

Application Sheet



Laboratory Name
Test Name: **Bicarbonate Liquid**

System information

For **cobas c** 311/501 analyzers:

CO2-L: ACN 156

SCO2L: ACN 763 (STAT, reaction time: 5) (for **cobas c** 501 analyzer)

SCO2L: ACN 763 (STAT, reaction time: 4) (for **cobas c** 311 analyzer)

For **cobas c** 502 analyzer:

CO2-L: ACN 8156

SCO2L: ACN 8763 (STAT, reaction time: 5)

Intended use

In vitro test for the quantitative determination of bicarbonate (HCO_3^-) in human serum and plasma on Roche/Hitachi **cobas c** systems.

Summary

Bicarbonate is the second largest fraction of the anions in plasma. Included in this fraction are the bicarbonate (HCO_3^-) and carbonate (CO_3^{2-}) ions, as well as the carbamino compounds. At the physiological pH of blood, the concentration of carbonate is 1/1000 that of bicarbonate. The carbamino compounds are also present in such low quantities that they are generally not mentioned specifically. Several different methods for the determination of bicarbonate in serum and plasma have been reported. Most of these procedures utilize acidification of the sample and conversion of all carbon dioxide forms to CO_2 gas.¹ The amount of gas formed is measured by manometric or volumetric devices, ion selective electrodes, or spectrophotometric techniques.^{2,3} These methods are either cumbersome, time-consuming, technique-oriented, and/or require special equipment.

Enzymatic procedures using phosphoenolpyruvate carboxylase (PEPC) have been described.^{4,5}

The bicarbonate content of serum or plasma is a significant indicator of electrolyte dispersion and anion deficit. Together with pH determination, bicarbonate measurements are used in the diagnosis and treatment of numerous potentially serious disorders associated with acid-base imbalance in the respiratory and metabolic systems.

Test principle

Bicarbonate reacts with phosphoenolpyruvate (PEP) in the presence of PEPC to produce oxaloacetate and phosphate:



The above reaction is coupled with one involving the transfer of a hydrogen ion from NADH analog to oxaloacetate using MDH.



The resultant consumption of NADH analog causes a decrease in absorbance, which is proportional to the concentration of bicarbonate in the sample being assayed.

Reagents - working solutions

R1 Phosphoenolpyruvate: ≥ 40 mmol/L; NADH analog: ≥ 2 mmol/L; MDH (porcine): ≥ 314.3 $\mu\text{kat/L}$; PEPC (microbial): ≥ 30.8 $\mu\text{kat/L}$

R1 is in position B.

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Precautions and warnings

For in vitro diagnostic use.
Exercise the normal precautions required for handling all laboratory reagents.
Disposal of all waste material should be in accordance with local guidelines.
Safety data sheet available for professional user on request.
For USA: For prescription use only.

Reagent handling

Ready for use

Storage and stability

CO2-L
Shelf life at 2-8 °C: See expiration date on **cobas c** pack label.
On-board in use and refrigerated on the analyzer: 6 weeks

Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.
Only the specimens listed below were tested and found acceptable.

Serum

Plasma: Li-heparin plasma

The preferred specimen is from venous blood collected anaerobically in the usual manner for bicarbonate analysis. Bicarbonate content in uncapped tubes decreases approximately 4 mmol/L after one hour.⁶ It has been reported that alkalinized serum stored in open cups is stable for up to 4 hours.⁶ Storage of serum at -20 °C or -80 °C for up to 6 months had no significant effect.⁷

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay.

Stability: 7 days at 4-8 °C⁸
40 hours at 15-25 °C^{9,10}

Separate from erythrocytes and store tightly stoppered.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

See "Order information" section

General laboratory equipment

In addition, other suitable control material can be used.

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions. The performance of applications not validated by Roche is not warranted and must be defined by the user.

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Application for serum and plasma

cobas c 501 test definition

Assay type	2-Point Rate		
Reaction time / Assay points	10 / 4-29 (STAT 5 / 4-29)		
Wavelength (sub/main)	505/415 nm		
Reaction direction	Decrease		
Unit	mmol/L		
Reagent pipetting		Diluent (H ₂ O)	
R1	50 µL	130 µL	
R2	–	–	
<i>Sample volumes</i>	<i>Sample</i>	<i>Sample dilution</i>	
		<i>Sample</i>	<i>Diluent (H₂O)</i>
Normal	2 µL	–	–
Decreased	2 µL	–	–
Increased	2 µL	–	–

Calibration

Calibrators	S1: H ₂ O S2: Ammonia/Ethanol /CO ₂ Calibrator
Calibration mode	Linear
Calibration frequency	2-point calibration - after reagent lot change - as required following quality control procedures

Traceability: This method has been standardized against a primary standard.

Quality control

At least once daily run solutions at two levels of a quality control material with known concentrations.
Refer to Brown Clinic Quality Control Requirements, Rules and Reviews Policy
Refer to Brown Clinic Quatliy Control Specialty and Subspecialty Policy

Calculation

Roche/Hitachi **cobas c** systems automatically calculate the analyte concentration of each sample.

Limitations – interference

Criterion: Recovery within $\pm 10\%$ of initial value at a bicarbonate concentration of 22 mmol/L.
Icterus:¹¹ No significant interference up to an I index of 60 for conjugated and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 1026 µmol/L or 60 mg/dL).
Hemolysis:¹¹ No significant interference up to an H index of 600 (approximate hemoglobin concentration: 372.6 µmol/L or 600 mg/dL).
Lipemia (Intralipid):¹¹ No significant interference up to an L index of 1800. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.
Drugs: No interference was found at therapeutic concentrations using common drug panels.^{12,13}

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In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.¹⁴

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

An abnormally elevated concentration of ambient carbon dioxide (CO₂) may occur under certain environmental conditions in the laboratory. The fluctuating ambient CO₂ concentration may interfere with the CO₂-L assay leading to higher CO₂ results. Under these circumstances, the reduction of the re-calibration interval may become necessary if the laboratory is unable to keep the ambient CO₂ concentration at a normal level by appropriate countermeasures.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on Roche/Hitachi **cobas c** systems. The latest version of the carry-over evasion list can be found with the NaOHD-SMS-SmpCln1+2-SCCS Method Sheets. For further instructions refer to the operator's manual. **cobas c** 502 analyzer: All special wash programming necessary for avoiding carry-over is available via the **cobas** link, manual input is not required.

Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.

Limits and ranges

Measuring range

2-50 mmol/L

Lower limits of measurement

Lower detection limit of the test:

2 mmol/L

The lower detection limit represents the lowest measurable analyte level that can be distinguished from zero. It is calculated as the value lying 3 standard deviations above that of the lowest standard (standard 1 + 3 SD, repeatability, n = 21).

Expected values¹

22-29 mmol/L

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

For Known Interfering Substances section refer to package insert.

For Known Non-Interfering Substance refer to package insert.

For Additional Technical Information refer to package insert.

References

- 1 Scott MG, Heusel JW, LeGrys VA, et al. Electrolytes and blood gases, in Tietz NW. Textbook of Clinical Chemistry. 3rd ed. Philadelphia, PA: WB Saunders Co 1999;1065-1066.
- 2 Natelson S. Microtechniques of Clinical Chemistry. Springfield, IL: Charles C Thomas 1975;147.
- 3 Segal MA. A rapid electrotitrimetric method for determining CO₂ combining power in plasma or serum. Am J Clin Pathol 1955;25:1212-1216.
- 4 Wilson W, Jesyk P, Rand R, et al. Use of Vickers discrete analyzer for enzymatic determination of the bicarbonate content of serum. Clin Chem 1973;19(6):640.
- 5 Norris KA, Atkinson AR, Smith WG, et al. Colorimetric enzymatic determination of serum total carbon dioxide, as applied to the Vickers Multichannel 300 discrete analyzer. Clin Chem 1975;21:1093-1101.
- 6 Gambino SR, Schreiber H. The measurement of CO₂ content with the autoanalyzer. A comparison with 3 standard methods and a description of a new method (alkalinization) for preventing loss of CO₂

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- 7 Elfath D, Cooney J, McDaniel R, et al. Effect of frozen storage of serum on the level of 22 chemistry analytes. Clin Chem 1991;37:931.
 - 8 Guder WG, da Fonseca-Wollheim F, Heil W, et al. Quality of Diagnostic Samples, in brochure: Recommendations of the Working Group on Preanalytical Quality of the German Society for Clinical Chemistry and Laboratory Medicine, 3rd Ed. 2010.
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 - 15 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

Alternative method

Refer to Brown Clinic Back-up Testing Policy

Source document

Reagent Name: CO2-L
Method Sheet Version: V11.0 English

Order information

REF	CONTENT	System-ID	Analyzer(s) on which cobas c pack(s) can be used
03289923 190	Bicarbonate Liquid (250 tests)	07 6725 5	Roche/Hitachi cobas c 311, cobas c 501/502
20751995 190	Ammonia/Ethanol/CO2 Calibrator (2 x 4 mL)	Code 688	
20752401 190	Ammonia/Ethanol/CO2 Control Normal (5 x 4 mL)	Code 100	
20753009 190	Ammonia/Ethanol/CO2 Control Abnormal (5 x 4 mL)	Code 101	
12149435 122	Precinorm U plus (10 x 3 mL)	Code 300	
12149435 160	Precinorm U plus (10 x 3 mL, for USA)	Code 300	
12149443 122	Precipath U plus (10 x 3 mL)	Code 301	
12149443 160	Precipath U plus (10 x 3 mL, for USA)	Code 301	

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Effective date

Effective date for this procedure:

Author

Source documentation compiled by Roche Diagnostics

Revised by: Heather J Hall, MBA, MT(ASCP), CG(ASCP)^{cm} Date: 4/9/2018

Approved by: _____ Date: _____

REVIEW – REVISION SUMMARY DOCUMENTATION

Date: _____ By: _____ Revision Summary: _____
