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| Throat Culture | | | | | | | |
| **Purpose** | This procedure provides instruction for THROAT CULTURE for the Microbiology laboratory. | | | | | | |
| **Policy Statements** | This procedure applies to Microbiologists/Virologists who perform culture set-up and plate reading. | | | | | | |
| Principle and Clinical Significance | The majority of cases of pharyngitis are of viral etiology. At least 9 different viruses have been implicated, some of them producing symptoms indistinguishable from those of streptococcal infection such as severe sore throat with ulceration, exudate, and fever. Most cases of bacterial pharyngitis (up to 98%) are caused by beta-hemolytic streptococci, primarily those in group A. There is some evidence that other beta-hemolytic streptococci, especially groups C and G and *Arcanobacterium haemolyticum* (*Corynebacterium*) may occasionally cause pharyngitis. These organisms are thought to be self-limiting and so far have not been associated with serious sequelae as seen with group A.  A throat swab may also be helpful in determining the cause of epiglottitis, which can be life threatening causing airway obstruction. Epiglottitis is usually caused by Haemophilus *influenzae* type b but is occasionally caused by *S. pneumoniae* and *S. pyogenes.*  Young cystic fibrosis (CF) patients usually do not produce sputum. In these patients, deep throat swabs are cultured for potential respiratory pathogens. Placing a swab in the pharynx induces coughing, resulting in lung secretions being brought up into the pharynx.  Refer to the Corynebacterium diphtheriae Culture Procedure for *C. diphtheriae* information | | | | | | |
| **Test Code** | TC | | | | | | |
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|  | **Reagents** | | **Supplies** | **Equipment** | | **Media** | |
| **Materials** | * 10% sodium desoxycholate reagent * 3% hydrogen peroxide * Vitek® GP, GN, NH, YST and AST cards * Catarrhalis Test disk * Gram Stain reagents * Mueller Hinton for Cefoxitin screening * Strep grouping kit * Oxidase reagent * Staphaurex™ kit * Other supplies as necessary for the identification of common agents | | * Glass slides | * Ambient air incubator * Anaerobic jar * CO2 incubator * Incinerator * Inoculating loop * Microscope | | Refer to the Sunquest specimen label for media information. The specimen site determines appropriate media, i.e., CF patients require a PCM, SABC and CSA.   * Anaerobic Sheep Blood agar,   --1 day -- (ASB1)   * Chocolate agar (CHOC) * CNA agar (CNA) * MacConkey agar (MAC) * Selective Strep agar (SSBA) * for CF and PCD patients only: * Pseudomonas cepacia agar (PCM), * Sabouraud with CC (SABC) * Chrome Staph aureus (CSA) * Serum tellurite agar (TELL)   + for Corynebacterium diphtheriae Culture | |
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|  |  | | | | | | **Related document** |
| Sample | 1. Acceptable specimens  * Throat swab  1. SDES codes/Specimen type  * EPI – epiglottis * SUBG – subglottis * THCF – throat, CF * THR – throat * THPCD-throat, primary ciliary dyskinesia * THRD – throat for *C. diphtheriae* * *—*See *Corynebacterium diphtheriae* Culture Procedure MC 1.10  1. Specimen Collection and Transport  * Refer to Lab Test Directory – [Throat Culture, Routine](http://www.childrensmn.org/Manuals/Lab/MicroBioViral/033688.asp)  1. Specimen assessment  * Refer to the Specimen Rejection section of Lab Test Directory – [Throat Culture, Routine](http://www.childrensmn.org/Manuals/Lab/MicroBioViral/033688.asp) | | | | | | [Lab Test Directory – Throat Culture, Routine](http://www.childrensmn.org/Manuals/Lab/MicroBioViral/033688.asp) |
| **Special Safety Precautions** | Microbiologists/virologists are subject to occupational risks associated with specimen handling. Refer to the safety policies**:**   1. Biohazard Containment 2. Safety in the Microbiology/Virology Laboratory  * Biohazardous Spills | | | | | | |
| **Procedure** | InoculationWarm all media before inoculation.Label all plates and slide properly with the patients name, accession number and date.Inoculate the media in the order of the least selective first to prevent carryover of inhibitory substances to another medium. Refer to the Sunquest specimen label for the order of inoculation.Specimen processing  1. Roll swab across the upper quadrant of the CHOC, SB, CNA, MAC, SSBA, (CSA, SABC, PCM) touching all surfaces of the swab. 2. Streak plates semi quantitatively for primary isolation. 3. Sterilize the inoculating loop in the incinerator for 5 s to 10 s. Allow the loop to cool. 4. Pass the loop through the edge of the first quadrant approximately 4 times while streaking into the second quadrant. Continue streaking in the second quadrant without going back into the first quadrant 3-4 times. 5. Flame loop again, turn the plate another quarter of a turn, and pass the loop through the edge of the second quadrant approximately four times while streaking into the third quadrant. Continue streaking in the third quadrant without going back into the second quadrant 3-4 times 6. Make small cuts in the primary area of the SB for hemolysis.  1. Incubation 2. Incubate CHOC, CNA and SSBA in 4-10% CO2 at 35ºC 3. Place ASB1 in anaerobic holding chamber to be closed in an AnaeroPack™ or Ana bag system for 1 day. 4. Place MAC, (CSA, SABC, and PCM) in ambient air incubator at 35ºC. 5. Culture examination 6. Day 1 7. Examine plates. 8. Gram stain each colony type and perform initial identification procedures as needed, i.e., catalase, oxidase, bile solubility, etc. Bile solubility only needs to be performed if the alpha strep is predominant and suspicious for S. pneumoniae. 9. Set up definitive biochemical or identification procedures on significant organisms if well isolated. 10. Perform antimicrobial susceptibility testing on significant organisms if well isolated. 11. Subculture organisms that are not well isolated to appropriate media for further work-up. 12. Reincubate primary plates and subcultures for an additional day. 13. Report preliminary results. 14. Day 2 15. Examine primary plates from the previous day for additional microorganisms. 16. Read and record identification tests and susceptibilities from the previous day. 17. Set up additional tests as needed. 18. Hold CF culture plates for a minimum of three days. Hold PCM for 5 days. 19. Confirm *B. cepacia* identification on commercial systems with conventional biochemical tests. Send *B. cepacia* to the University of Michigan reference laboratory (see appendix) and freeze isolate for future reference. Alert physician. 20. MRSA isolation requires a “Called to” if not from E.D. (disch.), or a repeat isolate. Freeze isolates for future reference. 21. Send updated or final report. 22. Save a representative primary plate, whether a complete work-up was performed or not, at room temperature for 7 days in case a physician calls for further studies. 23. Additional Days 24. Complete identification procedures until all significant isolates are finished. 25. Send updated report and finalize. | | | | | | |
| **Method Performance Specifications** | 1. Report the presence of “USUAL UPPER RESPIRATORY FLORA” (Sunquest code **UOF**) which includes:   *Actinomyces* sp. *Haemophilus influenzae*, non-typable  alpha-hemolytic streptococci, viridans grp. *Haemophilus parainfluenzae*  *Capnocytophaga* sp. *Lactobacillus* sp.  Coagulase-negative staphylococci *Micrococcus* sp.  *Corynebacterium* sp. *Moraxella catarrhalis*  *Eikenella corrodens* *Neisseria* sp. (not GC or NMEN)  Gamma-streptococci, viridans grp. *Stomatococcus* sp.  *Streptococcus pneumoniae* Yeast in low numbers (1+) (non-CF patients)  Gram-negative rods in low numbers (1+)(non-CF patients)  Beta-hemolytic streptococcus group F  Small colony β strep (colonies <0.5 mm)   1. Perform identification tests for the following:   *Arcanobacterium haemolyticum*  Beta-hemolytic streptococci, large colony (>= 0.5 mm) groups A, C, and G  *Haemophilus influenzae*, mucoid strains (send typable strains to MDH)  *Neisseria meningitides*  *S. aureus*   1. Perform identification tests on possible pathogens that are the predominate organism. These may include the following:   *Haemophilus influenzae*  *M. catarrhalis*  *Streptococcus pneumoniae*   1. *Streptococcus pyogenes* (group A beta-hemolytic strep) is generally susceptible to penicillin and its derivatives; therefore, susceptibility need not be routinely performed. The principal reason for considering an alternative drug for individual patients is allergy to penicillin. Erythromycin, a Cephalosporin, or Clindamycin might be substituted in these cases. Patients allergic to penicillins may also be allergic to cephalosporins.  * Add Sunquest code **BHSS** to report: Beta hemolytic streptococci are susceptible to penicillin, cephalosporins and vancomycin. Some strains may be resistant to erythromycin. * Call Children’s Respiratory Clinic with *Streptococcus pyogenes* (group A β strep) isolates.  1. Traditional pathogens for cystic fibrosis (CF) patients are as follows:  |  |  |  |  | | --- | --- | --- | --- | | Organism | Usual oral flora | Identify | Processing level | | *S. aureus* | | NEVER | Any amount | Perform cefoxitin screening and/or AST (Vitek2) | | *Ps. aeruginosa* (matte and mucoid colony types) | | NEVER | Any amount | Perform ID and AST (Vitek2, KB or MSCN)—**include KB AZT disk** | | *Alcaligenes xylosoxidans* | | NEVER | Any amount | Perform ID and AST (Vitek2 or MSCN) —**include KB AZT disk** | | *Burkholderia cepacia* complex  **Confirm ID with conventional tube biochemicals** | | NEVER | Any amount | Perform ID and AST (Vitek2 MSCN) —**include KB AZT disk** | | *S. maltophilia* | | NEVER | Any amount | Perform ID and AST-all 4 drugs  MSCN OR Vitek2 | | *Burkholderia gladioli,*  *B. bronchiseptica, Pandoraea sp.*  and *Ralstonia sp.* | | NA  (Considered emerging pathogens) | Any amount | Perform ID only; AST upon request (MSCN) | | *Enterobacteriaceae sp.* | | NEVER | Any amount | Perform ID only; AST upon request | | *H.* *influenzae* | | Not predominant | Predominant | Perform ID and β-lactamase; Serotype if colony is mucoid or wet | | *S. pneumoniae* | | Not predominant | Predominant | Perform AST (Vitek2 or MSCN) | | Moulds, *Aspergillus* sp. | | NEVER | Any amount | Perform presumptive ID only; send to MDH if complete ID requested. AST upon request | | Rapid growing mycobacteria | | NEVER | Any amount | Identify to species level; send to MDH for ID |   **CF Notes:**   1. *S. aureus* (CF):  * Multiple strains may be present in one specimen; look for subtle differences. * Small colony variant strains may be present due to long-term trimethoprim-sulfamethoxazole (SXT) treatment. They are small, flat, gray colonies, generally MRSA.  1. *Burkholderia cepacia* complex*:*  * *B. cepacia* will appear pink on PCM. Confirm with conventional biochemicals. * *S*end isolates to the *B. cepacia* Research Laboratory and Repository. See appendix for details. * Freeze isolate for future reference. Alert physician.  1. Yeast: Report as “YEAST, No further identification.” Perform identification upon request. 2. Molds: Identify all molds. Perform Lacto Phenol Cotton Blue exam. If conidia are consistent with *Aspergillus* sp., report presumptive *Aspergillus* sp. SUMP-ASPE. All other filamentous fungi: refer to MDH for ID. | | | | | | |
| **Result Reporting** | Culture Results: Record culture results and culture work-ups in Sunquest MRE *Culture Entry* tab in Observations or Workups by using customized keyboards or by entering a code in the result box. Report results semiquantitatively, i.e., 1+, 2+, 3+ or 4+.  |  |  |  |  | | --- | --- | --- | --- | | Quantity | 1st quadrant  # colonies | 2nd quadrant  # colonies | 3rd quadrant  # colonies | | 1+ | <10 |  |  | | 2+ | >10 | <5 |  | | 3+ | >10 | >5 | <5 | | 4+ | >10 | >5 | >5 |  1. Culture with no predominate organisms or no potential pathogens (PP), report as follows:   Observations: 1. 4+ USUAL UPPER RESPIRATORY FLORA Sunquest code: **UOF**  2. Susceptibilities not performed. Please contact Microbiology if further testing is required (MPLS 813-5866). Sunquest code: **SNP**  Workups: Wkup# 1 Workup Components  Med : SB BS : NEG  Desc : AHS-GHS-NEIS-HAEM-SMUC GMS : STR-HAE-NEIS (Add Wkld: 3)  Id : UOF   1. Culture with PP, report as follows:   Observations: 1. 4+ USUAL UPPER RESPIRATORY FLORA  2. 3+ PSEUDOMONAS AERUGINOSA (MUCOID)  3. 2+ PSEUDOMONAS AERUGINOSA (MATTE)  4. 2+ STAPHYLOCOCCUS AUREUS Further identification to follow  5. 1+ ASPERGILLUS SPECIES  6. Susceptibilities to follow.  Workups: Wkup# 1 Workup Components  Med: SB BS: NEG  Desc: AHS-GHS-NEIS-SMUC  Id: UOF   1. MRSA isolation requires a “Called to” if not from E.D. (disch.), or a repeat isolate. Document date and time called in computer.   Observations: 3. 3+ METHICILLIN-RESISTANT STAPH AUREUS \*\*\*MDRO\*\*\* Sunquest code: **MRSA**  4. MULTIPLE DRUG RESISTANT ORGANSIM (MDRO): This organism requires SPECIAL CONTACT PRECAUTIONS. Please call Infection Control. Sunquest code: **DRO**  5. \*\*Called to Linda S., RN L8 @ 1300 7/7/03 Report strains of *P. aeruginosa* as matte or mucoid using the Sunquest codes **PMUC** or **PMAT**Report mucoid strains of S. pyogenes using the Sunquest code **BSA – MUCO.** Review **Culture Summary** for accuracy before filing report.If a culture requires a correction, the code **CORR** (corrected report) must be reported on an observation line in the *Direct Exam* or *Culture Entry* tab. Refer to policy MC 5.1 LABELINGERRORS/SPECIMEN MIXUPSfor Sunquest report entry information.If growth should occur or additional testing should be requested after the culture has been finalized, remove the final status and send out a supplementary report. The code SRPT (supplementary report) must be used in SREQ or *Culture Observations* as follows:Updated or new culture information: In the *Culture Entry* tab, enter SRPT on an observation line followed by new results.Requests for additional testing: In the *Misc. Updates* tab, enter **SRPT** in SREQ followed by the request.  * Refinal the culture when identifications and/or testing are complete. | | | | | | |
| **References** | 1. York, M., et al., Section 3, Aerobic bacteriology, 3.11.2, 3.11.3, *In* L.S. Garcia (ed) *Clinical Microbiology Procedures Handbook*, 2010, American Society for Microbiology, Washington, D.C. 2. Gilligan, P.H., et.al, 2006, Cumitech 43, *Cystic Fibrosis Microbiology*, ASM Press, American Society for Microbiology, Washington, D.C. 3. Forbes, B.A., et al., Bailey & Scott’s *Diagnostic Microbiology*, twelfth edition, 2007. Mosby, Inc., St. Louis, MO., pg. 814-818. 4. Versalovic, James. et al, *Manual of Clinical Microbiology*, 2011, ASM press, American Society for Microbiology, Washington, D.C. 5. Pezzlo, M., Section 1, Aerobic bacteriology, 1.14, Processing and interpretation of upper respiratory tract specimens, *In* H.D. Isenberg (ed) *Clinical Microbiology Procedures Handbook*, 1994, Vol. 1, American Society for Microbiology, Washington, D.C. 6. Whittier, Susan, Update on the microbiology of cystic fibrosis: traditional and emerging pathogens. *Clinical Microbiology Newsletter*, Vol. 23, No. 9, pg. 67-71. 7. Hoppe, J.E., U. Theurer-Mainka, M. Stern, Comparison of three methods for culturing throat swabs from cystic fibrosis patients, *Journal of Clinical Microbiology,* Vol. 33, No. 7, 1995, pg 1896-1898. | | | | | | |
| **Appendices** | WORKLABEL MEDIA FORM DEFINITION  BATTERY: TC  SPEC MEDIA  0 CHOC, ASB1, MAC, SSBA, CUT3  EPI CHOC, SB, CNA, MAC, CUT3  SUBG CHOC, SB, CNA, MAC, CUT3  THCF CHOC, ASB1, CNA, MAC, PCM, SABC, CSA, CUT3  THR CHOC, ASB1, MAC, SSBA, CUT3  THRD SB, TELL  *B. cepacia* Research Laboratory and Repository—see “white binder” on shelf for protocol and forms  Dr. LiPuma’s Lab -- CFF *Burkholderia cepacia* Research Laboratory and Repository  [jlipuma@umich.edu](mailto:jlipuma@umich.edu)  The University of Michigan Health System  1150 W. Medical Center Drive  Ann Arbor, MI 48109-5646 | | | | | | |
| **Training Plan/ Competency Assessment** | **Training Plan** | | | | **Initial Competency Assessment** | | |
| 1. Employee must read the procedure 2. Employee will observe trainer performing the procedure. 3. Employee will demonstrate the ability to perform procedure, record results and document corrective action after instruction by the trainer. | | | | 1. Direct observation | | |
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| **Historical Record** | **Version** | **Written/Revised by:** | | **Effective Date:** | | **Summary of Revisions** | |
| 1.0 | Pat Ackerman | | 1978 | | Initial Version | |
| 1.1 | Pat Ackerman | | 04/1983 | |  | |
| 1.2 | Pat Ackerman | | 04/1988 | |  | |
|  | 1.3 | Pat Ackerman | | 07/29/2003 | |  | |  |  |
| 1.4 | Pat Ackerman | | 12/10/2004 | |  | |
| 1.5 | Pat Ackerman | | 08/22/2007 | | Updated Sunquest 6.2 reporting information. Revised SRPT and CORR statements. Updated CF traditional organisms into table format. Limit yeast identification to physician request only. Added code information BHSS. | |
| 1.6 | Jessica Craig | | 06/16/2010 | | Updated into online format. | |
| 1.7 | Becky Carlson | | 3/19/2014 | | Updated CF table; removed mucoid GAS referral to Dr Kaplan | |
| 1.8 | Becky Carlson | | 4/18/2015 | | Re-numbered from MC 429. Added SSBA to list of set up medias | |
|  |  | |  | |  | |
| **3** | Eileen Brinkman  Susan DeMeyere | | **2/5//2018** | | Updated bile solubility testing on alpha streps. | |
| **Archived by:** |  | | **Archived Date:** | |  | |