# Wipe Testing for Amplicon or Nucleic Acid Contamination

**PURPOSE**

* This procedure provides instruction for environmental monitoring of laboratory equipment and surfaces for DNA/RNA and amplicon contamination

#### POLICY STATEMENT

* *Bordetella* wipe testing is performed monthly
* RVP wipe testing is performed monthly
* GAS wipe testing is performed monthly
* RIP wipe testing is performed monthly
* HSV wipe testing is performed monthly
* Other targets are performed as determined
* Notify section technical director and/or designee regarding positive or unresolved results
* Discontinue patient testing during a contamination event if the technical director has determined that it is unsafe to continue

**ABBREVIATIONS**

|  |  |
| --- | --- |
| * CMA: ChromAgar MRSA
* NA: nucleic acid
* NEGC: negative control
* NFW: nuclease free water
 | * POSC: positive control
* PPE: personal protective equipment
* SB: 5% sheep blood agar
 |

## DOCUMENTATION/RECORDS

* Assay run-specific Results Report
* RVP Wipe Testing worksheet MB 3.02.F2
* BORDP Wipe Testing worksheet MB 3.02.F3
* GAS Wipe Testing worksheet MB 3.02.F4
* RIP Wipe Testing worksheet MB 3.02.F5
* HSV Wipe Testing worksheet MB 3.02.F6

## SAFETY CONSIDERATIONS

* Standard precautions

#### MATERIALS REQUIRED

|  |  |  |
| --- | --- | --- |
| **Equipment** | **Reagents** | **Supplies** |
| Room 2* Refrigerator 2 – 8° C
* VWR Multi-tube vortex
* BSC BSL-2
* Vortex Mixer
* !00 µl Concept pipettor
* 10 µl pipette
* Cold block
* Simplexa reaction disks and sealer

Room 3* + BD MAX instrument
	+ eSensor XT
	+ -20⁰ C freezer
	+ Simplexa thermocycler
	+ Pipettes, 20 µl, 200 µl
	+ Vortex Mixer
	+ UVP Hood
	+ Mini-centrifuge (2)
	+ ABI Thermocycler
 | BD MAX MRSA XT kit | Orange barrier wipes |
| eSensor XT-8 RVP kit | Nitrile gloves (powder-free) |
| BORDP ASR reagents | Disposable lab coats |
| GAS reagents | Absorbent clothes |
| RIP reagents | Test tube rack |
| BD MAX CDT kit | 2 ml Cryovial |
| Household Bleach | BBL CultureSwab |
| Eliminase | Disposable lab coats |
| Alconox | Pipette tips |
| 1% Sani-Cloth Bleach wipes (equivalent to 1:10 bleach solution) | Pipette disposal containers |
| 70% alcohol | BioHazard ZipLock baggies |
| NFW |  |

**PROCEDURE A:** Follow the general guidelines below for decontamination and cleaning

General guidelines for decontamination

| **Activity** | Step | **Action** | **Related Doc** |
| --- | --- | --- | --- |
| **PPE** | 1 | Change gloves frequently during the decontamination process, especially during the first steps of decontamination and before touching any clean surface | ***Refer to******Proc. I***Decontamination procedure following a contaminationevent |
|  | 2 | All PPE should be disposed of after decontamination |
| **Room 2****Processing** | 3 | Room 2: Processing: Perform swab testing on work surfaces and equipment, including but not limited to: | RVP Wipe testing MB 3.02.F2BORDP Wipe Testing MB 3.02.F3GAS Wipe TestingMB 3.02.F4RIP Wipe TestingMB 3.02.F5HSV Wipe Testing MB 3.02.F6 |
| * door handles
* pipettes
* work surfaces
 | * centrifuges
* vortexers
* anything with a button
 |
| **Room 3****Amplification** | 4 | Room 3: Amplification: Perform swab testing on work surfaces and equipment, including but not limited to: |
| * thermocycler block and lid
* door handles
* pipettes
* work surfaces
 | * centrifuges
* vortexers
* anything with a button
 |
| **Results** | 5 | Based on results of the wipe testing, focus decontamination efforts on the room(s) showing contamination |  |
|  | 6 | Repeat swab testing after cleaning to confirm decontamination |
|  | 7 | Re-run any swab that comes up positive for contamination and individual swabs of each component, such as pipettes, listed sites on worksheet for that area and possible expanded sites to narrow down the source |
|  | 8 | If samples continue to be positive for amplicon or genomic DNA/RNA, prepare a 1% v/v bleach/Alconox cleaning solution  |  |
|  | 9 | Repeat testing until all environmental swabs are negative |  |
|  | 10 | If any samples are positive consult with the Technical Specialist and/or Technical Director |  |

**PROCEDURE B:** Follow the steps below to collect and test Respiratory Viral Panel PCR (RVP) environmental samples

RVP environmental testing

| **Activity** | Step | **Action** | **Related Doc** |
| --- | --- | --- | --- |
| **Worksheet** | 1 | Print RVP Wipe testing worksheet * *Test expanded environmental sites during a contamination event such as drawer handles, light switches, phones, etc.*
 | RVP Wipe testing MB 3.02.F2 |
|  | 2 | Aliquot 1 mL NFW into a 2 ml cryovial for each sample to be tested |  |
| **Sample processing** | 3 | Transfer 1 ml NFW into a sterile tube to pre-moisten swabs prior to collection; can be used for multiple swabs4* Soak each swab in NFW for 5 s
 |  |
|  | 4 | Swab area by rotating the swab 2 – 3 times across the surface, approx 10 cm2  |  |
|  | 5 | Using an orange barrier protector, break the swab off into labeled sample tube (NFW) |  |
|  | 6 | Mix 10 seconds, vortex speed 8 |  |
| **PCR** | 7 | Extract with internal control; perform PCR testing directly from elution | MB 11.05RVP assay |
| **Notification** | 8 | If results are positive * *Notify* section technical director and/or designee
* Proceed to Procedure F, page 4
* Perform result “Look-Back”
* Document problem and corrective action
* Contact GenMark for further assistance if needed, 1-800-373-6767
 | **Procedure G****Decontamination** |
| **Archive** | 9 | Attach run-specific report to RVP Test Form MB 3.02.F2; place in Service manual |  |

**PROCEDURE C:** Follow the steps in the table below to collect and test *Bordetella* PCR (BORDP) environmental samples

*Bordetella* PCR Environmental Testing

| **Activity** | Step | **Action** | **Related Doc** |
| --- | --- | --- | --- |
| **Worksheet** | 1 | Print BORDP Wipe testing worksheet MB 3.02.F3* *Test expanded environmental sites during a contamination event such as drawer handles, light switches, phones, etc.*
 | BORDP Wipe Testing MB 3.02.F3 |
|  | 2 | Aliquot 250 µl NFW into a 2 ml cryovial for each sample to be tested |
| **Sample processing** | 3 | Transfer 1 ml NFW into a sterile tube to pre-moisten swabs prior to collection; can be used for multiple swabs* Soak each swab in NFW for 5 s
 |  |
|  | 4 | Swab area by rotating the swab 2 – 3 times across the surface, approx 10 cm2  |  |
|  | 5 | Using an orange barrier protector, break the swab off into labeled sample TE Buffer tube |  |
|  | 6 | Mix 5 min, vortex speed 8 |  |
| **PCR** | 7 | Perform PCR testing according to BORDP assay protocol  | MB 6.05BORDP assay  |
| **Notification** | 8 | If results are positive * *Notify* section technical Director and/or designee
* Proceed to Procedure F, page 4
* Perform result “Look-Back”
* Document problem and corrective action
 | **Procedure G Decontamination** |
| **Archive** | 9 | Attach run-specific report to BORDP Test Form MB 3.02.F3; place in Service manual |  |

**PROCEDURE D:** Follow the steps in the table below to collect and test Group A Strep (GAS) environmental samples

Group A Strep PCR Environmental Testing

| **Activity** | Step | **Action** | **Related Doc** |
| --- | --- | --- | --- |
| **Worksheet** | 1 | Print GAS Wipe testing worksheet MB 3.02.F4* *Test expanded environmental sites during a contamination event such as drawer handles, light switches, phones, etc.*
 | GAS Wipe Testing MB 3.0.2.F4 |
|  | 2 | Aliquot 250 µl TE Buffer into a 2 ml cryovial for each sample to be tested |
| **Sample processing** | 3 | Transfer 1 ml NFW into a sterile tube to pre-moisten swabs prior to collection; can be used for multiple swabs* Soak each swab in NFW for 5 s
 |  |
|  | 4 | Swab area by rotating the swab 2 – 3 times across the surface, approx 10 cm2  |  |
|  | 5 | Using an orange barrier protector, break the swab off into a labeled sample buffer tube |  |
|  | 6 | Process in the same way patient samples are |  |
| **PCR** | 7 | Perform PCR testing according to GAS assay protocol  | MB 8.05GAS assay  |
| **Notification** | 8 | If results are positive * *Notify* section technical Director and/or designee
* Proceed to Procedure F, page 4
* Perform result “Look-Back”
* Document problem and corrective action
 | **Procedure G Decontamination** |
| **Archive** | 9 | Attach run-specific report to GAS Wipe Testing Form MB 8.09.F38; place in Service manual |  |

**PROCEDURE E:** Follow the steps in the table below to collect and test RSV ad Influenza (RIP) PCR environmental samples

RIP PCR Environmental Testing

| **Activity** | Step | **Action** | **Related Doc** |
| --- | --- | --- | --- |
| **Worksheet** | 1 | Print RIP Wipe testing worksheet MB 3.02.F5* *Test expanded environmental sites during a contamination event such as drawer handles, light switches, phones, etc.*
 | RIP Wipe Testing MB 3.02.F5 |
|  | 2 | Aliquot 200 µl VTM into a 2 ml cryovial for each sample to be tested |
| **Sample processing** | 3 | Transfer 1 ml NFW into a sterile tube to pre-moisten swabs prior to collection; can be used for multiple swabs* Soak each swab in NFW for 5 s
 |  |
|  | 4 | Swab area by rotating the swab 2 – 3 times across the surface, approx 10 cm2  |  |
|  | 5 | Using an orange barrier protector, break the swab off into labeled sample tube (VTM) |  |
|  | 6 | Vortex 10 seconds, speed 8 |  |
| **PCR** | 7 | Perform PCR testing according to RIP assay protocol  | MB 9.05 RIP Simplexa RSV and Influenza A, B Direct Assay |
| **Notification** | 8 | If results are positive * *Notify* section technical Director and/or designee
* Proceed to Procedure F, page 4
* Perform result “Look-Back”
* Document problem and corrective action
 | **Procedure G Decontamination** |
| **Archive** | 9 | Attach run-specific report to Test Form MB 3.02.F35; place in Service manual |  |

**PROCEDURE F:** Follow the steps in the table below to collect and test HSV 1 and 2 Direct PCR environmental samples

HSV 1 and 2 Direct PCR Environmental Testing

| **Activity** | Step | **Action** | **Related Doc** |
| --- | --- | --- | --- |
| **Worksheet** | 1 | Print HSV Wipe testing worksheet MB 3.02.F6* *Test expanded environmental sites during a contamination event such as drawer handles, light switches, phones, etc.*
 | HSV Wipe Testing MB 3.02.F6 |
|  | 2 | Aliquot 200 µl VTM into a 2 ml cryovial for each sample to be tested |
| **Sample processing** | 3 | Transfer 1 ml NFW into a sterile tube to pre-moisten swabs prior to collection; can be used for multiple swabs* Soak each swab in NFW for 5 s
 |  |
|  | 4 | Swab area by rotating the swab 2 – 3 times across the surface, approx 10 cm2  |  |
|  | 5 | Using an orange barrier protector, break the swab off into labeled sample tube (VTM) |  |
|  | 6 | Vortex 10 seconds, speed 8 |  |
| **PCR** | 7 | Perform PCR testing according to HSV assay protocol  | MB 12.0 Simplexa HSV 1 and 2 Direct Assay |
| **Notification** | 8 | If results are positive * *Notify* section technical Director and/or designee
* Proceed to Procedure F, page 4
* Perform result “Look-Back”
* Document problem and corrective action
 | **Procedure G Decontamination** |
| **Archive** | 9 | Attach run-specific report to Test Form MB 3.02.F6; place in Service manual |  |

**PROCEDURE G:** Follow the steps in the table below for cleaning following contamination

Decontamination procedure following contamination

| **Activity** | **Step** | **Action** | **Related doc** |
| --- | --- | --- | --- |
|  | 1 | Gloves and disposable lab coat required | MB 3.03 Cleaning and Decontamination of Equip and work areas |
| **General** | 2 | Use unidirectional motion when cleaning |
|  | 3 | Change gloves often during decontamination |
|  | 4 | Use 1% Sani-Cloth Bleach wipes on all surfaces or * Use swabs to reach inaccessible areas
* Use disposable Bleach Sani-Cloth for more accessible areas
 | For deeper cleaning, refer to **Procedure H**1% v/v bleach/Alconox solution |
| **Bench-tops, Hoods, Pipettes, and small equipment**Room 2 and 3 | 5 | Wipe bench-tops, BioSafety Hoods, centrifuges, vortex mixers, and pipettes with Sani-Cloth Bleach wipes Allow bleach to dry for 4 – 5 min * Rinse with water followed by 70% alcohol
 | Perform cleaning process twice before repeat wipe testing |
| 6 | Discard in biohazardous waste |  |
| 7 | Change gloves |  |
|  | 8 | Repeat step 5 with fresh Sani-Cloth Bleach wipesAllow bleach to dry for 4 – 5 min * Rinse with water followed by 70% alcohol
* Hoods: Turn on UV light for 15 min
 |  |
|  | 9 | Change gloves and discard disposable lab coat in biohazardous waste |  |
| **Racks, cold blocks** | 10 | Soak racks in 10% bleach for 5 min* Rinse well with water followed by 70% alcohol
* Refer to alternative cleaning solutions, pg. 6, for additional information
 |  |
|  | 11 | Discard disposable materials in hoods and on countertops, i.e., pipette tips, waste containers, pens, etc. |  |
|  | 12 | Repeat wipe testing | **Procedure H** Alternate Cleaning Solutions |
| **Repeat testing** | 13 | If samples continue to be positive for amplicon or genomic DNA/RNA, prepare a 1% v/v bleach/Alconox cleaning solution (Proc. H, page 6) |
|  | 14 | Repeat steps 5-12 |
|  | 15 | Repeat procedure until all environmental swabs test negative |  |

**PROCEDURE H: Alternative Cleaning Solutions**

Freshly prepared 10% bleach has been shown to be extremely effective in destroying DNA contamination. However, there are alternatives available that are less corrosive and are recommended for cleaning instrumentation.

* 1% v/v bleach/Alconox solution (reagents located in dishwashing room)

|  |  |  |
| --- | --- | --- |
| Step | Reagent | Volume |
| 1 | Household bleach (5 – 6 %) | 500 ml |
| 2 | Water | 2000 ml |
| 3 | Alconox | 25 ml |

* ELIMINase – Decon Laboratories
* DNA *AWAY* – Molecular BioProducts, Inc
* DNAZap - Invitrogen Life Science Technologies

**REFERENCES**

1. Pershing, David H., et al, Molecular Microbiology, Diagnostic Principles and Practices, 2nd edition, 2011, ASM Press, Washington, DC, pg. 134 - 135
2. CLSI *Molecular Diagnostic Methods for Infectious Diseases;* Approved Guideline – Second Edition, CLSI document MM3-A2, Wayne, PA, Clinical and Laboratory Standards Institute; 2006
3. CLSI. Establishing Molecular Testing in Clinical Laboratory Environments. Approved Guideline November 2011; CLSI document MM19-A
4. *Good Molecular Practices Guide*, eSensor® Respiratory viral Panel, Clinical Micro Sensors, Inc. dba GenMark Diagnostics, Inc., 5964 La Place Court, Carlsbad, CA 92008, 1-800-373-6767, ww.genmarkdx.com

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| Historical Record |  |
|  | **Version** | **Written/Revised by:** | **Effective Date:** | **Summary of Revisions** |
| 1 | P. Ackerman | 12/20/2006 | Initial Version |
| 1.1 | P. Ackerman | 5/24/08 | Added MRSA wipe testing, added appendix 1 and 2 forms |
| 1.2 | P. Ackerman | 2/4/09 | Modified MRSA procedure to exclude the lysis step; modified appendix 2 form by increasing number of samples to be tested. Added MRSA interpretation table. |
|  | 1.3 | P. Ackerman | 7/1/09 | Modified procedure format |
|  | 5 | P. Ackerman | 7/7/2013 | Reformatted procedure; added proc. C and D, updated proc. F, added alternative cleaning solutions |
|  | 6 | P. Ackerman | 1.8.2014 | Remove SmartCycler information; revised for BD MAX |
|  | 7 | P. Ackerman | 7.8.15 | Added RVP information |
|  | 8 | P. Ackerman | 5.31.16 | Added BORDP Proc. F; formatted for CMS upload; changed logo; added technical director; changed proc. # from MB003.2 to MB 3.02; name change – added Amplicon |
|  | 9 | P. Ackerman | 6.17.17 | Added Proc. A General guidelines, reorganized remaining procedures |
|  | 10 | J. Laramie | 12.01.17 | -Eliminated 4 week monitoring of negative follow-up swabs-Wipe testing frequency for all tests: monthly-Added extraction and IC addition to RVP wipe checks-Added testing for GAS, RIP, CDT-Removed testing of additional targets section (for GAS, RIP, and CDT) |
|  | 11 | J. Laramie | 01.01.17 | -Specified what tubes to break swabs off into for RIP (VTM) and RVP (nuclease free water) wipe testing. |
|  | 11 | J. Laramie | 01.01.17 | Biennial review: JL 03.07.18 |
|  | 12 | J. Laramie | 02.26.19 | Removed MRSA and CDT wipe testing procedures  |
|  | 13 | Susan DeMeyere | 4.24.19 | Added HSV testing |