal end or featheredge of the smear. Coverslip smears are more accurate since the distribution of leukocytes is dependent on capillary action rather than mechanical pushing. Practicality demands the routine smears be made by the glass slide technique. |
| SAFETY | All specimens, reagents and controls should be handled as though capable of transmitting infectious diseases. Wear appropriate personal protective equipment when running patient samples or performing scheduled maintenance. Refer to: Policy and Procedures Safety Manual Infection Control and Procedures 11-085-01. |
| MATERIALS AND **REAGENTS** | Microscope  Sterile Transfer Pipet  Slide Stainer  Glass Slides  Diff Safe Blood Dispenser |
| SPECIMEN | The blood for films may be obtained from capillary blood (finger puncture) or from venous blood anticoagulated with EDTA. If any clotting has occurred in venous blood samples, severe alterations in leukocyte enumeration and differential counting will have occurred and platelet estimations will be valueless |

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| PROCEDURE | Glass Slide Technique:   |  |  | | --- | --- | | Step | Action | | 1. | Make a capillary puncture. Wipe away the first drop of blood and transfer a drop of blood 3-4 mm in diameter to a slide about a half-inch from one end. (Or place a drop of blood from an EDTA tube on a slide with a capillary tube or diffsafe technique). | | 2. | Lay slide on a flat surface. Place an end of a second slide, balanced on the fingertips at an angle no greater than 30 degrees in front of the drop of blood. | | 3. | Pull the spreader slide into the drop of blood. When the blood has spread along 1/2 of the width, push the spreader slide forward with a steady, even motion. The weight of the slide is the only pressure applied. | | 4. | Allow the smear to air dry. Stain with Wright’s stain. | | 5. | A good film should meet the following criteria:   1. The entire smear should cover no more than 1/2 of the area of the slide. 2. No portion of the smear should extend to the edges of the slide. 3. There should be no ridges, lines, or holes. 4. Leukocytes should be evenly distributed. 5. Red cells should touch but not overlap on about 1/3 of the blood film. | | 6. | The thickness of the film is dependent on the size of the drop, speed of pushing the spreader slide, and the angle of the pusher slide. | | 7. | Errors to avoid are:   1. Too large or too small a drop. 2. Delay between transferring the drop of blood to the slide and making the smear. 3. Dirty smear slide. 4. A chipped or dirty spreader slide. 5. Use of a spreader slide with cut off corners does not prevent margination of leukocytes. If the smear is made quickly and properly, margination can be avoided. | | |
| LABELING | | Outpatient Slides:   |  |  | | --- | --- | | Step | Action | | 1. | CLS prepares the smear and labels the slide with patient’s first and last name and date or accession number. | | |
|  | | PATHOLOGY SLIDES:   |  |  | | --- | --- | | Step | Action | | 1. | CLS prepares the smear and labels the slide with patient’s first and last name, medical record and date. | | |

Document History Page

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| Change type: New, Major, Minor etc. | Changes Made to SOP – describe | Name of responsible person/date | Med. Dir. Reviewed/ Date | Director of Lab Ops. reviewed/ date | Date change Implemented |
| MINOR | 1. Regional Template Update  2. Revised index no.  3. Added Safety  4. Added Materials and Reagents | Yvette Lingat  3/20/2020 |  | Mary Lou  Beaumont | 4/28/2020 |
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