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| Blood Culture - Bactec™ FX | | | | | | | | | | | |
| **Purpose** | Blood cultures are essential in the diagnosis and treatment of the etiologic agents of sepsis. The bacterial detection of microorganisms in a patient’s blood has diagnostic and prognostic importance. Sepsis constitutes one of the most serious infectious diseases and, therefore, the expeditious detection and identification of blood-borne bacterial pathogens is an important function of the diagnostic microbiology laboratory.  The Bactec™ FX is designed for the rapid detection of microorganisms in clinical specimens. The sample to be tested is inoculated into the vial, which is entered into the Bactec™ instrument for incubation and periodic reading.  When microorganisms are present in culture vials, they metabolize nutrients in the culture medium, releasing carbon dioxide into the medium. A dye in the sensor at the bottom of the vial reacts with CO2. This modulates the amount of light that is absorbed by a fluorescent material in the sensor. A photo detector at each station measures the level of fluorescence, which corresponds to the amount of CO2 released by organisms. Then the measurement is interpreted by the system according to pre-programmed positivist parameters.  At system startup, the onboard computer performs self-diagnostics and downloads operating instructions to the drawer rows. Then the instrument(s) automatically begin testing. Light Emitting Diodes (LEDs) behind the vials illuminate the rows, activating the vials’ fluorescent sensors. After a warm-up period, the instrument’s photo detectors then take the readings. A test cycle of all rows is completed every ten minutes. Positive cultures are immediately flagged by an indicator light on the front of the instrument, an audible alarm, and are displayed on the LCD display. | | | | | | | | | | |
| **Test Code** | BC, BRCL | | | | | | | | | | |
| **Materials** | Reagents | | | Supplies | Equipment | | | | | Media | |
|  | • Gram stain reagents  • Vitek™ GNI, GPI, GNS, GPS, YST, NH cards | | | • 70% isopropyl alcohol wipes  • BD™ Blood Transfer Device  • Venting needle  • 1 cc syringe  • Snap cap tubes  • Glass slides  • Sterile transfer pipettes | • BACTEC™ FX Microorganisms, if present in the blood samples, metabolize nutrients in the BACTECTM culture vial and release CO2 into the medium or utilize the oxygen in the medium. The instrument monitors the fluorescence of the vial sensor, which increases as CO2 is produced or oxygen is utilized. Analysis of the rate and amount of CO2 produced or O2 utilized enables the instrument to determine if the vial is positive; i.e., the presumptive presence of viable organisms  • Computer and Peripherals  The system computer stores all the system software, including the application software which controls instrument operations and the user interface, which enables the user to enter patient information, view results, print reports, identify errors, etc  • Barcode Scanner  The barcode scanner is located at the front of each drawer.  • CO2 incubator 35°C  • Anaerobic chamber 35°C  • Ambient air incubator 35°C  • Incinerator  • Inoculating loop  • Microscope | | | | | • Bactec™ Peds Plus/F Culture Vial1 (pink bottle): Optimum blood volume for each vial is 1 to 3 mL; 0.5 to 5 mL of blood is acceptable.   1. Each vial contains:  * 40 mL Enriched Soybean-Casein Digest Broth * 0.02% SPS * Resins * CO2 * O2 * Sensor for the detection of fluorescence  1. Store at 2° to 25° C 2. Bactec™PlusAnaerobic/F Culture Vial2 (orange bottle): Optimum blood volume for each vial is 8 to 10 mL; 1 to 10 mL of blood is acceptable. 3. Each vial contains:  * 25 mL Enriched Soybean-Casein Digest Broth * 0.05% SPS * CO2 and Nitrogen Gas * Sensor for the detection of fluorescence  1. Store at 2° to 25° C.  * Chocolate Agar (CHOC) * Sheep Blood Agar (BAP) * CDC Anaerobe Agar (ASB2) * CNA Agar (CNA) * MacConkey Agar (MAC) * Sabouraud Dextrose Agar, Emmons (SAB) * Candida Chromagar (CCAN) | |
| Sample |  | | | | | | | | | | **Related document** |
|  | 1. Volume    * The volume of blood cultured is critical because the number of organisms per mL of blood in most cases of bacteremia is low, especially if the patient is on antimicrobial therapy. In infants and children, the number of organisms per mL of blood during bacteremia is higher than adults, so less blood is required for culture.4 For pediatric patients; 2 to 6 mL of blood is drawn per blood culture set. Inject 1 to 3 mL into each vial using the following guidelines:  |  |  | | --- | --- | | Weight in Kg (lb) | Volume per bottle (mL) | | <1.5 Kg (<3.3 lb) | 1.0 | | 1.5-3.9 Kg (3.4🡪8.6 lb) | 1.0 | | 4.0-13.9 Kg (8.7🡪31 lb) | 2.0 | | >14 Kg (over 31 lb) | 3.0 |  * If only the minimum volume of blood can be drawn, inoculate the Bactec™ Peds Plus/F vial only.  1. Sampling Time    * Draw 2 to 3 sets of blood cultures per febrile episode at least 60 minutes apart. Do not draw more than 3 sets in a 24-hr period. This provides maximum recovery of microorganisms in patients with intermittent bacteremia, and documentation of persistent bacteremia in patients with intravascular infections (e.g.endocarditis, intravenous catheter site infections). 2. Special instructions    * Inoculated vials should be transported as quickly as possible to the laboratory.    * Bottles should not be refrigerated or frozen.    * **DO NOT USE** culture bottles past their expiration date.    * **DO NOT USE** culture bottles that exhibit any cracks or defects; discard the vial in the appropriate manner.    * **DO NOT USE** culture bottles that have had their caps removed prematurely. | | | | | | | | | | [Lab Test Directory – Blood Culture](http://www.childrensmn.org/Manuals/Lab/MicroBioViral/033016.asp) |
| Special Safety Precautions | Pathogenic microorganisms, including Hepatitis B Virus and Human Immunodeficiency Virus, may be present in specimens. “Standard Precautions” and institutional guidelines should be followed in handling all items contaminated with blood or other body fluids.   * Wear gloves while handling inoculated vials. * Perform all blood culture processing in a biological safety cabinet. * Properly dispose of all contaminated materials. Place syringes, needles and other sharp contaminated materials in a puncture proof container. * NEVER ATTEMPT TO RECAP A NEEDLE.   Microbiologists/virologists are subject to occupational risks associated with specimen handling. Refer to the safety policies**:**   1. [*Biohazard Containment*](file:///\\kidsnet.childrenshc.org\chcdfs\dept\LAB\Micro%20Procedure%20Manuals\MC%20200%20%20%20%20Safety\MC%20201%20%20%20Biohazard%20Containment.doc) 2. [*Safety in the Microbiology/Virology Laboratory*](file:///\\kidsnet.childrenshc.org\chcdfs\dept\LAB\Micro%20Procedure%20Manuals\MC%20200%20%20%20%20Safety\MC%20202%20%20%20Safety%20in%20the%20Microbiology%20Lab%20Policy.doc)  * [*Biohazardous Spills*](file:///\\kidsnet.childrenshc.org\chcdfs\dept\LAB\Micro%20Procedure%20Manuals\MC%20200%20%20%20%20Safety\MC%20204%20%20%20Biohazardous%20spills.doc) | | | | | | | | | | |
| **Quality Control** | **Media Quality Control**  Commercially prepared blood culture media do not require additional in-laboratory QC per CLIA and CLSI M22-A3.  Each case of media has a Quality Control certificate from BD indicating the organisms tested and the acceptability of those tests. An example from each media type is kept on file.  **Bactec FX Instrument Maintenance**  The following procedures should be performed daily:   1. Check the paper supply to the printer. If the paper supply is low or exhausted, replace the paper as explained in the operating manual furnished separately. 2. Tap the “maintenance” tab. The Test display appears. 3. Open drawer A. Then tap the “red” button to illuminate the red station indicators. Make a note of any station that does not illuminate red. 4. Next tap the “green” button to illuminate the green station indicators. Make a note of any station that does not illuminate green. 5. Repeat Steps 3 - 4 for each of the drawers in the system. 6. Close the drawer. 7. Tap the “alarm” button to verify that the audible alarm is functioning. 8. Finally, tap the “status” button to illuminate the system status indicators on the mullions. Both sides of all the indicators (amber, red, and green) should illuminate. If any indicator does not light, contact your local BD representative for service. 9. Check the temperature on the temperature vial(s) in each drawer. 10. Information should be recorded on the Maintenance QC Chart, which is located in the Bactec Maintenance book. Daily QC reports that print automatically should be checked and filed in the Bactec Maintenance book as well.   **Daily Backup:**  The automatic Epicenter Backup is programmed to happen at 3:00am. The backup DVD should be changed each morning during daily maintenance. The DVD will be automatically ejected after backup is complete each morning.  **Monthly Maintenance:**  Change both sets of filters on each Bactec instrument. Rinse filters thoroughly with water and allow to dry completely. | | | | | | | | | | |
| **Procedure (computer)** | **Ordering in the Sunquest Computer:**   1. Test is ordered as non-restricted, so labels will print in Microbiology upon order in Cerner. 2. Receive cultures orders in either Order entry in Mysis, or CVIS in Sunquest. 3. Use VP (if lab drawn) or RN (if nurse drawn) for Phleb Workload. 4. Use one of the following for a site code:  * ARL – Art line * ARTP – Arterial puncture * BLDN – Blood collect site not specified * BLUL – Blue lumen * BROV - Broviac * CENL – Central venous line * CORD – Cord blood * CVP – CVP line * FEM – Femoral * HICK – Hickman * IVS – IV start * LD – Line draw * MEDL – Medcomp line * PEBL – Peripheral blood * PERC – Perc line * PICL – PIC line * PORT – Port-a-cath (PAC) * REDL – Red lumen * UART – Umbilical arterial catheter (UAC) * UVC – UVC line * WHL – White lumen * For ECMO circuit—use LD-; ECMO circuit  1. Place the Sunquest generated barcode label on the Bactec™ vial without covering the vial’s barcode label. 2. Disinfect the top of each vial with 70% isopropyl alcohol. | | | | | | | | | | |
| **Procedure** | * + - 1. **Entering Data And Loading Instrument**   To enter vials in the instrument, select a drawer where there are available stations. (The number of available stations is shown below the “vial entry” icon on the Status display.) Then follow one of the two methods described below.   * + - 1. Method 1 (Vial Activated)  1. Select a drawer that has available stations, and open that drawer 2. The barcode scanner turns on 3. Scan a vial sequence barcode label 4. The Vial Entry display appears and the Sequence, Media, and default Protocol are automatically entered 5. If you did not scan the Accession, scan or enter it now 6. To change the protocol tap the “modify” button, then tap the up arrow to increase or down arrow to decrease the protocol length. 7. Place the vial into an available station (solid green indicator)    * + 1. Method 2 (Icon Activated) 8. Select a drawer that has available stations, and open that drawer 9. Tap the “vial entry” button on the Status display 10. The Vial Entry display appears and the barcode scanner turns on 11. Scan the vial sequence barcode label 12. The Sequence, Media, and default Protocol are automatically entered 13. If you did not scan the Accession, scan or enter it now 14. To change the protocol tap the “modify” button, then tap the up arrow to increase or down arrow to decrease the protocol length 15. Place the vial into an available station (solid green indicator) 16. When a vial is placed into the last available station in a drawer, the Activity Complete tone sounds (3 beeps). 17. To continue entering vials, select another drawer with available stations.     * + 1. **Inserting Vials in the Instrument**         2. Before inserting vials into the stations, visually inspect all vials for positives. Evidence of microbial growth includes hemolytic, turbidity, and excess gas pressure (causing the vial septum to bulge outward). All such vials should be treated as positives; they should be stained and subcultured.         3. After all vials have been inspected and inserted in stations, close the drawer.         4. A vial presence sensor immediately senses the insertion of a vial in a station and the instrument updates the station LED indication and the status shown on the LCD.         5. Once vials are placed in their stations, you should avoid moving them to other stations unnecessarily.         6. Avoid opening the drawer unnecessarily. Drawers should not remain open longer than 10 minutes. Continuous loud alarm will sound if drawer is open more than 10 minutes.         7. Make sure all vials are fully inserted in the stations before closing the drawer.   Vials that are not read for 40 minutes (because of an open drawer or being unseated) need to be subbed and an AO performed. If AO stain is positive, perform gram stain.   * + - 1. **Vials Delayed in Transport—**(add **DELA** to the **SDES** when receiving these cultures).   Vials that are delayed in transportation to lab 8 hours or more need to be subbed to CHOC, SB, and ASB2 plates, and a gram stain and AO performed before being placed into Bactec.  Subculture the bottle(s) according to the positive bottle BC protocol.  Read the preliminary gram stain. Leave results for day shift Micro with a label indicating the status.  If the gram stain is negative, put the bottle(s) in the Bactec according to the processing new vials protocol.  Bottles can be held up to 48 hours at room temperature and up to 24 hours in a 35-degree incubator, and still be placed into the Bactec for reading.   * + - 1. **Anonymous Vial Entry**       2. Vials can be placed into available (GREEN indicator) stations without being scanned into the instrument. Vials that are not scanned into the instrument are called “anonymous” vials. The instrument recognizes anonymous vials when they are placed in stations, but are assigned an “unknown” medium type and default protocol of 5 days. Anonymous vials are evaluated with general positivity criteria. They cannot use the specific positivity criteria tied to the characteristics of the medium since the instrument does not know the medium type.       3. These anonymous vials need to be identified in the system using the ID(entify). Do not perform Negative Vial Removal until all Anonymous vials have been resolved. You could lose data if you accidentally remove an Anonymous vial.       4. To identify anonymous vials:  1. Open drawer and remove vial from flashing yellow station or open drawer and tap ? to activate ID Anonymous workflow. 2. Scan the sequence and accession for the anonymous vial. The patient information is filled on the workflow display and the station the vial was pulled from will be flashing green. 3. Return the vial to the flashing green station.    * + 1. NOTE: Once an anonymous vial has been placed in the instrument, do not remove the vial and reenter it without identifying it (ID Anonymous activity). All test readings are discarded if you remove the vial without identifying it.   **Positive / Negative/ Ongoing Vials**   1. **Notification of positive and negative vials**    1. The system notifies you of new positive cultures in several ways       1. Positive Vial audible alarm sounds       2. Station Indicators: FLASHING RED or FLASHING AMBER / RED (alternating) -Anonymous Positive       3. Message box appears on Epicenter screen.       4. Positive vial system indicator for that drawer illuminates       5. On the Status display, the “positives” icon is active (color is red, not grayed out) and the number of positive vials in the drawer is shown          1. Out-of-Protocol Negatives are indicated by the following             1. Negative vial system indicator for that drawer illuminates             2. On the Status display, the “negatives” icon is active and the number of negative vials in the drawer is shown             3. Station indicators: FLASHING GREEN   In Protocol Negatives (ongoing) are indicated by LED with no light lit up.  **Removing positive vials**   1. Print “**Current positive** report”. At the FX screen, touch the **Reports** tab. Touch the drop-down menu and select **Current positives.** Touch the **Print** button at the bottom of the screen. 2. Select a drawer that has positive stations, and open the drawer by pulling it out.   The barcode scanner turns on.  All positive, final negative, available, and anonymous (all variations) are indicated by the appropriate lit or flashing station indicators.  Tap the “remove positives” button on the Status display, OR  Remove a vial from a FLASHING RED (positive) or FLASHING AMBER / FLASHING RED (anonymous-positive) station  The Positive Removal display appears. Scan vial sequence. (If an anonymous positive vial was removed, the ID Anonymous display appears. Scan the sequence and accession for the anonymous positive vial and tap the “Save” button. Then tap the “Exit” button to return to the Positive Removal display.   * + - * 1. **Negative Vial Removal**   Negative bottles with be removed at the beginning of each shift.  Open drawer.  Remove negative vial from Flashing Green station (Vial activated workflow).  OR:  Tap the “remove negatives” button to activate Negative Removal Workflow (Icon Activated workflow).  “Remove Negative” Workflow display is activated.  Only negative vial station LEDs are illuminated flashing green and the barcode reader is not turned on.  Continue removing negative vials until all vials with flashing green LEDs are removed.  Dispose of bottles in biohazard waste containers.  Retain bottles labeled with white tape “flag” labeled TSUB. Follow False Positive Bottle instructions.  If a completed Out-of-Protocol vial is accidentally left in the instrument, it will remain negative and can be removed at a later time.  Triple beep (workflow complete) will sound.  **Processing an Instrument – Positive Vial**  Remove the vial from the instrument and place in a biological safety cabinet.  Reprint the specimen label to use on the subculture plates.  If it is necessary to release pressure in the vial, place a 70% isopropyl alcohol wipe over the septum and insert a venting needle through the alcohol wipe and septum. Remove the needle after the pressure is released. Place the venting needle into a sharps container.  Invert the vial to mix the contents.  Disinfect the septum of the vial with a 70% isopropyl alcohol wipe. Allow to dry.  Attach a 1 ml syringe to a blood transfer device.  Push the blood transfer device into the septum of the vial, invert and withdraw 1ml.  Remove the blood transfer device with the syringe from the vial.  Remove the syringe from the blood transfer device and discard the blood transfer device into a sharps container.  Using the contents from the syringe, inoculate a CHOC, SB and ASB2 for each bottle. Label the plates with the current date, the current time, mark them “A” for the aerobic bottle and “N” for the anaerobic bottle (use the barcode labels).  Positive vials suspicious of Brucella should be sealed with tape to prevent exposure upon opening.  Make a gram stain slide.  Expel the remaining sample into a sterile and labeled snap cap tube.  After the slide is dry and heat fixed, perform the gram stain procedure as soon as possible.  Read and report the gram stain results. On eves/nights, a 2nd tech must review slide before reporting. (See Reporting section).  Perform an AO if gram stain result is negative.  Results of the gram stain may require additional plates to be inoculated.  Inoculate the following agar plates according to the gram stain results:   1. Gram-negative rod: CNA and MAC from the positive bottle and the related bottle. 2. Yeast: CCAN and SAB from the positive bottle and related bottle.    1. Perform identification and susceptibility (AST) of organism(s) grown on solid media according to laboratory protocol.    2. Always consult caregiver regarding AST, if not performed, except for cultures with multiple isolates drawn from IV start (IVS) drawn in E.D.    3. Positive bottles are saved for one month in case of additional testing.    4. Returning ‘False’ (smear negative) Positive Vials       * + Place tape on the neck of the vial to mark it for a terminal subculture and AO stain.         + Go to Vial Entry, scan sequence, and place vial in flashing green station.         + False positive vials must be returned to the instrument within 5 hours.   21. For positive related vials, follow steps 1-20 in this section. A related vial is the second bottle in the  blood culture set (aerobic and anaerobic bottle) to become positive. The related vial will have the  same Accession number, therefore the same collection date, time and source, as the first positive  bottle. MANUAL BLOOD CULTURES If blood bottles are received into the laboratory that does not meet the criteria for the Bactec™ system, they will be monitored off-line for growth.   1. Place the bottle(s) into the 35° C incubator. 2. Macroscopically examine the bottle(s) each day for seven days. 3. Perform blind subcultures to CHOC and ASB2 and perform acridine orange (AO) stains at 24h, 48h, and 5 days. 4. Examine plates at 24hand 48h before discarding as negative. 5. Perform identification and susceptibility of organism(s) grown on solid media according to laboratory protocol.  ANAEROBIC BOTTLE ONLY If only an anaerobic bottle is received, contact the station and see if the patient is still available to be drawn for an aerobic bottle. If not, the anaerobic bottle should be incubated off-line according to the following protocol.   1. Release the CO2 in the vial by wiping the septum with a 70% alcohol wipe and inserting a venting needle. 2. Place the bottle into the 35º C incubator. 3. Macroscopically examine the bottle each day for seven days. 4. Perform blind subcultures to CHOC and ASB2 and perform AO stains at 24h, 48h, and 5 days. 5. Examine plates at 24h and 48h before discarding as negative. 6. Perform identification and susceptibility of organism(s) grown on solid media according to laboratory protocol.   **FALSE POSITIVE BOTTLE**   1. Mark False Positive (gram stain smear negative) bottles with a tape “flag” labeled TSUB.   This will alert day shift techs for AO stain and terminal subculture.   1. Return the flagged bottle to the Bactec. 2. When the False Positive bottle becomes a Bactec out of protocol negative at 5 days: 3. Perform terminal subculture (TSUB): Inoculate CHOC and ASB2 plates. 4. Label the plates with the current date, the current time, mark them “A” for the aerobic bottle or “N” for the anaerobic bottle (use the barcode labels) and incubate. 5. Minneapolis day Micro also performs AO stain from these TSUB bottles.    1. St Paul Bactec FX (Day shift) – Same day-- aspirate a one mL aliquot from the flagged bottle, and send in a labeled tube and the bottle to Minneapolis Micro for the AO stain.    2. St Paul Micro -- Send subculture plates to Minneapolis Micro next day    3. Minneapolis Bactec FX (Day shift) – CDSK will provide an aliquot for AO stain. 6. Minneapolis Micro --examine plates at 24h and 48h before discarding as negative.   **Contamination**  Care must be taken to prevent contamination of the sample during collection and inoculation into the Bactec™ vials. A contaminated sample will give a positive reading, but this does not indicate a clinically significant result. Such a determination must be made by the user, based on such factors as type of organisms recovered, occurrence of the same organism in multiple cultures, patient history, etc.  **Recovery of SPS Sensitive and Fastidious Organisms**  Because blood can neutralize the toxicity of SPS toward organisms sensitive to SPS (such as some *Neisseria* species and Streptobacillus sp), the presence of optimum volumes of blood, based on media type, benefits the recovery of these organisms. To enhance the growth of SPS sensitive organisms when less than optimum volumes of blood are inoculated, additional whole human blood may be added.  Some fastidious organisms, such as certain *Haemophilus* species, require growth factors, such as NAD, or factor V, which are provided by the blood specimen. If the blood specimen volume is 0.5 mL or less for Bactec™ Peds Plus/F or 3.0 mL or less for Bactec™ Plus Anaerobic/F, an appropriate supplement may be required for recovery of these organisms. Bactec™ BRAND FOS™ Fastidious Organism Supplement or whole human blood may be used as nutritional supplements.  **Non-viable Organisms**  A gram-stained smear from a culture medium may contain small numbers of non-viable organisms derived from medium constituents, staining reagents, immersion oil glass slides and specimens used for inoculation. In addition, the patient specimen may contain organisms that will not grow in the culture medium or on media used for subculture. Such specimens should be subcultured to special media as appropriate.  **Antimicrobial Activity**  Neutralization of the antimicrobial activity by resins varies depending on dosage level and timing of specimen collection. Studies have demonstrated that the resins present in this medium do not adequately neutralize imipenem-cilastatin antimicrobial preparations.  **Susceptibility Testing of *Salmonella* isolates**  To set up susceptibilities on *Salmonella* isolates use the Urine/GN KB disks on MH agar. Report ampicillin, ciprofloxacin, trimethoprim-sulfa, and ceftriaxone.  **Recovery of *Streptococcus pneumoniae***  In aerobic media, *S. pneumoniae* will typically be visually and instrument positive, but in some cases no organisms will be seen on gram stain or recovered on routine subculture. If an anaerobic vial was also inoculated, the organism can usually be recovered by performing an aerobic subculture of the anaerobic vial, since this organism has been reported to grow well under anaerobic conditions.  **Subacute Bacterial endocarditis-SBE**  The causative agents of bacterial endocarditis grow on the valves of the heart, and often are shed intermittently, and at a low level. Therefore in order to allow them time to grow for detection by the Bactec system, the protocol should be changed to 14 days.  **Recovery of *Brucella* spp. and *Francisella tularensis*** Special handling is required for the recovery of *Brucella spp*. and *Francisella tularensis* from blood cultures. Incubate *Brucella* (BRCL) for 10 days. Do a blind subculture, gram and Acridine orange stain at 5 days and terminal subculture with gram and AO at 10 days. Refer to the LRN Level Bioterrorism Laboratory Protocols Procedure, in the Safety folder for more specific information. | | | | | | | | | | |
| **Limitations** |
| **Method Performance Specifications** | Optimum recovery of isolates will be achieved by adding the appropriate volume of blood for the type of vial inoculated. Use of lower or higher volumes may adversely affect recovery and/or detection times. Blood may contain antimicrobials or other inhibitors, which may slow or prevent the growth of microorganisms. False negative readings may result when certain organisms do not produce enough CO2to be detected by the system or if significant growth has occurred before placing the vial into the system. False positives may occur when the white blood cell count is high.  It is recommended that related vials remain out of the instrument for no more than 10 minutes to minimize the possibility of the vial becoming a “false” positive vial. | | | | | | | | | | |
| **Result Reporting** | **REPORTING POSITIVE BLOOD CULTURES ON EVENING AND NIGHT SHIFTS**   1. **Positive** results must be reviewed by a second tech on the shift before reporting. 2. Record ALL results on the Bactec “Current Pos” print-out.   Record whether ‘A” or ‘N”  Record/write the gram results.  Record initials and initials of second tech.  Record the “Called to”, with date and time.   1. Critical Value: All positive blood cultures are reported immediately by phone to the physician or nursing station. A related vial gram stain does not need to be called if the results are identical. New organisms in a related vial will qualify for immediate reporting. Call Infection Control with gram stain results that appear to be gram-negative diplococci/gram negative cocci. 2. Document in the computer, the person called, their credentials (MD,RN,CNP,etc) and the date and time of the call. 3. Report the gram stain results using codes or the F8 function keys. Gram stain results are not quantified for blood cultures.   A related vial gram does not need to be phoned only if the results are identical to the first positive bottle. A related vial is the second bottle in the set to become positive and it will have the same accn as the first bottle.  **RESULTS:** Codes: Function keys:  GRAM POSITIVE COCCI GPC key 2  IN CLUSTERS CLS key 3  IN PAIRS PA no key for PA  IN PAIRS AND CHAINS PCHS no key for PCHS  IN CHAINS CHS key 4  GRAM NEGATIVE RODS GNR key A  GRAM POSITIVE RODS GPR no key for GPR  GRAM NEGATIVE COCCI GNC no key for GNC  GRAM NEG COCCOBACILLI GNEG-CC no keys  YEAST YEAS key O  BEING ISOLATED AND IDENTIFIED BIID key >  \*\*Called to and read back by CAL key C  GRAM STAIN GMS no key for GMS  **EXAMPLES:**  Observations: 1. GRAM POSITIVE COCCI IN CLUSTERS BEING ISOLATED AND IDENTIFIED  2. \*\*Called to ER (Dr. Smith) 2230 05/19/2008 GRAM STAIN  Using codes: 1. **GPC** (tab) **CLS** (tab) **BIID** (down arrow)  2. **CAL** (tab) (2 semicolons) **ER (Dr. Smith) 2230 05/19/08** (tab) **GMS**  Using the Function keys:  1. **key 2** (tab) **key 3** (tab) **key >** (down arrow)  2. **key C** (tab) (2 semicolons) **ER (Dr. Smith) 2230 05/19/08** (tab) **GMS** (no key, have to use code)  Observations: 1. GRAM NEGATIVE RODS BEING ISOLATED AND IDENTIFIED  2. \*\*Called to L8 (Mary, RN) 1715 05/20/2008 GRAM STAIN  Using codes: 1. **GNR** (tab) **BIID** (down arrow)  2. **CAL** (tab) (2 semicolons) **L8 (Mary P.,RN) 1715 05/20/08** (tab) **GMS**  Using the Function keys:  1. **key A** (tab) **key >** (down arrow)  2. **key C** (tab) (2 semicolons) **L8 (Mary P.,RN) 1715 05/20/08** (tab) **GMS** (have to use code, no key)  Observations: 1. YEAST BEING ISOLATED AND IDENTIFIED  2. \*\*Called to NICU (Dan, RN) 0320 05/21/2008 GRAM STAIN  Using codes: 1. **YST** (tab) **BIID** (down arrow)  2. **CAL** (tab) (2 semicolons) **NICU (Dan B., RN) 0320 05/21/08** (tab) **GMS**  Using the Function keys:  1. **key O** (tab) **key >** (down arrow)  2. **key C** (tab) (2 semicolons) **NICU (Dan B., RN) 0320 05/21/08** (tab) **GMS** (have to use code, no key) REPORTING POSITIVE BLOOD CULTURES ON DAY SHIFT  1. Critical Value: All positive blood cultures are reported immediately by phone to the Physician or nursing station. Call Infection Control with Gram stain results that appear to be gram-negative diplococci/gram negative cocci and also all *Neisseria meningitidis* isolates. 2. Document in the computer, the person called, their credentials (MD, RN, CNP, etc) and the date and time of the call. 3. Report and record all results and workups in Sunquest Microbiology Result Entry, in the Culture Entry tab using customized keyboards or by entering a code in the result box.   Observations: 1. GRAM POSITIVE COCCI IN CLUSTERS BEING ISOLATED AND IDENTIFIED  2. \*\*Called to Dr. Plouff at 0830 09/23/2006 GRAM STAIN  3. Susceptibilities to follow  Workups: Wkup # 1 Workup components:  Med : BPNK SC : CHOC SB ASB2  Desc : POS GMS : STPH  Id : UNKN  Wkup # 2 Workup components:  Med : BORG SC : CHOC SB ASB2  Desc : RELA GMS : NOS  Id : UNKN  Observations: 1. GRAM NEGATIVE RODS BEING ISOLATED AND IDENTIFIED  2. \*\*Called to HOC (Mary P.,RN) at 0830 09/23/2006 GRAM STAIN  3. Susceptibilities to follow  Workups: Wkup # 1 Workup components:  Med : BPNK SC : CHOC SB ASB2 CNA MAC  Desc : POS GMS : GMNR  Id : UNKN   1. Culture plates that are no growth after 1 day should be taped closed and labeled as ’NG1-work up in hood’.   These cultures may be a slow growing highly infectious organism and should be worked up in the BSC for our own safety. Please refer to the LRN Level Bioterrorism Laboratory Protocols Procedure, in the Safety folder for more specific information.   1. Negative blood cultures are updated each day in Sunquest Microbiology Automatic No-Growth Result Entry. 2. Enter BC in the Worksheet box and click **Add**. Then enter BC2 and click **Add**. 3. Click the **Add** button a little lower on the screen. 4. Click the **Start Update** button at the bottom of the screen. 5. Sunquest will complete the update and the window will close. 6. Blood cultures are automatically finaled as: No Growth 5 Days  **Call Infection Prevention with Gram stain results that appear to be gram-negative diplococci morphologically resembling *Neisseria* sp. Also inform Infection Prevention when *Neisseria meningitidis* has been isolated and confirmed. Document date and time called in the computer.**If growth should occur or additional testing should be requested after the culture has been finalized, remove the final status and send out a supplementary report using the code SRPT in SREQ or CULTURE RESULTS. Refinal the culture when identifications and/or testing is complete. If a culture requires a correction, the code **CORR** (corrected report) must be used in CULTURE RESULTS. Refer to the procedure [*Corrected Laboratory Reports*](file:///\\kidsnet.childrenshc.org\chcdfs\dept\LAB\Micro%20Procedure%20Manuals\MC%20100%20%20%20%20Quality,Spec.%20mgmt.,Labeling,Proc.,Sendout%20Results,Billing,%20PT%20testing,Addl%20Projects\MC%20102%20%20%20Labeling%20Errors,%20Specimen%20mixups,%20Corrected%20reports.doc) | | | | | | | | | | |
| **References** | 1. Bactec™ Peds Plus/F Culture Vials Insert. Document PP-091-JAA, 2008. Becton Dickinson Microbiology Systems. 2. Bactec™ Plus Anaerobic/F Culture Vials Insert. Document 8085859, 2009. Becton Dickinson Microbiology Systems. 3. Bactec™ FX System User’s Manual. 08/2008. Becton Dickinson Microbiology Systems. 4. Garcia, Lynne. Editor in chief...3rd edition, 2010. Clinical Microbiology Procedures Handbook. American Society for Microbiology, Washington, DC. 5. Versalovic, James, editor in chief, Manual of Clinical Microbiology. 10th ed. American Society for Microbiology, Washington, DC, 2011 6. Howden, R.J. J. Clin. Path. 1976, 29:50-53. 7. Recommendations for preventing transmission of Human Immunodeficiency Virus and Hepatitis B Virus to patients during exposure-prone invasive procedures. MMWR 1991, Vol. 40, No RR-8. 8. Blood borne Pathogens. Code of Federal Regulations, Title 29, Part 1910.1030 Federal Register 1991, 56:64175-64182. 9. Baron E. J, M.P Weinstein, W.M. Dunne, Jr., P. Yagupsky, D.F. Welch, and D.M. Wilson. 2005. Cumitech 1C, Blood Culture IV. Coordinating ed., E.J.Baron, American Society for Microbiology, Washington, DC. 10. Principles and Procedures for Blood Cultures, CLSI Guidelines, 2007, M47-A, Vol. 27. 11. Quality Control for Commercially Prepared Microbiological Culture Media, CLSI, 2004, M22-A3, Vol. 24. | | | | | | | | | | |
| **Appendices** | WORKLABEL MEDIA-FORM DEFINITION  BATTERY: BC  SPEC MEDIA  0 BPNK, BORG | | | | | | | | | | |
| **Training Plan/ Competency Assessment** | Training Plan | | | | | | | Initial Competency Assessment | | | |
| 1. Employee must read the procedure 2. Employee will observe trainer performing the procedure. 3. Employee will demonstrate the ability to perform procedure, record results and document corrective action after instruction by the trainer. | | | | | | | Direct observation  1. Complete written exam. | | | |
| **Historical Record** | **Version** | | **Written/Revised by:** | | | | **Effective Date:** | | **Summary of Revisions** | | |
| 1.0 | | Eileen Brinkman | | | | 6/28/2010 | | Initial Version | | |
| 1.1 | | Eileen Brinkman | | | | 10/14/2010 | | Deleted related vial information and added delayed vial entry. | | |
| 1.2 | | Becky Carlson | | | | 1/05/2014 | | Added positive culture AST statement | | |
| 1.3 | | Becky Carlson | | | | 11/01/2014 | | Added gram review by 2nd tech | | |  |  |
| 2 | | Becky Carlson | | | | 4/14/205 | | Re-numbered from MC 403 | | |
| 3 | | Susan DeMeyere | | | | 5/23/2017 | | Changed procedure for Salmonella testing. | | |
| 4 | | Susan DeMeyere | | | | 8/30/2017 | | Added removal of negative bottles on all shifts. Added retaining negative bottles with TSUB flag for false positive workup. | | |
| 5 | | Susan DeMeyere | | | | 9/5/2017 | | Added instructions to tape plates closed that are suspicious of Brucella. Added to tape closed and label plates with no growth as NG1-work up in hood. | | |
|  | |  | | | |  | |  | | |
| Archived by: |  | | | | Archived Date: | | |  | | |