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| Transfusion Reaction Culture | | | | | | | | |
| **Purpose** | This procedure provides instruction for TRANSFUSION REACTION CULTURE for the Microbiology Laboratory. | | | | | | | |
| **Policy Statements** | This procedure applies to microbiologists who perform culture set-up and plate reading. | | | | | | | |
| Principle and Clinical Significance | Transfusion of blood products and blood components is usually safe and effective therapy. However, transfusion reactions can occur. A serious complication of transfusion therapy is the transmission of bacterial organisms. Although bacterial contamination of blood and blood products is rare, it remains a significant concern. The use of a closed system for blood collection, the implementation of rigid standards for blood bank operations and stringent attention to skin disinfection has contributed to the reduction of bacterial contamination. If a transfusion reaction should occur, inoculate two sets of blood culture bottles from the suspected blood unit, incubating one set at 35ºC and the other set at 25ºC. | | | | | | | |
| **Test Code** | TRXN | | | | | | | |
|  | **Reagents** | | | **Supplies** | **Equipment** | | **Media** | |
| **Materials** | * Gram Stain reagents * Acridine orange stain | | | * 10 cc syringe * 18 gauge needle * AnaeroPack sachet * BD© blood transfer device * Fenwal sampling site coupler (located in BB) * Glass slide | * Anaerobic jar/bag * BACTEC™ FX Fluorescent series * CO2 incubator * Fluorescent microscope * Incinerator * Inoculating loop * Microscope | | Refer to the Sunquest specimen label for media information. **Note**: One “set” consists of an aerobic and an anaerobic blood culture bottle.   * 2 BACTEC™PEDS PLUS/F aerobic medium (pink cap) * 2 BACTEC™ Lytic/10 Anaerobic/F medium (purple cap) * Chocolate agar (CHOC) * Sheep Blood agar (SB) * Anaerobic sheep blood agar (ASB2) | |
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| Sample | * Acceptable specimens * Blood or blood components from transfusion bag * SDES codes/Specimen type * DON -; unit number (use free text to add unit number) ; XXXXXXXX * If blood, use the code DON - BLD -; unit number XXXXXXXXXXXXX * If platelets, use free text DON -; PLATELETS unit number XXXXXXXXXXXXX * If transfusion bag is empty, order culture and result with cancel code QNS. Credit culture in CRW. | | | | | | |
| **Special Safety Precautions** | Microbiologists/virologists are subject to occupational risks associated with specimen handling. Refer to the safety policies**:**   1. [*Biohazard Containment*](https://starnet.childrenshc.org/References/labsop/mcvi/safety/mcvi-3.1-biohazard-containment.pdf) 2. [*Safety in the Microbiology/Virology Laboratory*](https://starnet.childrenshc.org/References/labsop/mcvi/safety/mcvi-3.2-safety-in-the-microbiology-lab.pdf)  * [*Biohazardous Spills*](https://starnet.childrenshc.org/References/labsop/mcvi/safety/mcvi-3.4-biohazardous-spills.pdf) | | | | | | | |
| **Procedure** | Specimen processing  1. Use the Fenwal sampling site coupler to enter the unused port on the blood bag. **Do not** use the blood administration kit. 2. Cleanse the rubber septum of each blood culture bottle with 70% alcohol and the port of the blood bag. Allow to dry. 3. With a syringe, remove 12ml (or as much as possible, if less than 12 ml) of blood or blood component from the unit. **Do not culture the segments**. 4. Inoculate approximately 3 ml into each of 4 blood culture bottles, saving a small amount for a Gram stain, and possible subculture. Inoculate bottles with equal amounts if less than 12 ml available. 5. If less than 4ml are available, inoculate to CHOC, SB, ASB2 and gram stain directly. 6. Prepare a slide for Gram stain, heat fix and examine immediately.  * **If organisms are observed**, report in the Sunquest MRE and notify the provider and Transfusion Services as for a positive BC. Directly inoculate a CHOC, SB and ASB2. * Incubate subculture plates as follows:   CHOC/SB - 35ºC in CO2 incubator for 2 days  ASB2 - 35ºC in anaerobic jar/bag for 2 days   1. Place one set of blood culture bottles in the Bactec Instrument. A set consists of one aerobic bottle (pink) and one anaerobic bottle (purple). 2. Hold the Room Temp set at 25ºC; Send RT bottles to Mpls with morning courier if St Paul TRXN. 3. **5 day standard incubation** 4. Incubate one set of blood cultures at 35ºC in the BACTEC™ FX following the 5-day Bactec™ protocol. 5. Incubate the other set of blood cultures at 25ºC for 5 days, (in Mpls). 6. Incubate any/all blind subculture plates as follows: 7. CHOC - 35ºC in CO2 incubator for 2 days 8. ASB2 - 35ºC in anaerobic jar for 2 days 9. **Acridine orange/Gram Stain examination** 10. Perform AO and interpret. 11. Read AO with fluorescent microscope. If positive, perform Gram stain. 12. Blot the oil from the slide and save for one week. 13. **Culture Procedure. Update negative TRXN manually in Sunquest MRE for 5 days.**      1. Day #1: 2. Perform 24 h blind subcultures and AO stain on the RT 25ºC set of blood culture bottles. 3. Cleanse the bottle septum with 70% alcohol. Allow to dry. 4. Remove 0.5 ml of blood and inoculate a CHOC from the aerobic bottle and a CHOC & ASB2 from the anaerobic bottle. Prepare a slide for acridine orange stain (AO). 5. Read AO with fluorescent microscope. If positive, perform Gram stain. 6. If positive, report in the Sunquest MRE and notify the provider and Transfusion Services as for a positive BC 7. Blot the oil from the slide and save for one week.   Day 1 cont.  Initial Blind subcultures   1. If initially set because of a positive initial gram, examine blind subculture plates at 24 h and 48 h before discarding as negative. 2. If growth is present, Gram stain each colony type and perform initial identification procedures, i.e., catalase, oxidase, etc. 3. Set up definitive biochemical or identification procedures on isolated colonies. 4. Perform antimicrobial susceptibility testing on isolated colonies. 5. Subculture organisms that are not well isolated to appropriate media for further work-up. 6. Reincubate plates and subcultures for an additional day. 7. Report preliminary results. 8. Notify Transfusion Services and the provider of any positive findings. 9. Day # 2: 10. Examine blind subculture plates at 24 h and 48 h before discarding as negative. 11. Perform 48 h blind subcultures and AO stain on the RT 25ºC set of blood culture bottles. 12. Cleanse the bottle septum with 70% alcohol. Allow to dry. 13. Remove 0.5 ml of blood and inoculate a CHOC from the aerobic bottle and a CHOC & ASB2 from the anaerobic bottle. Prepare a slide for acridine orange stain (AO). 14. Read AO with fluorescent microscope. If positive, perform Gram stain. 15. If positive, report in the Sunquest MRE and notify the provider and Transfusion Services as for a positive BC 16. Blot the oil from the slide and save for one week. 17. Examine plates from the previous day.   h. Read and record identification tests and susceptibilities if any, from the previous day.  i. Set up additional tests as needed.  j. Send updated report.  k. Save a representative primary plate, whether a complete work-up was performed or not, at room temperature for 7 days in case a physician calls for further studies.   1. Additional Days:      1. Examine blind subculture plates at 24 and 48 h before discarding as negative. 2. As needed, complete identification and susceptibility testing procedures until all significant isolates are finished. 3. Send updated report and finalize. 4. Terminal AO and subculture: 5. Perform terminal subcultures (TSUB) and AO stain on the RT 25ºC set of blood culture bottles at 5 days. 6. Cleanse the bottle septum with 70% alcohol. Allow to dry. 7. Remove 0.5 ml of blood and inoculate a CHOC from the aerobic bottle and a CHOC & ASB2 from the anaerobic bottle. Prepare a slide for acridine orange stain (AO). 8. Read AO with fluorescent microscope. If positive, perform Gram stain. 9. If positive, report in the Sunquest MRE and notify the provider and Transfusion Services as for a positive BC 10. Blot the oil from the slide and save for one week. 11. Examine plates from the previous day. 12. Hold blind subculture plates for 48 hours before discarding as negative.   Hold negative RT 25ºC bottles for 5-7 days until all plates have been read, before issuing a final report. | | | | | | | |
| **Interpretation** | 1. If the culture is negative, it is unlikely that the blood or blood product was contaminated with bacteria at the time of the transfusion reaction. 2. A positive culture does not positively identify infection as the cause of the transfusion reaction. A positive blood culture or blood product does not establish the source of contamination unless both the blood bank and patient’s samples yield the same organism. PFGE may be needed to determine if the strains are similar. | | | | | | | |
| **Procedure Notes** | 1. *Yersinia enterocolitica, Ps. fluorescens, Ps. putida,* and *Enterobacter* sp. can proliferate at 0-6ºC, the temperature at which blood is stored. These organisms have been implicated in transfusion reactions. Gram-negative bacilli can produce endotoxins that will cause rapid and sometimes irreversible shock when present in the blood. 2. Platelets are the most common cause of transfusion-related sepsis caused by organisms such as *Corynebacterium* sp., coagulase-negative staphylococci, *S. aureus, Pseudomonas* sp., and *Enterobacter cloacae.* Platelets are stored at 22ºC and provide an excellent medium for bacterial growth. 3. Contamination of blood products can be from skin flora, environmental organisms introduced at the time of collection or processing, or an asymptomatic bacteremia in the blood of a healthy donor. Water baths in which frozen blood products are thawed have also been a source of contamination. 4. If it is clinically necessary to culture a blood unit, the recipient’s blood should be cultured simultaneously. | | | | | | | |
| **Result Reporting** | Culture Results: Record culture results and culture work-ups in Sunquest MRE *Culture Entry* tab in Observations or Workups by using customized keyboards or by entering a code in the result box.  1. **No growth cultures**: Update culture status in the Observation result box (Culture Entry tab), by using the “No Growth” update key (‘). Report as “No growth “*x*” days". Final culture on day 5. 2. **Positive culture:**   Observations: 1. 4+ COAGULASE NEGATIVE STAPHYLOCOCCI (2 COLONY MORPHOLOGIES)    Workups: Wkup # 1 Workup Components  Med: SB GMS : STPH  Desc: WH SC : SB  Id: CNS SLC : POS  VMIC : 1       1. **Notify provider and Transfusion Services of positive results. Document in workup**.  Review **Culture Summary** for accuracy before filing report.If a culture requires a correction, the code **CORR** (corrected report) must be reported on an observation line in the *Direct Exam* or *Culture Entry* tab. Refer to policy [*LABELING ERRORS/SPECIMEN MIXUPS AND CORRECTING PATIENT DATA*](file://\\kidsnet.childrenshc.org\chcdfs\dept\Lab%20Procedures\Microbiology\1NEW%20Micro%20Procedure%20Manual.%20(same%20as%20in%20Starnet)\MC%20100%20%20%20%20Quality,Spec.%20mgmt.,Labeling,Proc.,Sendout%20Results,Billing,%20PT%20testing,Addl%20Projects\MC%20102%20Labeling%20Errors,%20Specimen%20Mixups,%20Corrected%20Reports%20R.docx)for Sunquest report entry information.If growth should occur or additional testing should be requested after the culture has been finalized, remove the final status and send out a supplementary report. The code SRPT (supplementary report) must be used in SREQ or *Culture Observations* as follows:Updated or new culture information: In the *Culture Entry* tab, enter SRPT on an observation line followed by new results.Requests for additional testing: In the *Misc. Updates* tab, enter **SRPT** in SREQ followed by the request.  * Refinal the culture when identifications and/or testing are complete. | | | | | | | |
| **Appendices** | WORKLABEL MEDIA DEFINITION, BATTERY : TRXN  SPEC MEDIA   1. BPNK,BPRL,BP25,BPR25 | | | | | | | |
| **References** | 1. Miller, J.Michael, Section 13, Epidemiologic and Infection Control Microbiology 3.13.1, Garcia, Lynne (ed) *Clinical Microbiology Procedures Handbook*, 2010, American Society for Microbiology, Washington, D.C. 2. Gilchrist, M. J. R., Section 11, Aerobic bacteriology, 11.11, Culture of blood bank products, *Garcia, Lynne*(ed) *Clinical Microbiology Procedures Handbook*, 2010, Vol. 2, American Society for Microbiology, Washington, D.C. | | | | | | | |
| **Training Plan/ Competency Assessment** | **Training Plan** | | | | | **Initial Competency Assessment** | | |
| 1. Employee must read the procedure 2. Employee will observe trainer performing the procedure. 3. Employee will demonstrate the ability to perform procedure, record results and document corrective action after instruction by the trainer. | | | | | 1. Direct observation | | |
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| **Historical Record** | **Version** | | **Written/Revised by:** | | **Effective Date:** | | **Summary of Revisions** | |
| 1.0 | | Pat Ackerman | | 04/11/1994 | | Initial Version | |
| 1.1 | | Pat Ackerman | | 08/12/2003 | |  | |
| 1.2 | | Pat Ackerman | | 01/14/2008 | | Updated Sunquest 6.2 reporting information. Revised SRPT and CORR statements; added notification of blood bank Technologist; added Hyper-link to labeling policy. Updated blood culture bottle information. | |
|  | 1.3 | | Pat Ackerman | | 10/12/2009 | | Updated protocol length to 5 days from 7 days. | |  |  |
| 1.4 | | Tina Gronquist | | 06/16/2014 | | Updated into online format. | |
| 2 | | Becky Carlson | | 4/14/2015 | | Re-numbered from MC 432 | |
| 3 | | Susan DeMeyere | | 12/7/2018 | | Add instructions for empty or low volume. Removed anaerobic culturing from aerobic bottles. | |
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