## Anaerobes

### Clues Suggestive of Anaerobic Infections

- <u>There are many clues that can be suggestive of</u> an anaerobic infection such as:
  - Foul odors
  - Black discoloration of specimens or colonies
  - Inability to grow organisms seen in Gram stain
  - Unique morphologies seen on direct smear Gram stain
    - Fusiform GNB
    - Large spore-forming GPB
    - Branching GPB

Additional characteristics are listed in Table 24-2

## **Specimen Types**

- Anaerobes can be isolated from a number of different specimen types that include:
  - Aspirates
  - Body fluids
  - Lower respiratory
    - BAL and BW only
  - Abscesses
  - Tissues
  - Deep wounds
  - Urine
    - Suprapubic and cystos only
  - Genital sources
    - Intrauterine and vaginal abscesses only

### **Specimen Processing**

#### Primary Media includes:

- CDC-Anaerobe Sheep Blood Agar
- Laked KV
  - Kanamycin to inhibit growth of Gram-negative facultative anaerobic bacilli
  - Vancomycin to inhibit growth of Gram-positive organisms
  - Contains Vitamin K and Hemin to aid in the growth of Bacteroides spp. and Prevotella sp.
- CDC-CNA
  - Colistin and Nalidixic acid: inhibits facultative GNBs

### Anaerobic Conditions

- Anaerobic environments can be generated through various systems that include:
  - Gas-impermeable bags, containers or jars
    - Anaerobic environment is achieved through chemical reactions within a gas pack
  - Anaerobic chamber
    - Environment is maintained by gas-tank delivery system and the use of a palladium catalyst
    - Hydrogen from the gas mixture combines with oxygen to form water in the presence of the catalyst
    - -Oxygen indicator is required

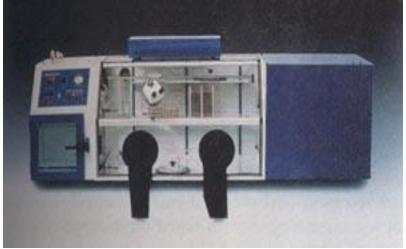
### Anaerobic Culture Environment

Atmosphere:

 Nitrogen: 80-90%
 Hydrogen: 5-10%
 CO<sub>2</sub>: 5-10%

H2 + O2----→H20 palladium catalyst





### **Anaerobic Systems**

#### GasPak Jar Anaerobic Chamber





#### Anaerobic Environment Indicator System

- Indicator systems include:
  - Resazurin
    - **PINK** when O2 is present
    - WHITE/COLORLESS when O2 is not present
  - Other indicator systems
    - Methylene Blue
      - -BLUE when O2 is present



- -**WHITE/COLORLESS** when O2 is not present
- –Up to 2 hours may be required to establish appropriate anaerobic conditions

# Important tests used on the ANA bench

- Aero-tolerant test (radials)
- Gram Stain
- Location of spores if present (terminal, subterminal, central)
- Potency antimicrobial disks (vankomycin,kanamycin,colistin)
- Egg yolk agar (lecithinase and/or lipase)
- Bacteroides Bile Esculin agar (BBE)
- Double zone hemolysis
- Beta lactamase
- Rapid ANA system (4 hrs ID system)

### Gram Stain of Aspirate Material

| Table 28-1. | Characteristics of | anaerobes | based on | Gram stain morphology |
|-------------|--------------------|-----------|----------|-----------------------|
|-------------|--------------------|-----------|----------|-----------------------|

| Organism                    | Gram stain reaction and morphology  |  |  |  |  |
|-----------------------------|---|--|--|--|--|
| Actinomyces spp.            | Branching gram-positive bacilli   |  |  |  |  |
| Clostridium perfringens     | Large gram-positive bacilli with blunt ends ("boxcar-shaped"), no spores                        |  |  |  |  |
| Clostridium tetani          | Gram-positive bacilli with round or oval terminal spore ("drumstick" or "tennis racket" shaped) |  |  |  |  |
| Propionibacterium spp.      | Small, thin pleomorphic gram-positive bacilli   |  |  |  |  |
| Bacteroides, Porphyromonas, | Faintly staining gram-negative coccobacilli   |  |  |  |  |
| or Prevotella spp.          |   |  |  |  |  |
| Fusobacterium nucleatum     | Thin, gram-negative bacilli with tapered ends   |  |  |  |  |
| Fusobacterium necrophorum   | Extremely pleomorphic, thin gram-negative bacilli with bizarre shapes                           |  |  |  |  |
| or F. mortiferium           | Anaerobic gram-positive bacilli V   |  |  |  |  |
| Veillonella spp.            | Tiny gram-negative cocci with a tendency to stain gram variable                                 |  |  |  |  |

#### Color Atlas of Medical Microbiology, ASM Press, p203.

### Gram Stain Pitfall

 Over decolorization can lead to the false impression of a Gram-Negative organism

 Susceptibility to special potency antimicrobial disks can help preliminarily categorize of the anaerobic pathogen

### Potency antimicrobial Disks

- Vancomycin (5 μg)
- Kanamycin (1 mg)
- Colistin (10 µg)
- Gram Positive: Vancomycin S, Colistin R
- Gram Negative: Vancomycin R
  - Exception: *Porphyromonas spp.* Vancomycin S

### Potency Antimicrobial Disks: Presumptive ID

Table 28-3. Presumptive identification of anaerobes based on special-potency antimicrobial disk results

|                                     | Result <sup>a</sup> with disk containing: |                      |                     |  |
|-------------------------------------|---|----------------------|---------------------|--|
| Organism                            | Kanamycin<br>(1,000 μg)                   | Vancomycin<br>(5 μg) | Colistin<br>(10 μg) |  |
| Bacteroides fragilis group          | muld in R floord o                        | R R                  | R                   |  |
| Bacteroides ureolyticus group       | ono ben S dave da                         | R                    | S                   |  |
| Fusobacterium spp.                  | S S S                                     | mosla a R disaré i   | S                   |  |
| Porphyromonas spp.                  | doooco o Ringen du                        | na goune S vitra Mi  | R m                 |  |
| Veillonella spp.                    | S   | R                    | S                   |  |
| Peptostreptococcus anaerobius       | neget de R <sup>S</sup> lood e            | Thin, g 8 m-negative |                     |  |
| Other anaerobic gram-positive cocci | S   | S                    | R                   |  |
| Anaerobic gram-positive bacilli     | V   | S <sup>b</sup>       | R                   |  |

<sup>*a*</sup> R, resistant; S, susceptible; R<sup>S</sup>, resistant, rarely susceptible; V, variable reaction. <sup>*b*</sup> Rare Lactobacillus spp. and Clostridium spp. may be vancomycin resistant.

#### Color Atlas of Medical Microbiology, ASM Press, p204.

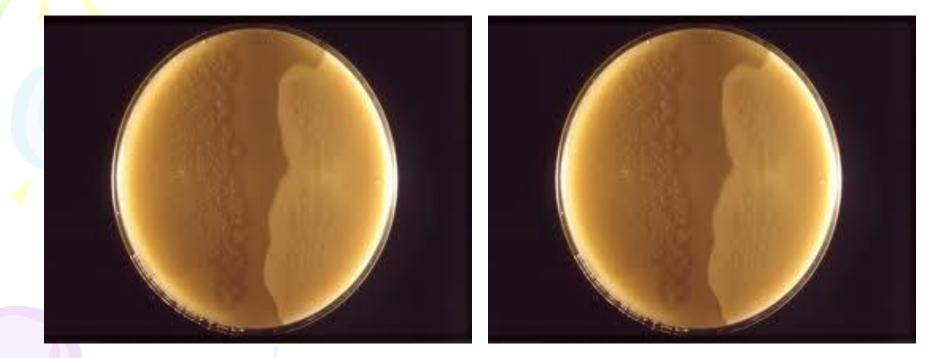
### Lecithinase (Naegler RX)

- Performed on an egg-yolk agar
- Tests the ability to break down lecithine into diglycerides
- Positive for lecithinase: <u>Produces opaque</u> <u>white clearings</u>
- Performed on suspected Clostridium spp.

### Lecithinase (Naegler RX)

#### Negative

#### Positive

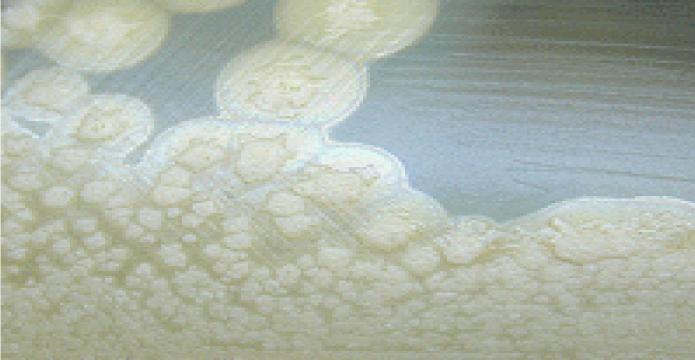


### Lipase

- Performed on an egg-yolk agar
- Tests the ability to <u>break down fat</u> with the enzyme lipase
- Positive: Production of lipase causes an <u>iridescent oil-on-water effect (oil</u> <u>slick)</u>
- Differentiates Fusobacterium spp.



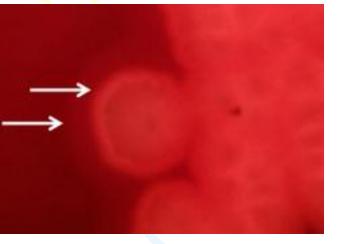
### Lipase (positive)



#### BBE, Double zone hemolysis, Rapid ANA

BBE (Bacteroides spp.)





#### Double Zone Hemolysis (Clostridium perfringens)

#### Rapid ANA (4 hour ID)



## Anaerobic, Spore-Forming Gram Positive Bacilli

### Clostridium spp.

- Obligate anaerobic GPB, form spores
- Ubiquitous in nature (soil)
- Frequent inhabitants of the GI tract
- Frequently isolated species in humans:
  - \*C. perfringens
  - \*C. septicum
  - -C. innocuum
  - -C. ramosum
  - \*C. difficile

## Clostridia spp.

- Wound infections
  - Surgical site
  - Trauma
- Gas Gangrene (*C. perfringens*)
- Abscess (abdominal, lung, organ, brain, oral)
- Septicemia
- Toxin mediated diseases:
  - C. difficile: antibiotic associated diarrhea
  - C. botulinum: botulism; flaccid paralysis
  - *C. tetani*: tetanus; muscle spasm; no relaxation
  - C. perfringens: food poisoning



Figure 28-16 Disk pattern of *Clostridium*. Although some of the clostridia stain gram negative, the disk pattern is consistent with a gram-positive organism and confirms the correct Gram reaction of the isolate. As shown here, clostridia are vancomycin and kanamycin susceptible and colistin resistant. Vancomycin – S Kanamycin – S Colistin - R

Color Atlas of Medical Microbiology, ASM Press, p210.

### **Spore Location**

- Spore location is one of the identifying characteristics used for *Clostridium sp.*
- Location of spores can include:
  - Central
  - Sub-terminal
  - Terminal

### C. botulinum

#### Botulinum toxin - most powerful EXOTOXIN known

- Neurotoxin prevents acetylcholine release from the nerve
   endings producing acute descending, flaccid paralysis
  - Bilateral involvement of Cranial Nerves (face, head, throat), followed by thorax, diaphragm, then extremities
- Death by respiratory failure
- Seven toxin types

#### Food disease = Ingestion botulism

- Improperly preserved canned foods will not
- Spores germinate and secrete botulinum toxin
- Ingestion of the preformed toxin



### C. botulinum

- Neonates ingest bacterial spores
- Typical food source is raw honey
- Germinates in neonatal gut
- Same syndrome as intoxication botulism
- Adults are not susceptible to this type of infection
  - Spores will not germinate in adult gut

Clostridium botulinum
Key characteristics include:
Lipase: positive
Lecithinase: negative
Sub-terminal spore location

\*Diagnosed primarily by:
 —Serological testing
 —Clinical symptoms

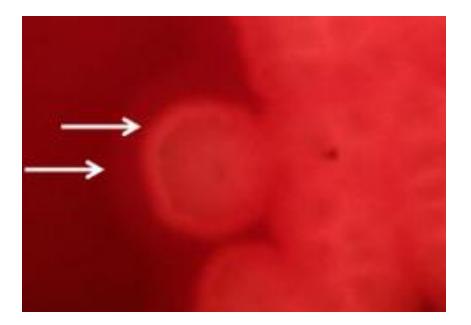


### Clostridium perfringens

- Associated with gas-gangrene/food poisoning
- Gram positive "boxcar" shaped bacilli
- Key characteristics include:
  - Lecithinase: Positive
  - Lipase: Negative
  - Central or sub-terminal spore location(rarely seen)
  - Double zone of Beta-hemolysis
  - Reverse CAMP test positive

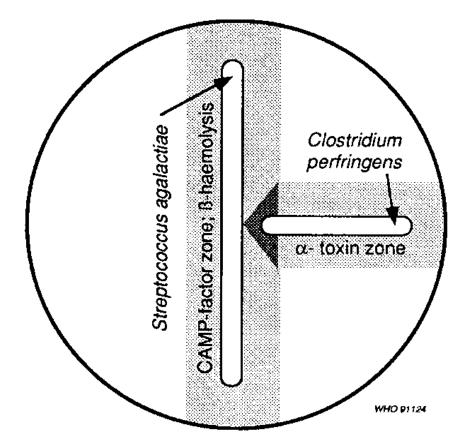
### **Double Beta-Hemolysis**

C. perfringens
 produces a
 characteristic
 double zone of
 Beta-hemolysis



### **Reverse CAMP**

- *C. perfringens* produces a positive reverse CAMP test.
  - Group B is streaked as a line down the center on the plate
  - C. perfringens is streaked in a line perpendicular
  - Arrow head clearing is positive



### **Clostridium difficile**

- Implicated in:
- -Antibiotic-associated diarrhea
- Pseudomembranous colitis
- Key characteristics include:
   Lecithinase negative
   Sub-terminal spore location

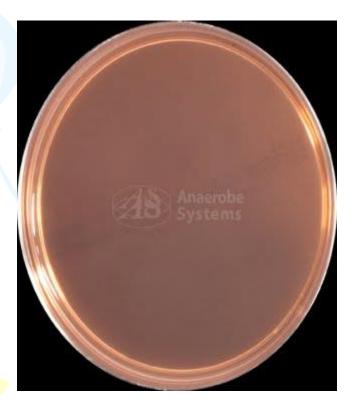


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 Must be a toxin producer to cause disease

#### C. difficile on CCFA

## Cycloserine-cefoxitin-fructose agar (CCFA) is be used to isolate C.difficile <u>Uninoculated</u> <u>Growth</u>





### Clostridium tetani

- Associated with lockjaw and tetanospasims
- Requires a neurological identification
- Key characteristics include:
  - Lecithinase: Negative
  - Lipase: Negative
  - Terminal spore location

#### • Tennis racket shaped cells

- Swarming colonies
- Indole positive(other Clostridia are neg)
- \*Diagnosed primarily by:
- -Serological testing
- -Clinical symptoms

### C. tetani

#### www.textbookofmicrobiology.net





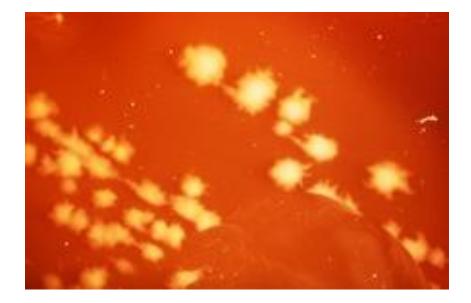
GPB with round/oval terminal spore "tennis racket, drumstick"

### Clostridium septicum

- Associated with malignancy when isolated outside the GI tract
- Key characteristics include:
  - Lecithinase: Negative
  - Lipase: Negative
  - Sub-terminal spore location
  - Swarming colonies
    - Produces "Medusa head" colonies
  - Indole negative

### "Medusa Head" colonies

- C. septicum
   produces fingerlike projections
   from the edge of
   colonies
- Identification on Rapid ANA system



### Anaerobic, Non-spore Forming Gram Positive Bacilli

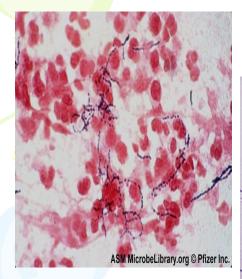
Sensitive to:Vancomycin,Kanamycin Resistant to: Colistin

### Actinomyces sp.

- Branching Gram positive bacilli
- Characteristic white and raised
   Molar Tooth" colonies
- Catalase: Negative
- Indole: Negative
- Associated with lumpy jaw

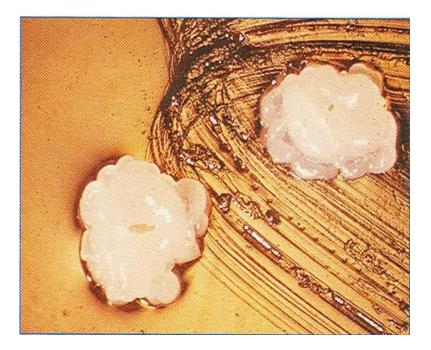


### Actinomyces spp.





Branching, GPB, may have beaded appearance, partial acid-fast negative, whereas *Nocardia spp.* are partial acid-fast positive



### A. israelli. "molar tooth" colonies produced on BHI agar.

Koneman's Color Atlas and Textbook of Diagnostic Microbiology, 6<sup>th</sup> Ed. Color plate 16-2, F

### Propionibacterium sp.

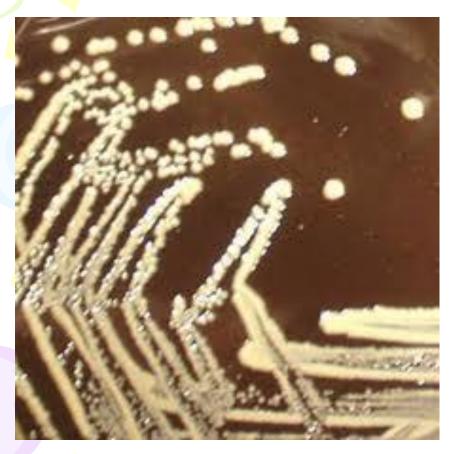
- Pleomorphic Gram positive bacilli
- Catalase positive
- Normal flora in oral cavity, gastrointestinal and urogenital tract and on skin
- Associated with acne, endocarditis, bacteremia, meningitis
- P. acnes: Catalase: Positive Indole: Positive

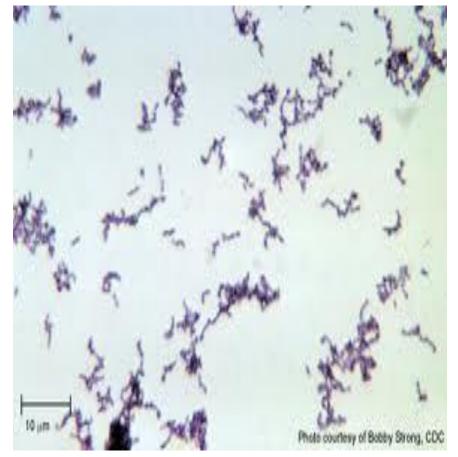


### Propionibacterium sp.

#### **CDC Blood agar**

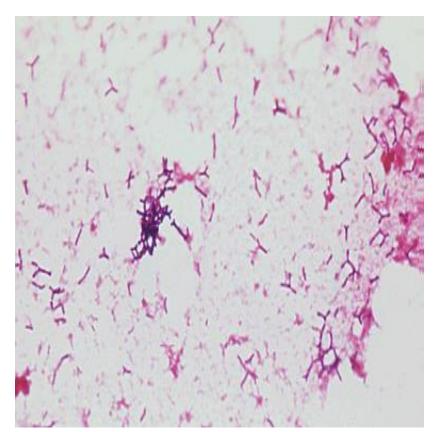
#### **Gram stain**





### Bifidobacterium sp.

- Dog bone or fork shaped gram positive bacilli
- Catalase negative
- Indole negative
- Normal intestinal and oral flora
- Rarely causes disease

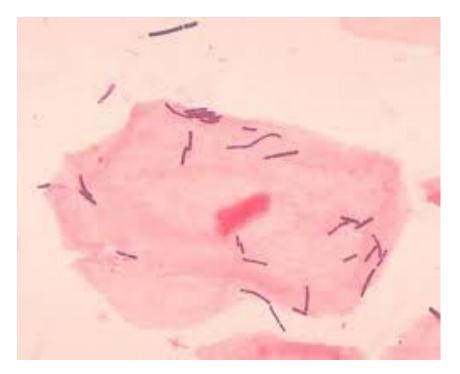


### **Eubacterium species**

- Pleomorphic GPB, may be branching
- Normal intestinal and oral flora
- Catalase : Negative
- Nitrate: Positive
- Not considered a pathogen
- E. lentum is a rare human pathogen

## Lactobacillus sp.

- Long, thin gram positive bacilli
- Catalase negative
- Indole negative
- Normal flora in mouth, intestines and vagina



# Anaerobic, Gram Negative Bacilli

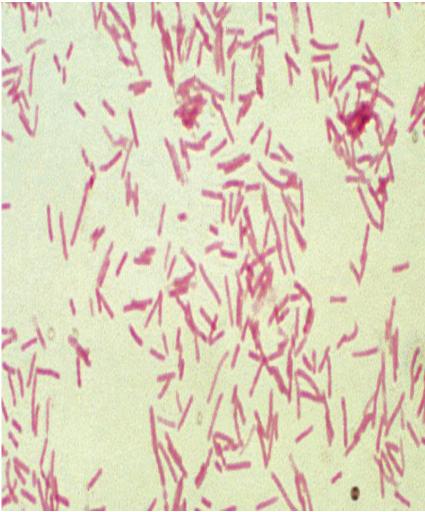
all are normal flora of human mucous membranes

## Bacteroides fragilis Group

- Gram negative bacilli
- Can be found in wounds, tissues and blood cultures
- All species:
  - Resistant to Vancomycin, Kanamycin and Colistin
    - Grows on Laked KV plate
  - Catalase positive
  - Stimulated to grow in 20% bile
    - Bacteroides Bile Esculin (BBE) plate
      - Up to 20% bile concentration
      - Hydrolysis of esculin turns plate black
  - CDC-ANA BAP
    - Grey, speckled colonies

## Bacteroides Bile esculin (BBE) Gram stain





## **Bacteroides fragilis Group**

|                       | Esculin | Salicin | Trehalose | Indole |
|-----------------------|---------|---------|-----------|--------|
| B. fragilis           | +       | -       | -         | -      |
| B. distanosis         | +       | +       | +         | -      |
| B.<br>thetaiotamicron | +       | -       | +         | +      |
| B. vulgatus           | -       | -       | -         | -      |

#### Pigmented Gram Negative Bacilli (dark brown to black colonies)

#### Prevotella sp.

- <u>No growth on BBE</u>
   <u>plate</u>
- Growth on KV plate
  - <u>Resistant</u> to <u>Vancomycin</u> and Kanamycin
  - Variable to Colistin
  - Requires vitamin K and hemim

#### - \*Indole negative

- Red fluorescence under UV light

- Porphyromonas sp.
  - <u>No growth on BBE</u>
     <u>plate</u>
  - <u>No Growth on KV</u>
     <u>plate</u>
    - <u>Susceptible</u> to <u>Vancomycin</u>
    - Resistant to Kanamycin and Colistin
  - \*Indole positive
  - Red fluorescence under UV light

### Fusobacterium sp.

### • Key features include:

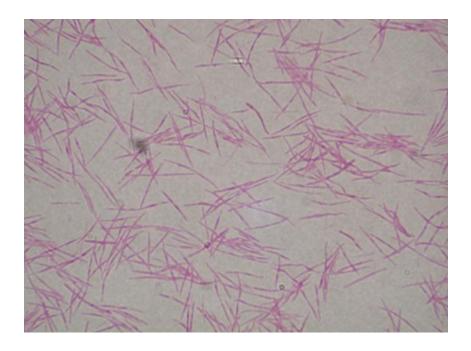
- Resistance to Vancomycin
- Susceptible to Kanamycin and Colistin
- Catalase negative
- Indole positive
- Not stimulated to grow in 20% bile (no growth on BBE)
- Colonies produce a chartreuse fluorescence under long-wave UV light

### Fusobacterium sp.

- F. nucleatum
  - Long, slender, fusiform gram negative bacilli
  - -\*Lipase negative
- F. necrophorum
  - Pleomorphic gram negative bacilli or coccobacilli
  - -\*Lipase positive
- Key features can be found in Table
   24-8

### Fusobacterium sp.

 F. nucleatum
 produces a characteristic fusiform shape on gram stain



# Anaerobic, Gram Positive Cocci

### Peptostreptococcus sp.

- Susceptible to Vancomycin
- Resistant to Kanamycin and Colistin
- Catalase negative
- Most common:
- ➢P. anaerobius: <u>Indole pos</u>,
  - SPS sensitive
- P. asaccharolyticus: <u>Indole neg</u>, SPS resistant

# Anaerobic, Gram Negative Cocci

### Veillonella sp.

- Non-pathogenic normal flora
- Resistant to Vancomycin and susceptible to Kanamycin and Colistin
- Red fluorescence under UV light

### Question 1

All of the following specimen types are acceptable specimens for processing anaerobic organisms except:

- A. Aspirates
- B. Clean catch urine
- C. Body fluids
- D. Lower respiratory

### Question 2

Describe some of the clues that would indicate the possibility of an anaerobic infection?

### Question 3

A 36 year old man arrived at the emergency room after noticing a black discoloration in his toe 5 days prior to his visit. Since that time, the discoloration has spread to his lower limb. The attending physician noted severe tissue necrosis and a build-up of gas under the skin and sent a specimen to the lab for examination.

- 1. Do you think this patient possibly has an anaerobic infection? Why?
- 2. List some of the laboratory tests that might be ordered for this patient?
- 3. Describe how this organism might appear on culture media and on a Gram stain?
- 4. What organism do you think is causing this patients symptoms?