

# The Wonderful World of Burkholderia

## A Review

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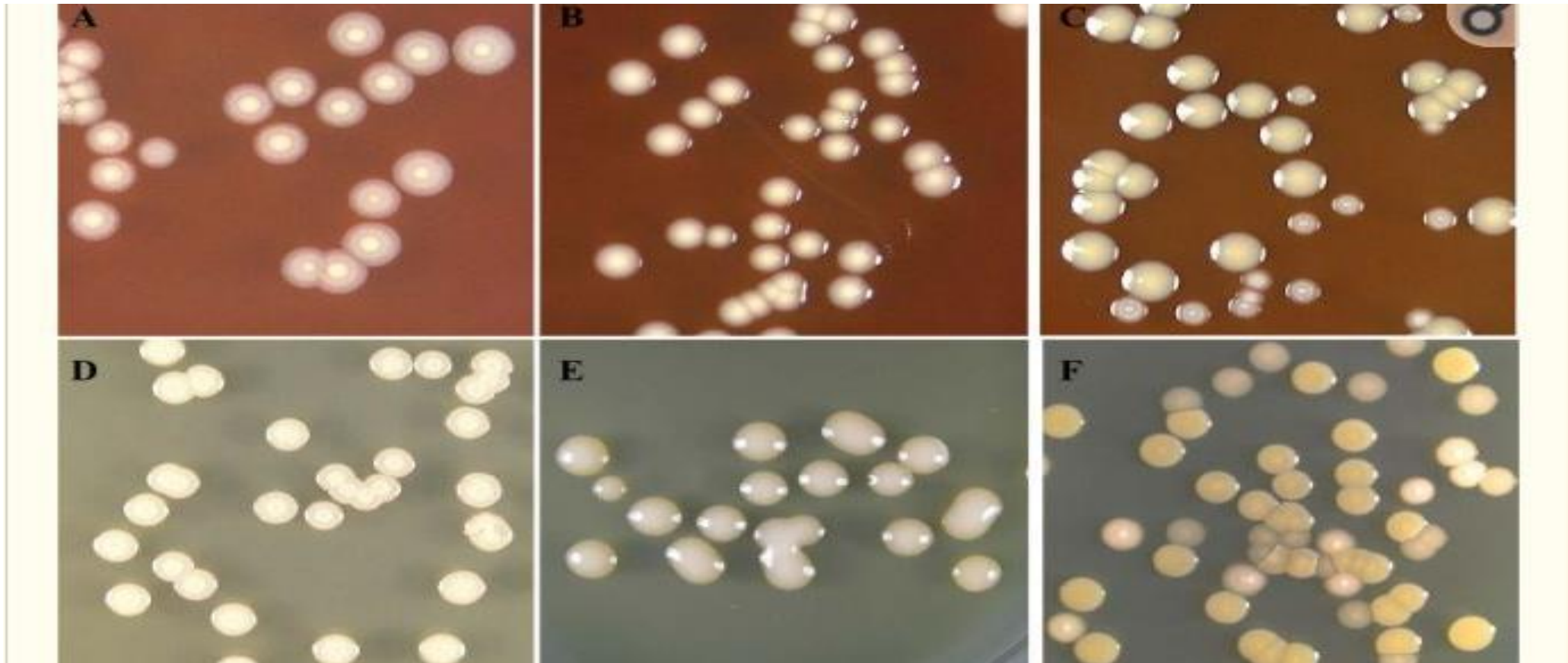
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9/12/2020

# Colony Morphology Can Vary



BAP

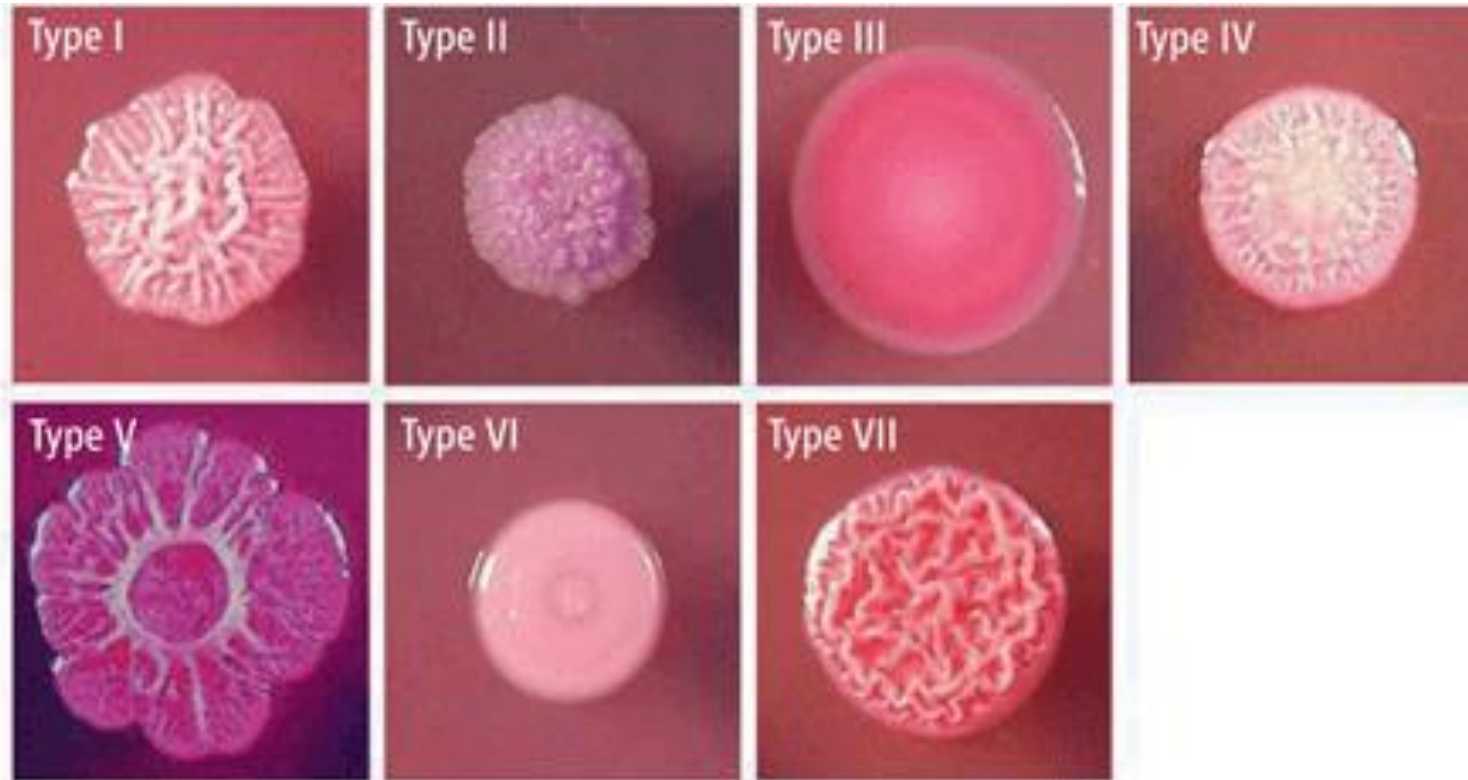
TSA

A & D = Non-mucoid colonies

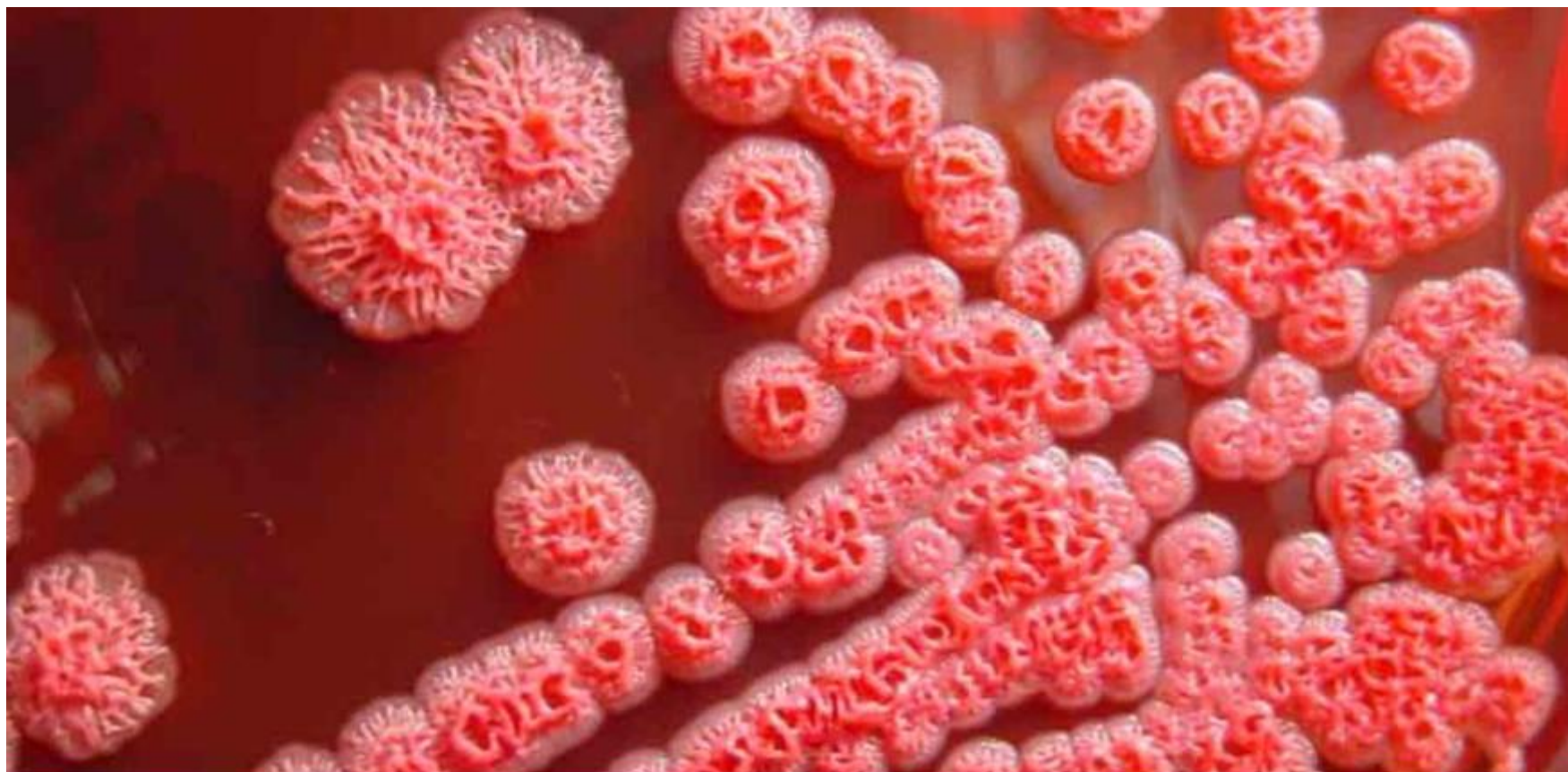
B & E = Mucoid colonies

C & F = Mixed Morphologies

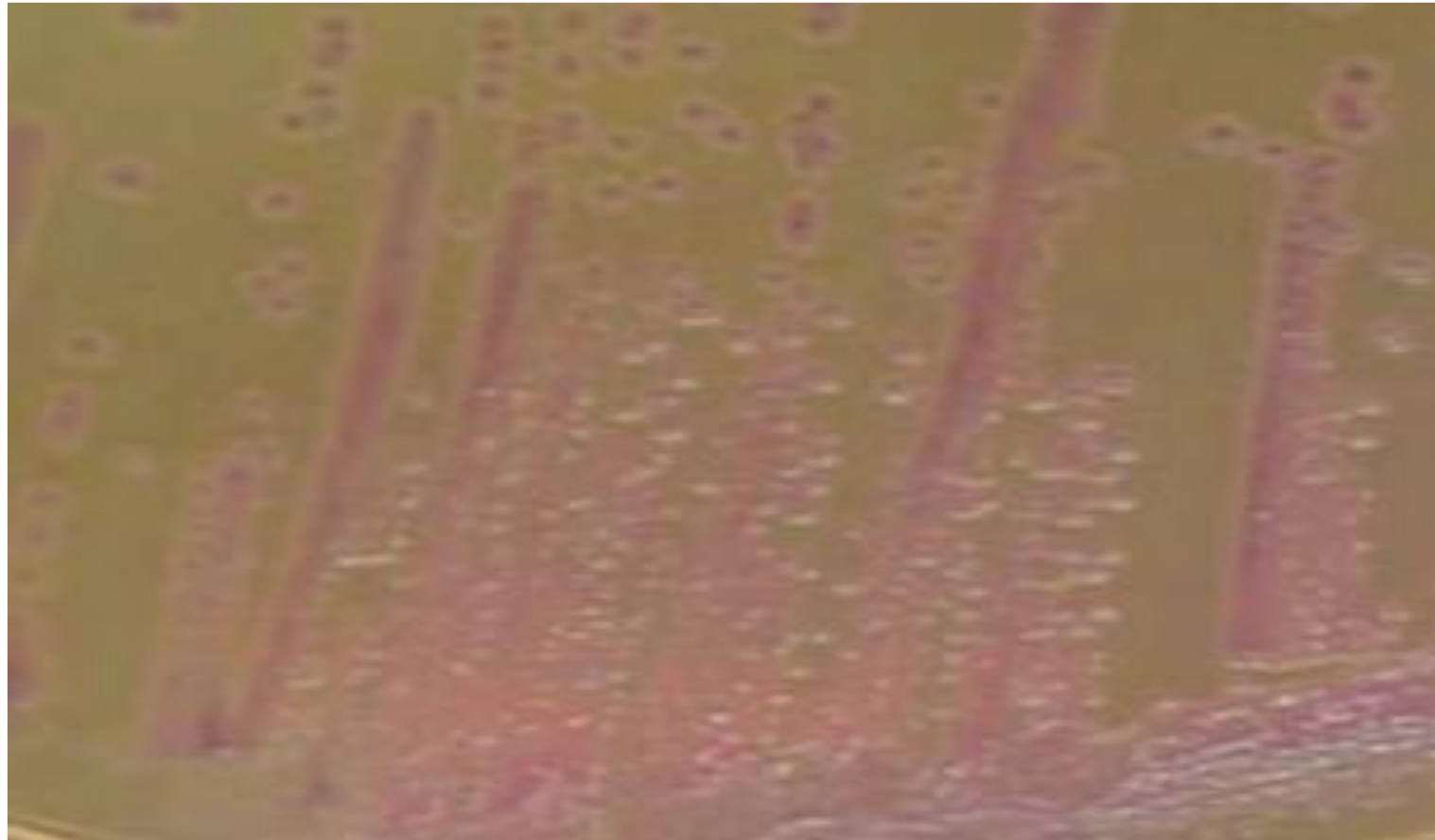
# Colony Morphology Can Vary



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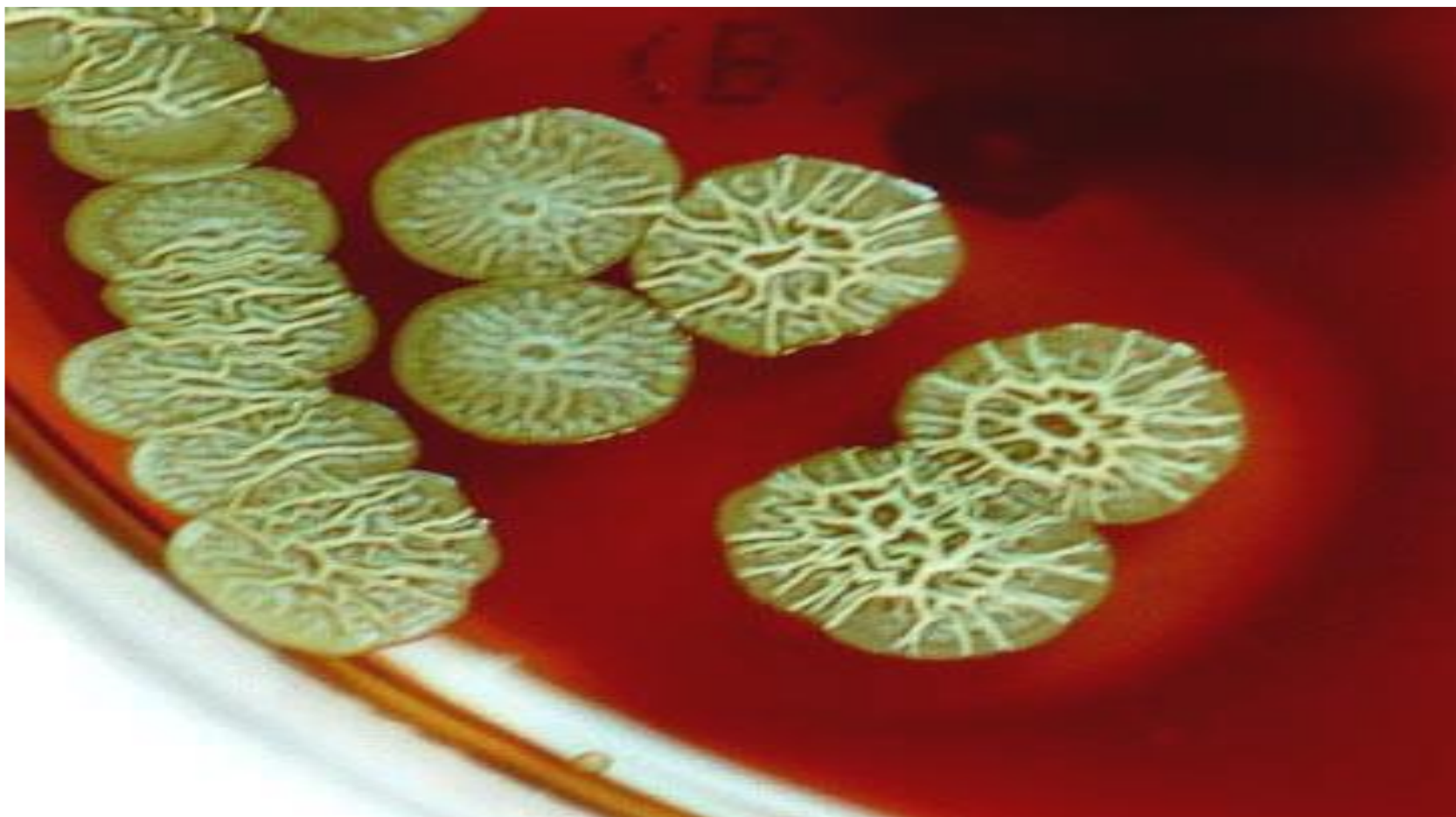
# Colony Morphology Can Vary



# Colony Morphology Can Vary



# Colony Morphology Can Vary





# Colony Morphology Can Vary



# The colonies are gorgeous, but...

Where do I begin with the identification?

# Key Tests

- Gram Stain
- Growth on MacConkey Agar
- Growth at 42°C Catalase
- Oxidase
- Indole
- Polymyxin B
- Motility (Use semi-solid agar only)
- Triple Sugar Iron Agar – *use only to determine glucose non-fermenter status*

# Burkholderia species - TNTC

*Are there really  
over 100 species of  
Burkholderia????*



# Burkholderia cepacia Complex

Tests	Burkholderia ambifaria	Burkholderia anthina	Burkholderia cenocepacia	Burkholderia cepacia	Burkholderia dolosa	Burkholderia multivorans	Burkholderia pyrrocinia	Burkholderia stabilis	Burkholderia vietnamiensis
Urea hydrolysis	-	-	✓	✓	+	+	-	-	✓
Growth at 42 degrees C	✓	✓	✓	✓	+	+	✓	-	-
Citrate	+	+	+	+	+	+	-	+	+
Lysine decarboxylase	+	+	+	+	-	✓	+	+	+
Ornithine decarboxylase	-	-	✓	✓	-	✓	✓	+	-
Adonitol	✓	✓	✓	✓	+	+	+	✓	-
ONPG (beta galactosidase)	+	✓	✓	✓	+	✓	+	-	✓
Sucrose	+	✓	✓	✓	-	-	✓	-	✓
Lactose	✓	✓	✓	✓	+	+	-	✓	✓
Trehalose			✓	✓	+	+	-	✓	
Gelatin hydrolysis	+	-	✓	✓	✓	✓	✓	✓	✓
D-Mannitol	+	+	+	+	+	+	✓	+	+
D-Mannose	+	+	+	+	+	+	✓	+	+
Growth on MacConkey agar	+	+	✓	✓	+	+	✓	+	+
L-Arabinose	+	+	✓	+	+	+	✓	✓	+
Nitrite to gas			-	-	-	-	✓	-	-
Maltose	+	+	✓	✓	+	+	+	✓	✓
Oxidase	+	+	✓	✓	✓	✓	+	+	+
Yellow pigment	-	-	✓	✓	✓	✓	-	-	-
Motile			✓	✓	+	+	+	+	+
Beta hemolysis	✓	✓	✓	✓	-	-	✓	-	✓
D-Xylose	+	✓	✓	✓	+	+	✓	✓	✓
Esculin hydrolysis	✓	-	✓	✓	✓	✓	-	✓	-
Raffinose			✓	✓	-	-	-	✓	✓
Casein hydrolysis			✓	✓	-	-		✓	-
Nitrate to nitrite	✓	✓	✓	✓	+	+	✓	✓	✓
Cellobiose			✓	✓	+	+	✓	✓	✓
Salicin			✓	✓	-	✓	✓	✓	-
Dulcitol			✓	✓	+	+		✓	✓
D-Sorbitol			✓	✓	+	+		✓	✓
myo-Inositol			✓	✓	+	+		✓	✓
Glycerol			✓		+	+		✓	✓

+ > 90%  
 ✓ 10% - 90%  
 - < 10%

# Other Important Burkholderia species

Tests	Burkholderia gladioli	Burkholderia mallei	Burkholderia pseudomallei	Burkholderia thailandensis
Nitrite to gas	-	-	+	+
Growth at 42 degrees C	-	-	+	+
Yellow pigment	+	-	+	+
Tartrate utilization	+	-	-	-
Motile	+	-	+	+
Starch hydrolysis	-	∇	+	+
Maltose	-	∇	+	+
Adonitol	+		-	+
Pyrrolidonyl-beta-naphthylamide	+		-	-
Dulcitol	+		+	-
Erthryritol	-		+	-
Arginine dihydrolase	-	+	∇	∇
Citrate	+	-	∇	∇
L-Arabinose			-	+
D-Mannitol	+	∇	+	+
Beta hemolysis	∇	-	-	-
Lipase	+	∇	+	+
Glucose oxidizer	∇	+	+	+
D-Xylose	+	∇	+	+
Nitrate to nitrite	∇	+	+	+
Growth on MacConkey agar	+	∇	+	+
Sucrose	-	-	∇	∇
Oxidase	∇	∇	+	+
Lactose	∇	∇	+	+
ONPG (beta galactosidase)	∇		-	-
Gelatin hydrolysis	∇	-	∇	∇
Esculin hydrolysis	∇	-	∇	∇
L-Rhamnose	-		∇	∇

+ > 90%  
 ∇ 10% - 90%  
 - < 10%

# Biothreat Agents:

1. *Burkholderia mallei*
2. *Burkholderia pseudomallei*

Two Choices: **RULE OUT OR SEND OUT**

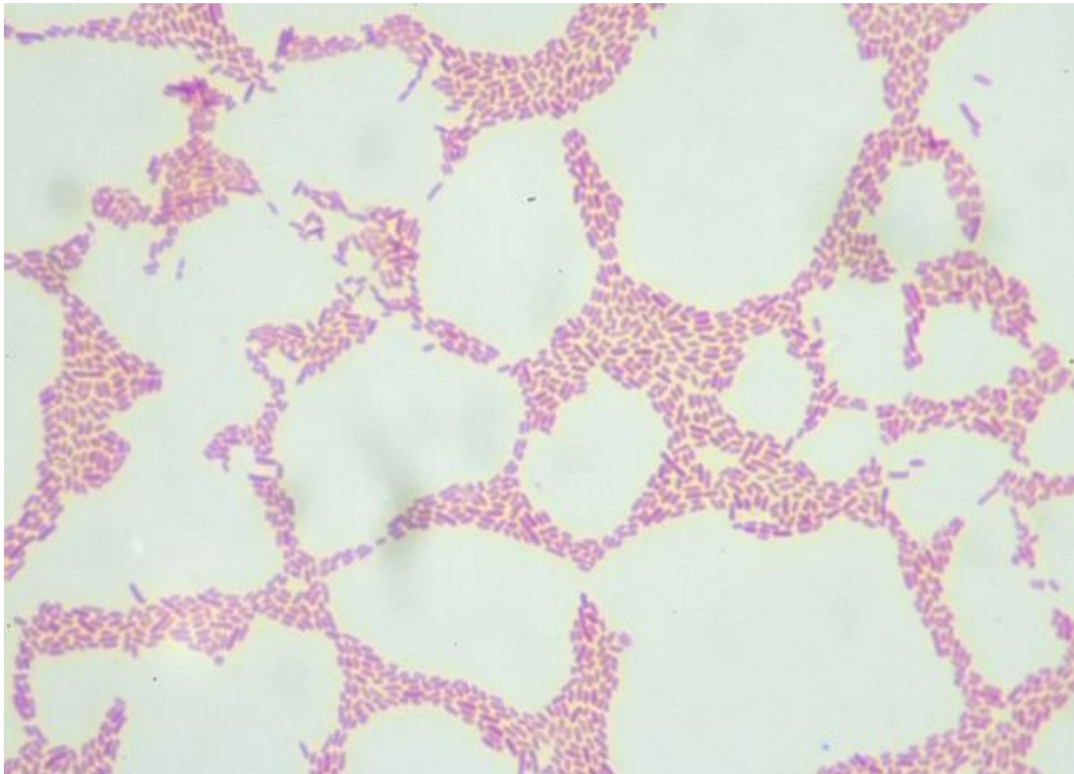
*If unable to rule out, send the isolate to the  
Maryland State Public Health laboratory.*

**DO NOT SEND TO A COMMERCIAL REFERENCE LABORATORY.**

# Gram Stain

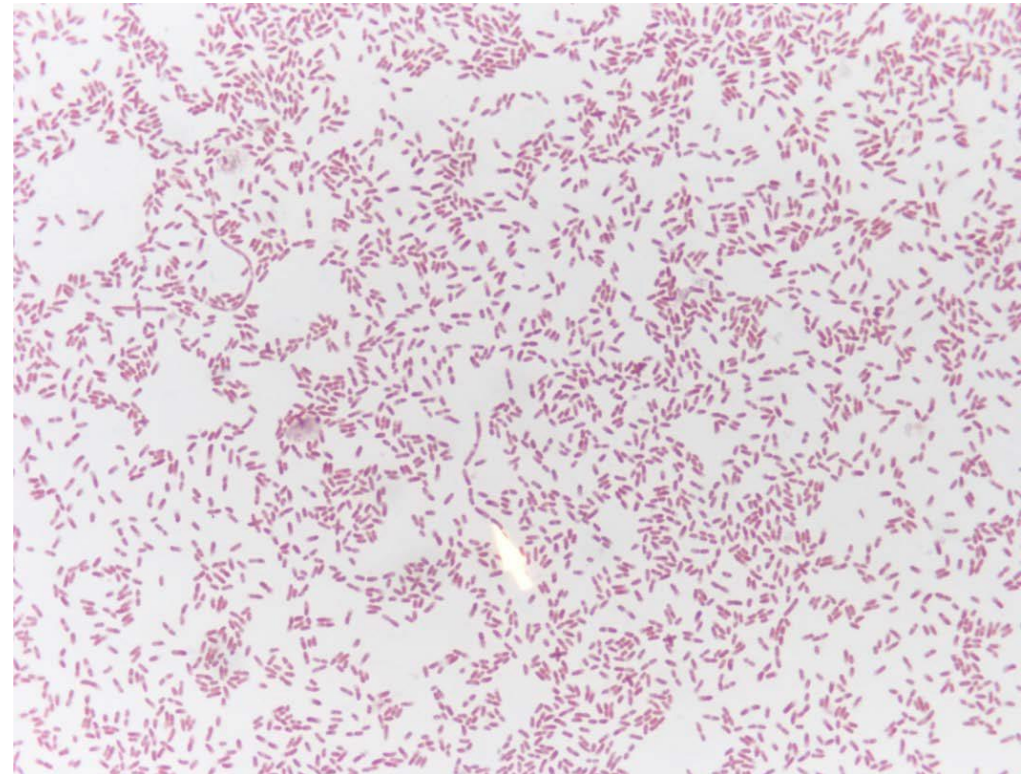
## **Burkholderia mallei**

*Gram negative coccobacilli with round ends, Chinese-letter forms in parallel or bundles*



## **Burkholderia pseudomallei**

*May appear bipolar in direct smears*





# Key Characteristics

## **Burkholderia mallei**

- Blood Agar = Nonhemolytic/No pigment
- MacConkey = Poor/No growth
- Growth at 42°C = No growth
- Catalase = Positive
- Oxidase = Variable/Mostly negative
- Spot Indole = Negative
- Polymyxin B = No zone
- Motility = Negative

## **Burkholderia pseudomallei**

- Blood Agar = Nonhemolytic/**Pigment**
- MacConkey = Growth
- Growth at 42°C = **Growth**
- Catalase = Positive
- Oxidase = Positive (may be delayed)
- Spot Indole = Negative
- Polymyxin B = No zone
- Motility = **Positive**
- **Diagnostic = Distinctive musty/earthy odor**

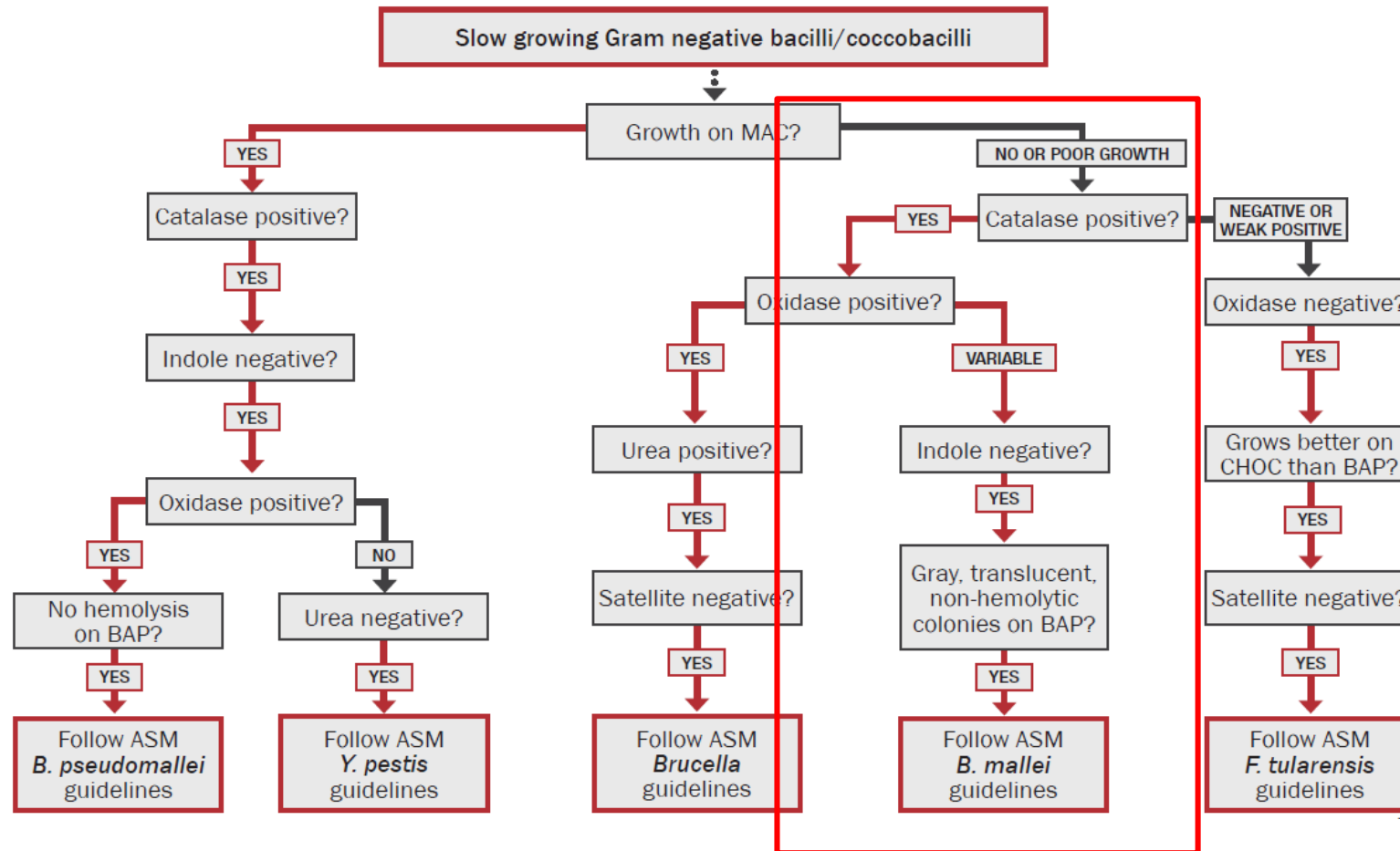
# Flowcharts – Rule In/Rule Out

The gateway to tables and the Laboratory Response Network (LRN) Facilities

# *Burkholderia mallei*

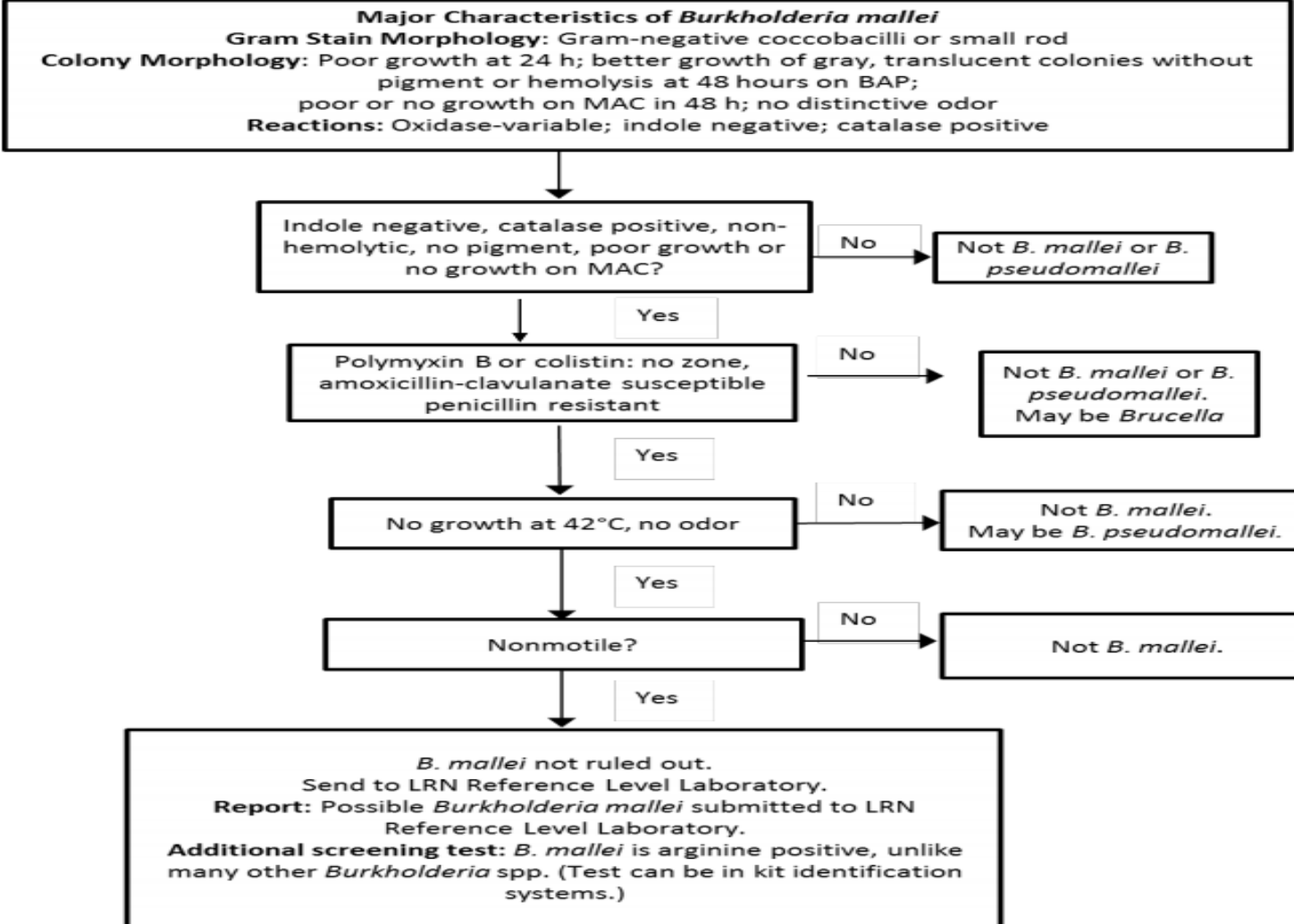
Oxidase **variable**; Indole negative; Catalase positive

# Go With The **APHL** Flow – *B. mallei*

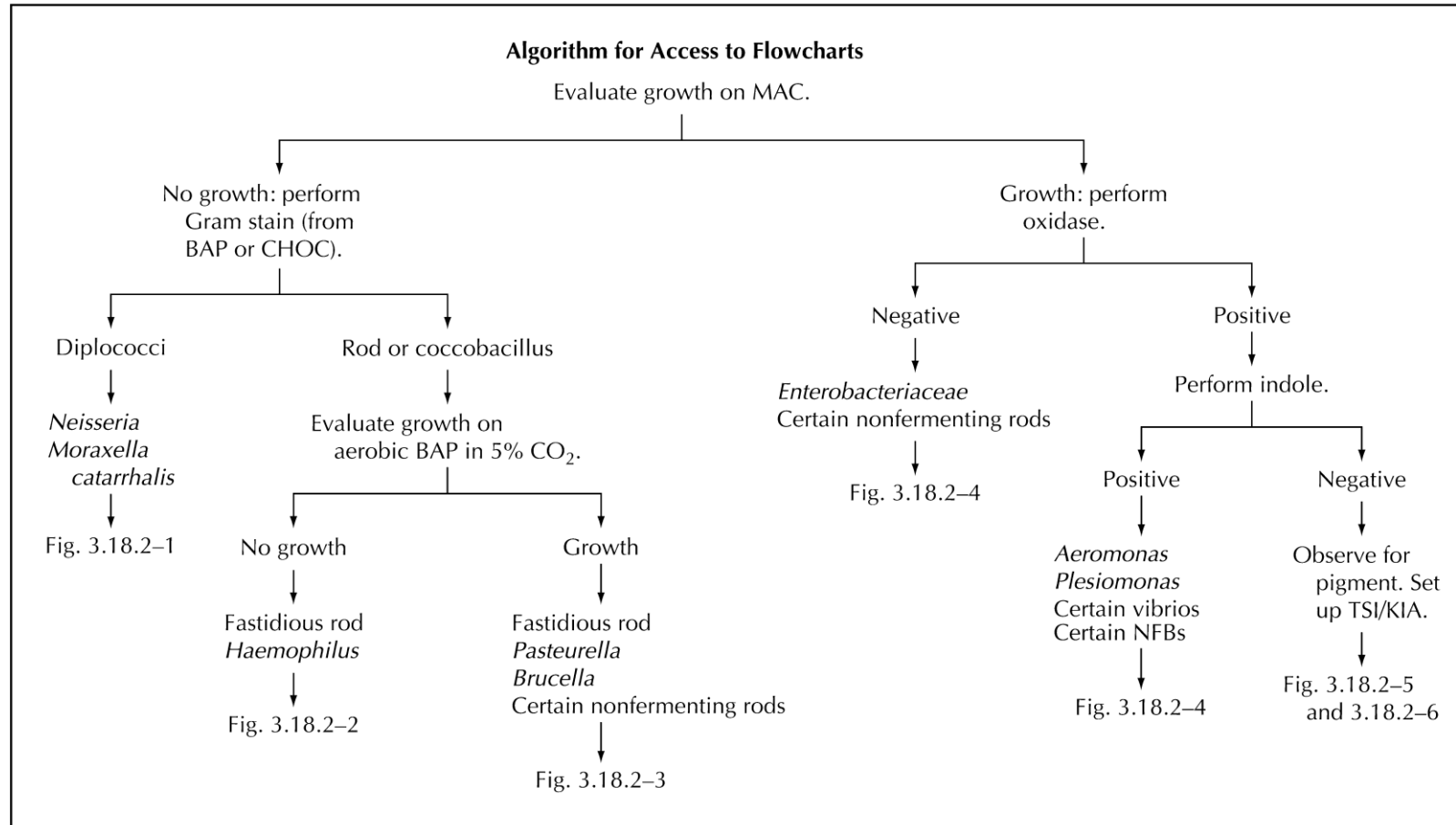


# Go With The **ASM Sentinel Lab** Flow – *B. mallei*

## ***Burkholderia mallei* Identification Flowchart**

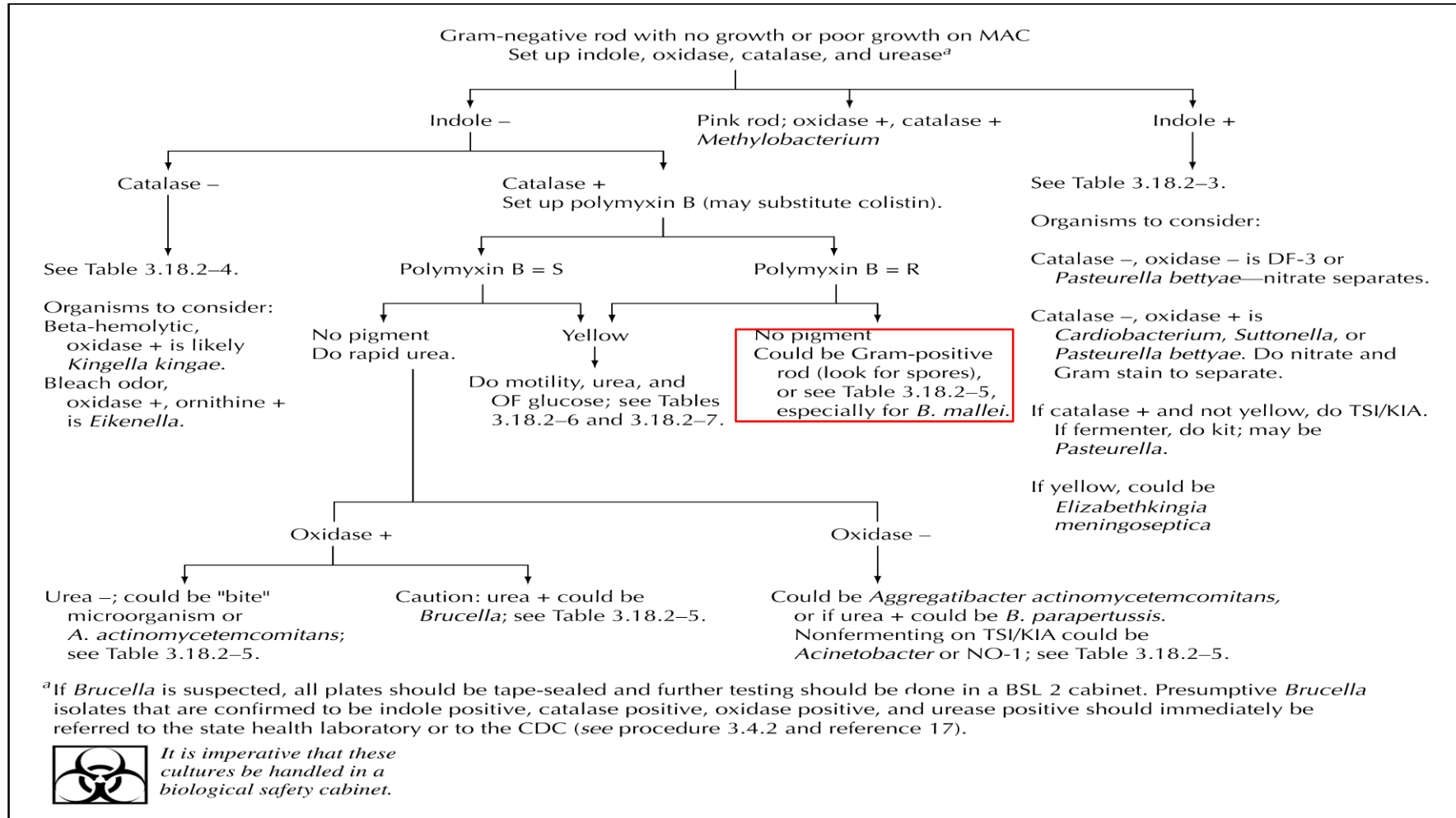


# Go With The **ASM CMPH** Flow – *B. mallei*



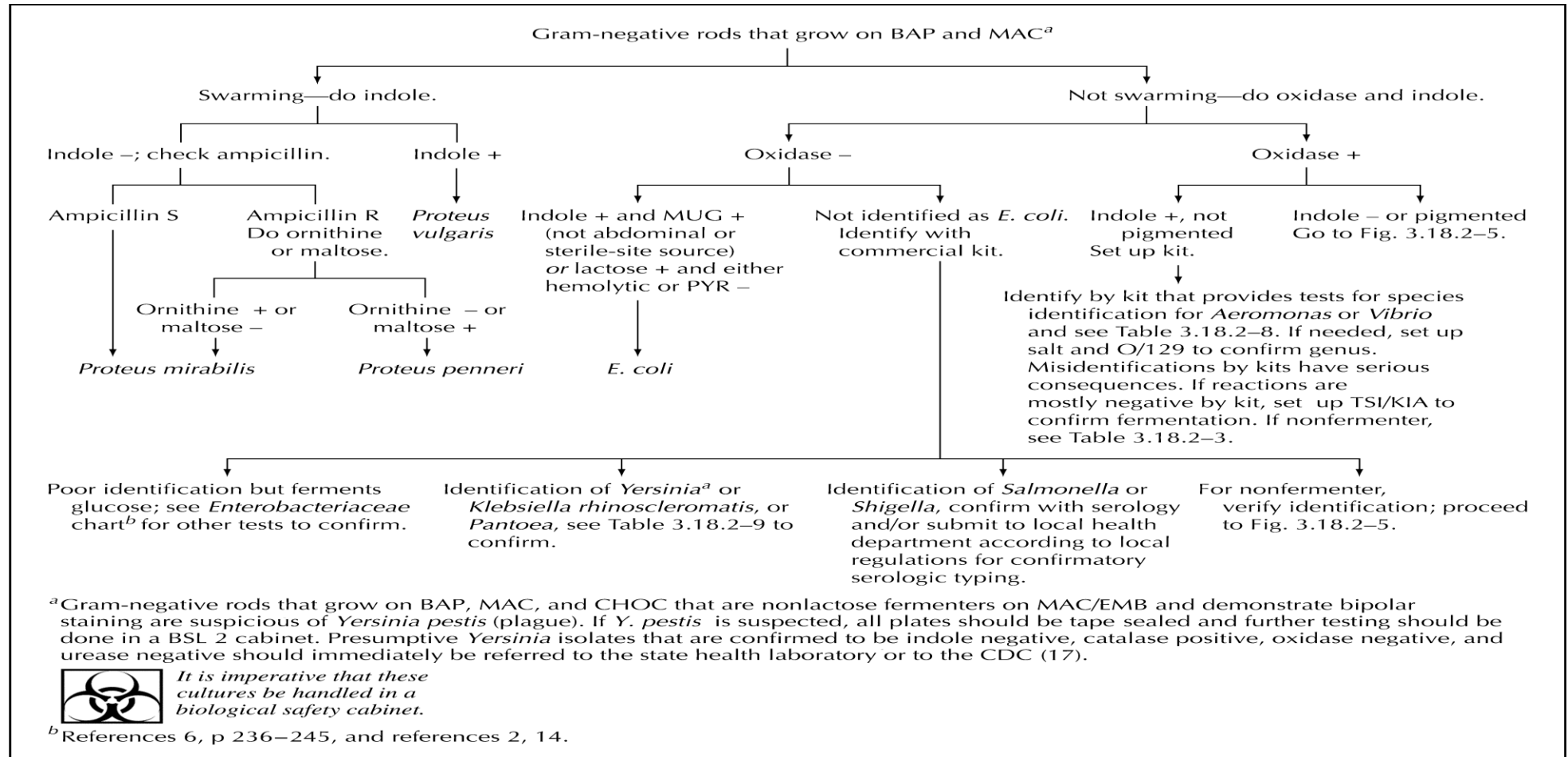
# Go With The ASM CMPH Flow – *B. mallei*

Figure 2-3



# Go With The ASM CMPH Flow – *B. mallei*

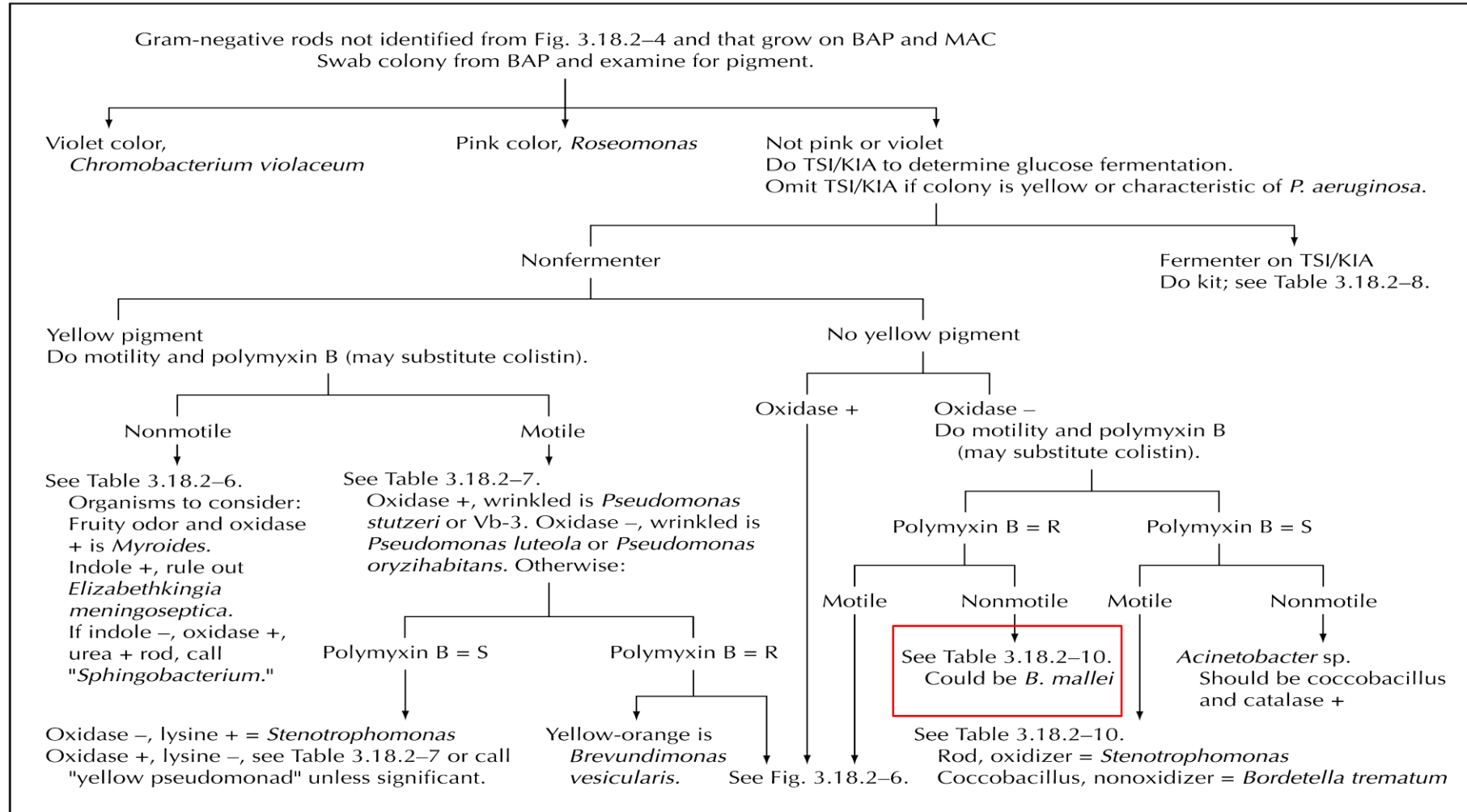
Figure 2-4





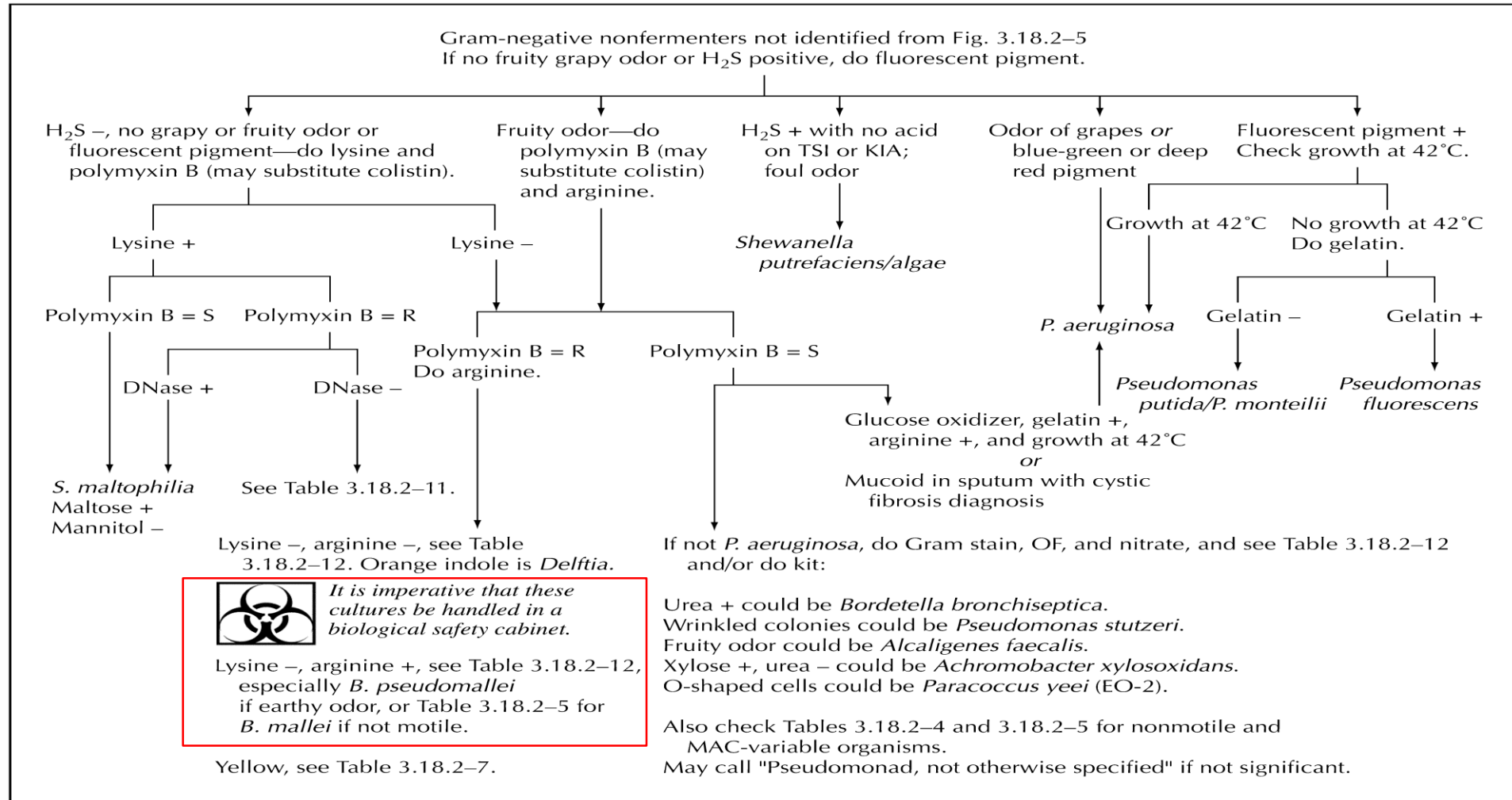
# Go With The ASM CMPH Flow – *B. mallei*

Figure 2-5



# Go With The ASM CMPH Flow – *B. mallei*

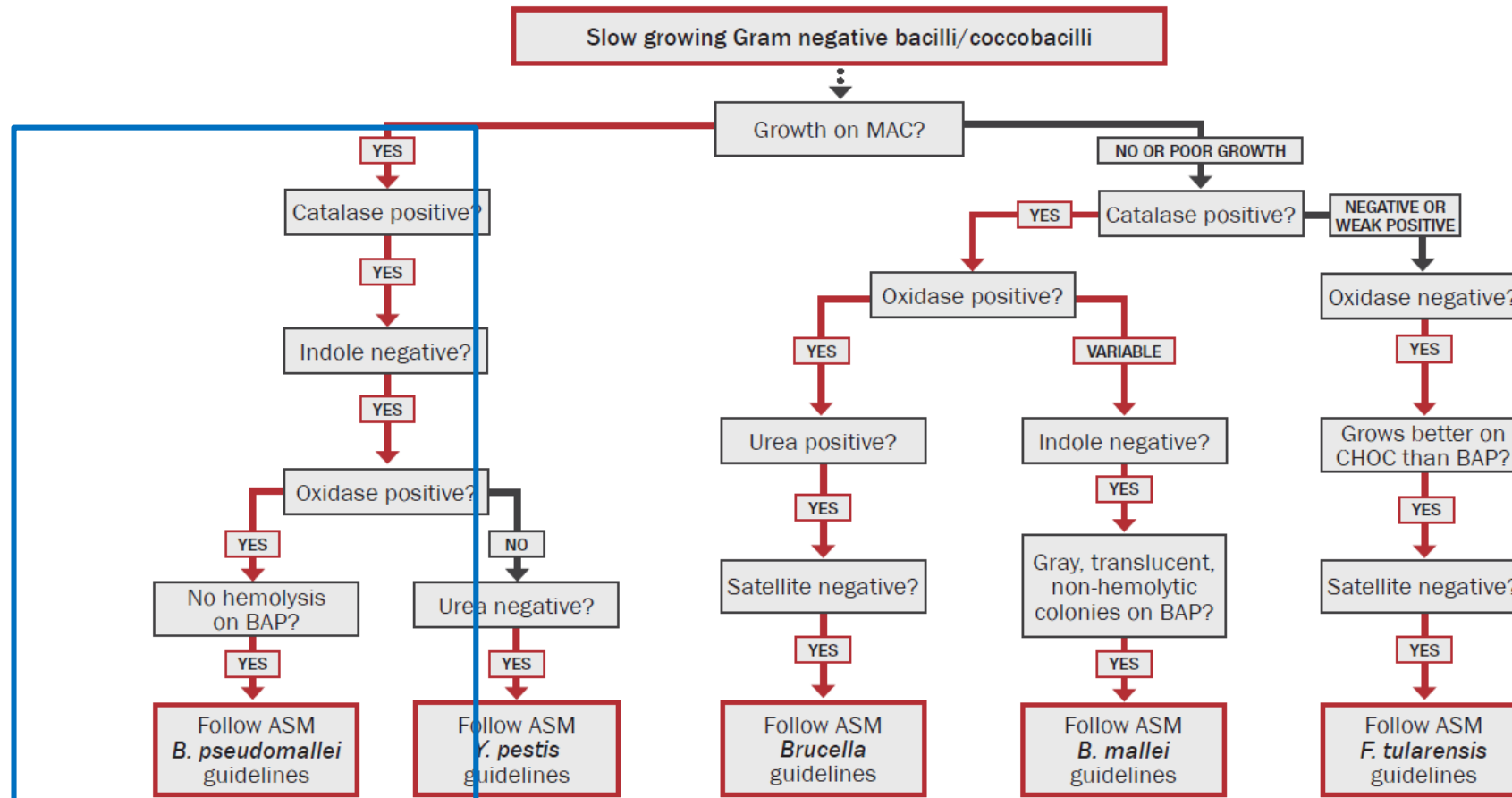
Figure 2-6



# *Burkholderia pseudomallei*

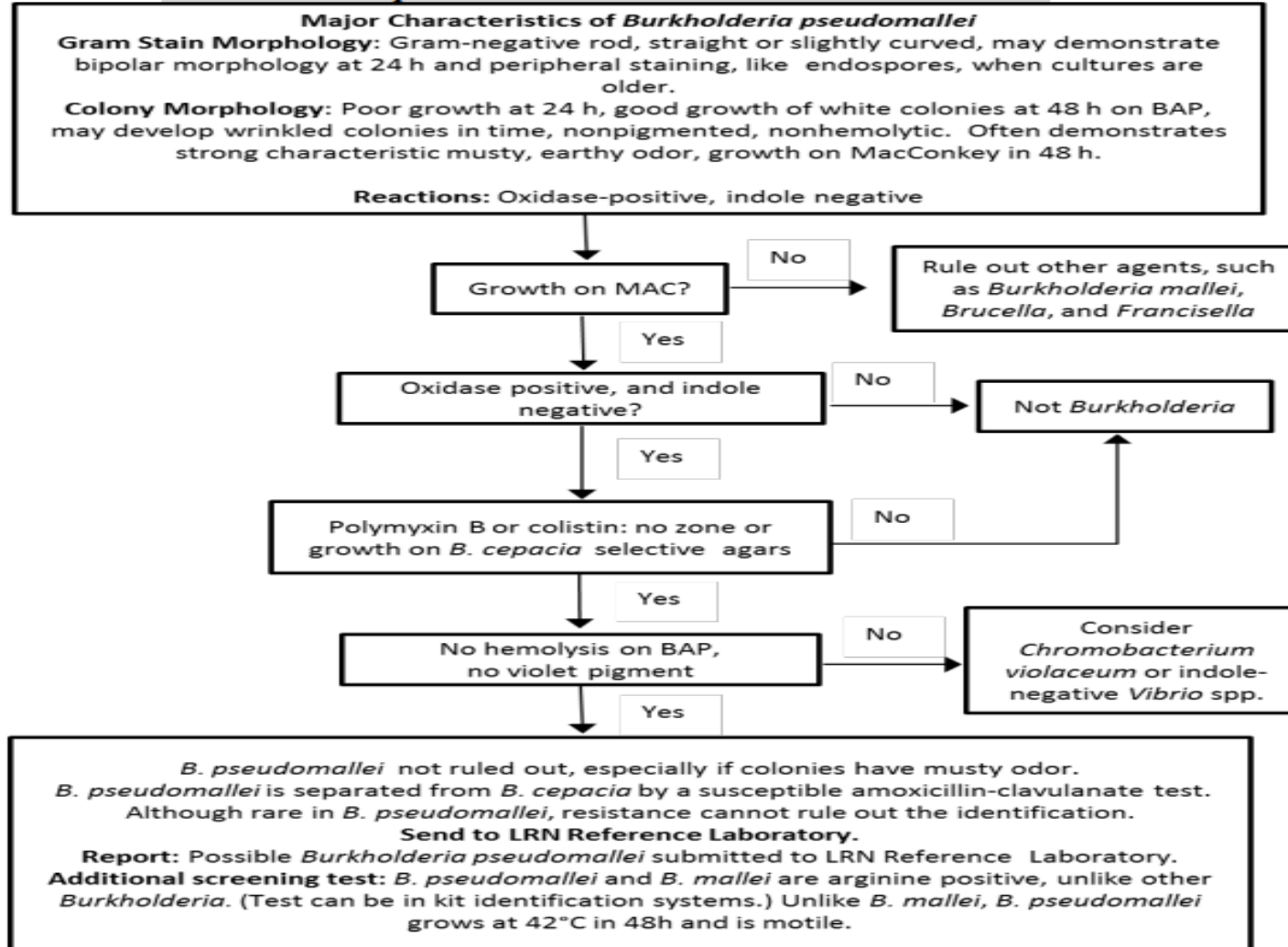
Oxidase positive; Indole negative; **Definitely down to earth**

# Go With The **APHL** Flow – *B. pseudomallei*

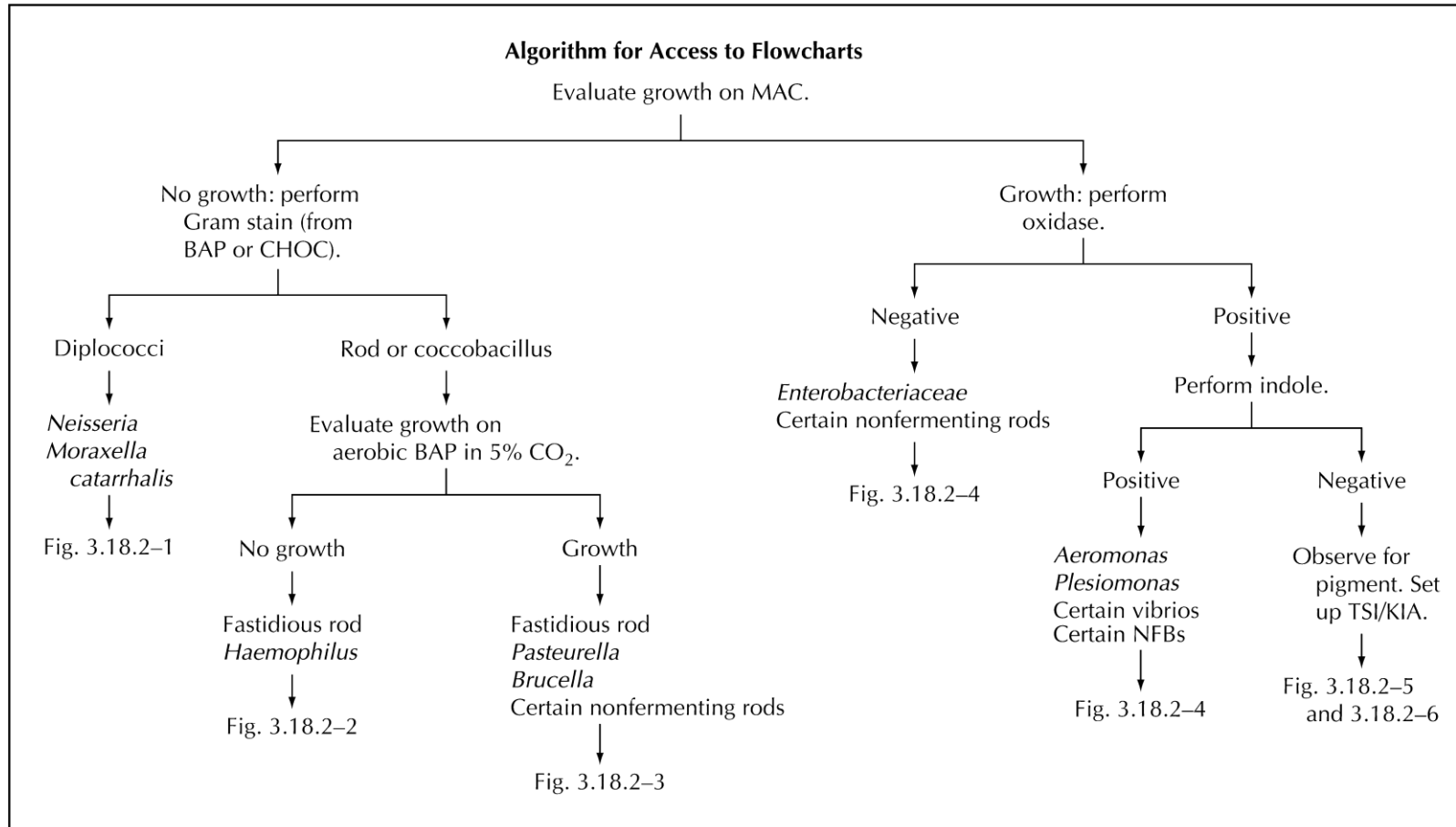


# Go With The **ASM Sentinel Lab** Flow – *B. pseudomallei*

## ***Burkholderia pseudomallei* Identification Flowchart**

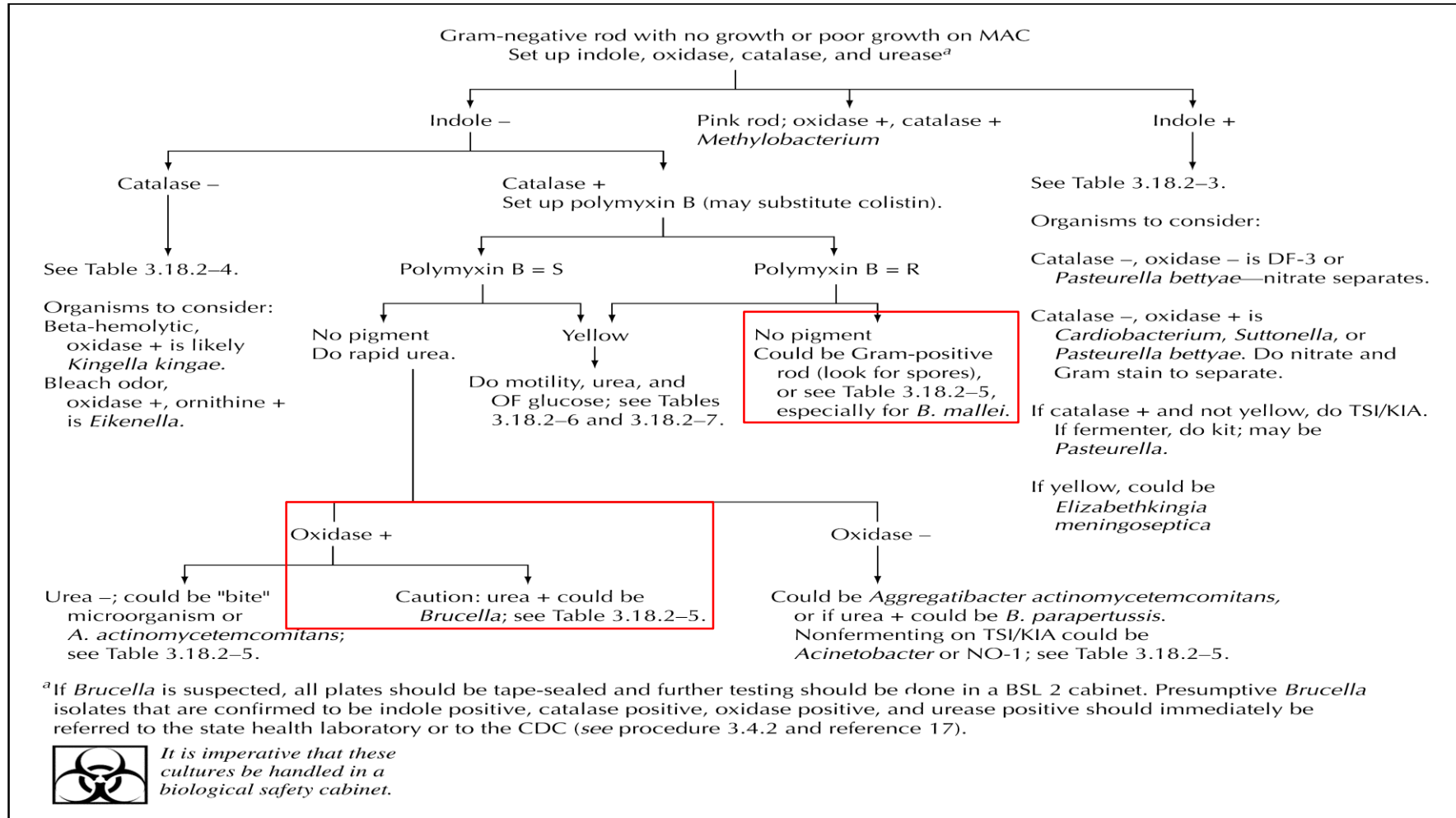


# Go With The **ASM CMPH** Flow – *B. pseudomallei*



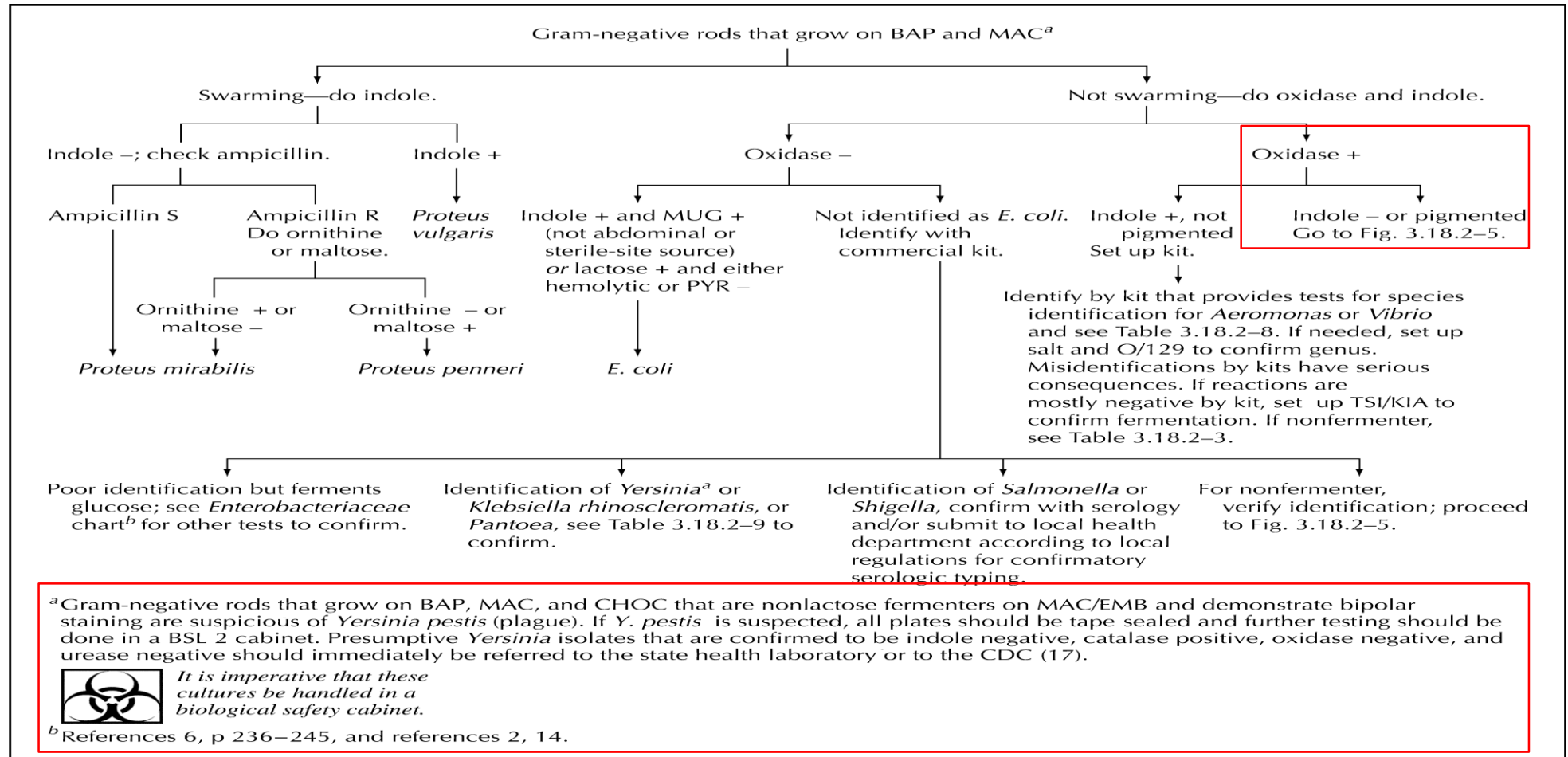
# Go With The **ASM CMPH** Flow – *B. pseudomallei*

Figure 2-3



# Go With The ASM CMPH Flow – *B. pseudomallei*

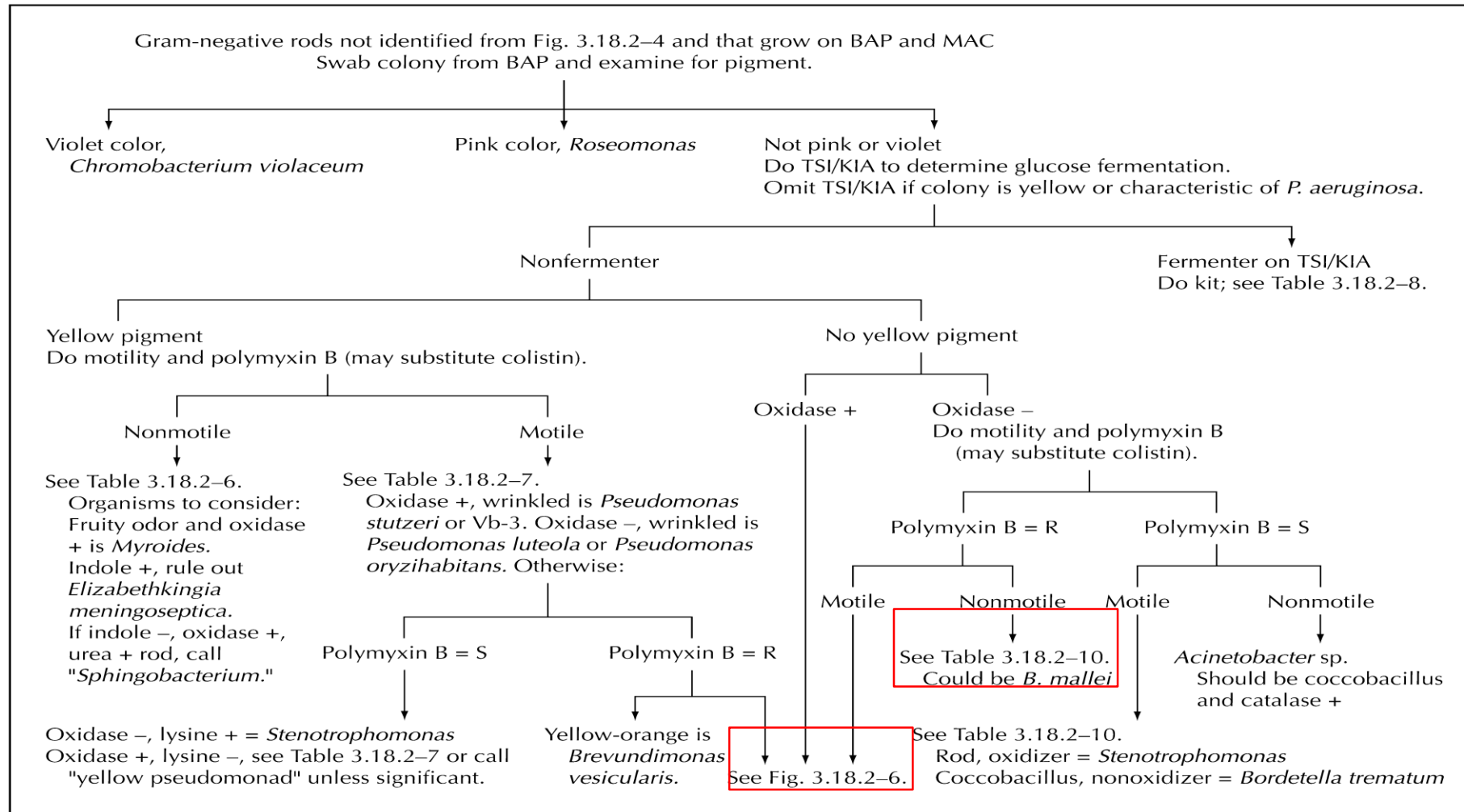
Figure 2-4





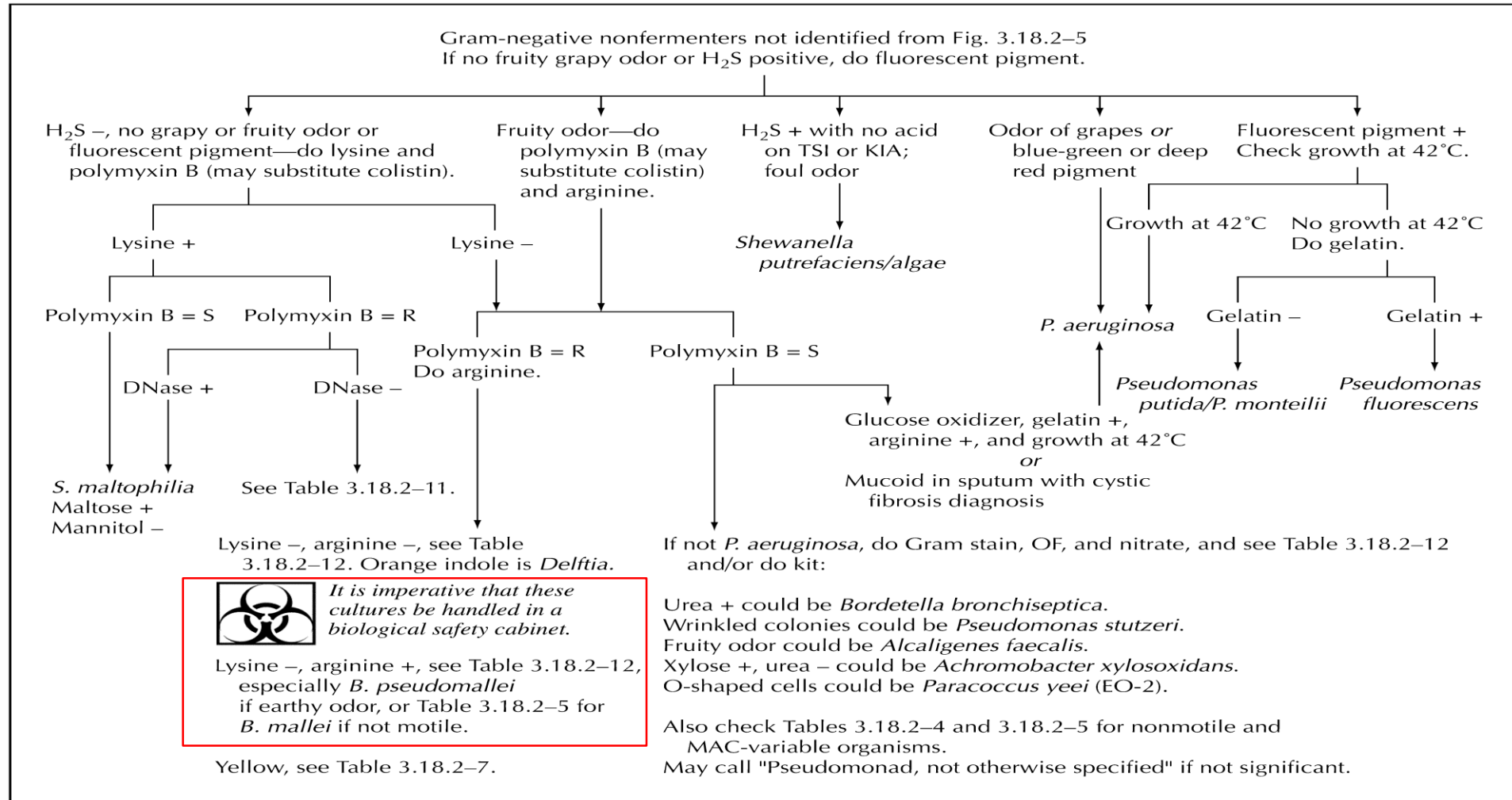
# Go With The **ASM CMPH** Flow – *B. pseudomallei*

Figure 2-5



# Go With The ASM CMPH Flow – *B. pseudomallei*

Figure 2-6



# Tables

**Troubleshooting Organism Mimicry**

**Table 3.18.2–5** Biochemical differentiation of non-yellow-pigmented, Gram-negative rods that are catalase positive and indole negative but do not grow well on MAC<sup>a</sup>

Organism(s) <sup>b</sup>	MAC	Oxidase	Urease <sup>c</sup>	Nitrate	Glucose	Arginine	Other differential trait(s)
<i>Aggregatibacter actinomycetemcomitans</i>	–	V	–	+	F, gas V	–	Tiny rod; esculin –; acid slant and butt in TSI; sucrose fermentation –.
<i>Avibacterium gallinarum</i>	V	+	–	+	F	–	Rod, acid slant and butt in TSI; sucrose +
<i>Bordetella parapertussis</i>	–	–	+	–	n-o	NA	Coccobacilli, brown pigment on MH agar, beta-hemolytic
<i>Brucella</i> <sup>c</sup>	V	+	+	+	O	NA	Coccioid tiny cells; work in safety cabinet
<i>Burkholderia mallei</i>	V	V	V	+	O	+	Coccobacilli, hazardous; resistant to polymyxin B; citrate –
<i>Capnocytophaga canimorsus/cynodegmi</i> (DF-2)	–	+	–	–	F <sup>a</sup>	+	Rod; gliding motility, ONPG +; from dog and cat bites
<i>Neisseria zoodegmatidis</i> (EF-4b)	V	+	–	+	O	–	Reduces nitrite with no gas; no reaction in TSI; from dog and cat bites
<i>Neisseria animaloris</i> (EF-4a)	V	+	–	+, gas	F	V	May reduce nitrite with no gas; 73% acid in butt of TSI; from dog and cat bites
CDC group EO-5	–	–	W	–	O	–	Coccobacilli; 20% are yellow
CDC group NO-1	V	–	–, W	+	n-o	–	Rod; from dog and cat bites. <i>Acinetobacter</i> organisms that fail to grow on MAC are coccioid and nitrate –.
<i>Methylobacterium</i> spp.	V	+	V	V	n-o, O	NA	Vacuolated rod pink in 72 h; grows faster on CHOC; motile
<i>Moraxella</i> spp.	V	+	–	V	n-o	NA	Coccioid, thick cells
<i>Neisseria weaveri</i>	V	+	–	–	n-o	–	Rod; PDA V, nitrite +; Gilardi rod group 1 similar, except nitrite – and PDA +
<i>Oligella ureolytica</i>	V	+	+	+, gas V	n-o	NA	Coccioid, PDA +, motile
<i>Oligella urethralis</i>	V	+	–	–	n-o	NA	Coccioid, PDA +, nitrite +
<i>Paracoccus yeei</i> (EO-2)	V	+	V	+	O	NA	Coccioid but large cells with vacuoles, appear as “o-shaped” cells; mucoid colony
<i>Pasteurella haemolytica</i>	V	+	–	+	F	–	Acid slant and butt in TSI; beta-hemolytic
<i>Psychrobacter immobilis</i>	V	+	V	V	n-o, O	NA	Coccioid; may have rose-like odor
<i>Psychrobacter phenylpyruvicus</i>	V	+	+	V	n-o	NA	Coccioid, PDA +, nitrite –, 6.5% salt +

Table 2-5

<sup>a</sup> For indole-positive strains, see Table 3.18.2–3. All strains nonmotile, except as noted, but even with those, motility is difficult to demonstrate. See Table 3.18.2–4 for catalase-variable rods. Data are from references 5, 22, 26, and 28. W, weak reaction; NA, not applicable or available. See footnote c to Table 3.18.2–1 and footnote a to Table 3.18.2–3 for other abbreviations and symbols.

<sup>b</sup> See reference 8 for guidelines on when DNA target sequencing may be useful for identification.



*It is imperative that these cultures be handled in a biological safety cabinet.*

<sup>c</sup> If *Brucella* is suspected, all plates should be tape sealed and further testing should be done in a BSL 2 cabinet. Presumptive *Brucella* isolates that are confirmed to be indole positive, catalase positive, oxidase positive, and urease positive should immediately be referred to the state health laboratory or to the CDC (see procedure 3.4.2 and reference 17).

**Table  
2-10**

**Table 3.18.2–10** Biochemical reactions of non-glucose-fermenting, Gram-negative rods that are catalase positive, oxidase negative or delayed, and grow well on MAC within 48 h<sup>a</sup>

Organism(s) <sup>b</sup>	Motility	Pigment	Polymyxin B	Lysine	Glucose	Urea	Nitrate	Arginine	Mannitol	Maltose	Other differential trait(s)
<i>Acinetobacter</i> spp. Saccharolytic	–	–	S	–	O	–	–	V	–	NA	Coccobacilli; most are <i>A. baumannii</i> , the only one that grows at 42°C
Asaccharolytic	–	–	S	–	n-o	–	–	V	–	–	Coccobacilli; not all grow on MAC; nitrate negativity and rod shape separates them from NO-1
<i>Bordetella trematum</i>	+	–	NA	–	n-o	–	V	–	–	–	Coccobacilli
<i>Burkholderia gladioli</i> / <i>Pandoraea</i>	+	Variable yellow	R	–	O (O)	V	V	–	V	V	Rod; see Table 3.18.2–11 for species identification; DNase –
<i>Burkholderia mallei</i>	–	–	R	–	O	V	+	+	V	V	Coccobacilli, hazardous; MAC and oxidase V, citrate –; no growth at 42°C
<i>Pseudomonas oryziha-</i> <i>bitans/luteola</i>	+	Yellow	S	–	O	V	V	V	+	+	Wrinkled colonies; PYR +
<i>Roseomonas</i> spp.	V	Pink	R	NA	n-o	+	V	NA	–	–	Coccioid, mucoid, delayed positive oxidase; does not absorb long-wave UV light
<i>Stenotrophomonas maltophilia</i>	+	Variable yellow	V	+	O	–	V	–	–	+	Rod; DNase +, PYR –

<sup>a</sup> Reactions from references 5, 22, and 28. All strains are catalase positive. For oxidase-negative, yellow-pigmented organisms, see Table 3.18.2–7. See footnote c to Table 3.18.2–1 and footnote a to Tables 3.18.2–3 and 3.18.2–5 for abbreviations and symbols.

<sup>b</sup> See reference 8 for guidelines on when DNA target sequencing may be useful for identification.

**Table  
2-12**

**Table 3.18.2–12** Biochemical reactions of nonyellow Gram-negative rods that are oxidase positive and grow well on MAC within 48 h<sup>a</sup>

Organism(s) <sup>d</sup>	Polymyxin B	Glucose	Nitrate	Arginine	Urea	PYR	Xylose	Mannitol	Sucrose	Maltose	Other differential trait(s)
<i>Achromobacter xylosoxidans</i> subsp. <i>denitrificans</i>	S	n-o	+, gas	–	–	+	–	–	–	–	<i>Achromobacter piechaudii</i> is similar but nitrite – and no gas from nitrate.
<i>Achromobacter xylosoxidans</i> subsp. <i>xylosoxidans</i>	V	O	+, gas V	V	–	+	+	–	–	–	Oxidizes xylose better than glucose
<i>Alcaligenes faecalis</i> <sup>c</sup>	S	n-o	–	–	–	–	–	–	–	–	Fruity odor; nitrite +, acetamide +
<i>Bordetella avium/hinzii</i>	S	n-o	–	V	– <sup>+</sup>	+	–	–	–	–	<i>B. avium</i> is beta-hemolytic, nitrite –
<i>Bordetella bronchiseptica</i>	S	n-o	+	–	+	–	–	–	–	–	Grows on SS agar
<i>Brevundimonas diminuta</i>	R	n-o/O	–	–	V	–	–	–	–	–	Brown on MH agar
<i>Burkholderia cepacia</i> complex, <i>Ralstonia</i> , <i>Pandoraea</i>	R	O (O)	V	–	V	V	V	V	V	V	See Table 3.18.2–11 for species identification.
<i>Burkholderia pseudomallei</i> <sup>b</sup>	R	O	+, gas	+	V	–	+	+	V	+	White opaque colonies with sheen, then wrinkled, not beta-hemolytic, earthy odor; lactose +
CDC group Ic	NA	O	+	+	V	NA	–	–	–	+	
<i>Comamonas</i> spp.	S	n-o	+	–	–	+ <sup>–</sup>	–	–	–	–	
<i>Delftia acidovorans</i>	R	n-o	+	–	–	+	–	+	–	–	Orange color of colony with Kovács' indole
<i>Ochrobactrum anthropi</i> and unnamed <i>Achromobacter</i> groups B, E, and F	V	O	V, gas V	V	+	+	+	V	V	V	PDA +
OFBA-1	S	NA	+, gas	+	–	NA	NA	NA	NA	NA	Beta-hemolytic; turns OF base +
<i>Oligella ureolytica</i>	NA	n-o	+, gas V	NA	+	NA	–	–	–	–	PDA +
<i>Paracoccus yeei</i> (EO-2)	S	O	+	–	V	NA	+	–	–	–	Coccoid but large cells with vacuoles; mucoid colonies
<i>Pseudomonas alcaligenes</i> , <i>Pseudomonas</i> CDC group 1, <i>Pseudomonas pseudoalcaligenes</i>	S	n-o	V	V	–	–	W	–	–	–	<i>P. pseudoalcaligenes</i> is nitrate + but no gas and 42°C +; <i>P. alcaligenes</i> is nitrate V and 42°C –; <i>Pseudomonas</i> CDC group 1 is nitrate + with gas.
	S	O	V	+	V	V	+	V	V	–	<i>Pseudomonas</i> fluorescent group See Fig. 3.18.2–6 for species separation.
<i>Pseudomonas stutzeri</i>	S	O	+, gas	–	V	–	+	V	–	+	Wrinkled colonies
<i>Cupriavidus pauculus</i> (IVc-2)	V	n-o	–	NA	+	+	–	–	–	–	No growth on SS agar
<i>Rhizobium radiobacter</i>	V	O	V	–	+	+	+	+	+	+	PDA +
<i>Shewanella putrefaciens/algae</i>	S	O	+	–	V	+	–	–	+	+	H <sub>2</sub> S +, ornithine +, brown, foul smelling

<sup>a</sup> All strains are motile and indole negative. Also see Table 3.18.2–5 for nonmotile, Gram-negative rods that are MAC variable. Verify that strains are nonfermenting rods using TSI or KIA. For fermenting rods, see Table 3.18.2–8. Data are from references 5, 22, 28, 35, and 36. SS, salmonella-shigella. See footnote c to Table 3.18.2–1 and footnote a to Tables 3.18.2–3, 3.18.2–5, and 3.18.2–7 for other abbreviations and symbols for reaction key.

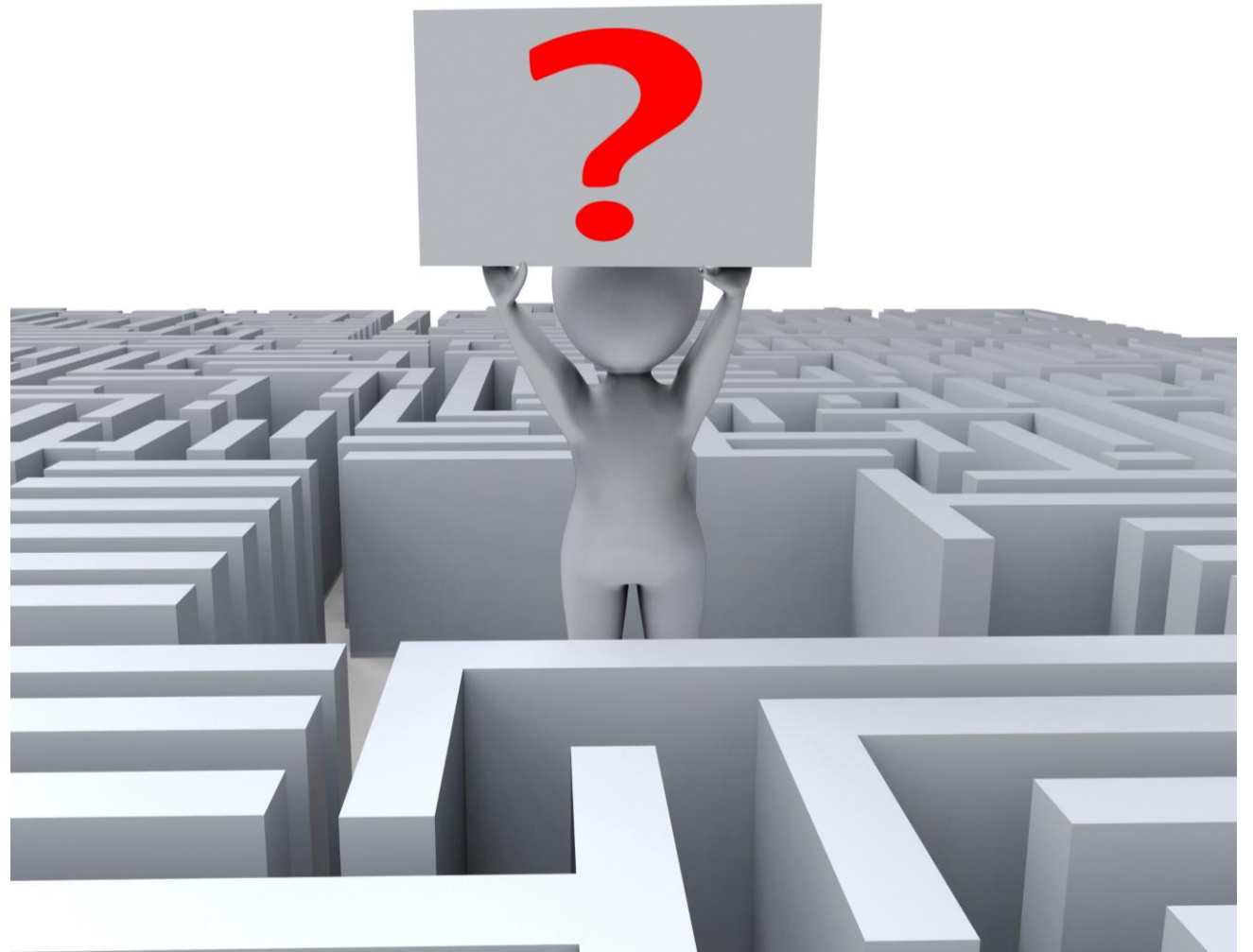
<sup>b</sup> *B. mallei* can have similar reactions but is nonmotile, has no odor, and does not produce gas from nitrate.

<sup>c</sup> To separate *Alcaligenes faecalis* from other related nonoxidizers: *Ralstonia gilardii* is nitrite negative; nonyellow *Myroides* is urea and PYR positive but nonmotile and polymyxin B resistant; and Gilardi rod group 1 is nonmotile and PDA positive.

<sup>d</sup> See reference 8 for guidelines on when DNA target sequencing may be useful for identification.

Now...

Are you  
ready for  
the real  
thing?



# References

1. American Society for Microbiology, SENTINEL LEVEL CLINICAL LABORATORY GUIDELINES FOR SUSPECTED AGENTS OF BIOTERRORISM AND EMERGING INFECTIOUS DISEASES - Glanders: *Burkholderia mallei* and Melioidosis: *Burkholderia pseudomallei*, Washington, DC, Revised March 2016.
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