

STANTON TERRITORIAL HEALTH AUTHORITY

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TITLE: MAST DISCS FOR ESBL	Revision Date: Issue Date:		
	20-April-2018	20-April-2016	
Document Number: MIC50645	Status: Approved		
Distribution: Microbiology Test Manual	Page: 1 of 6		
Approved by:	Signed by:		
S. Asmussen, Manager of Diagnostic Services	bulg	Successo	
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PURPOSE:

Detection of plasmid mediated broad spectrum beta-lactamases (BSBL) including extended spectrum beta-lactamases (ESBL) and AmpC cephalosporinases (AmpC) is important for both therapeutic and epidemiological purposes.

KIT INFORMATION:

	4 cartridges, each cartridge containing 50 discs			
	Cartridge A: Cefpodoxime 10ug discs			
	Cartridge B: Cefpodoxime 10ug + ESBL inhibitor discs			
Туре	Cartridge C: Cefpodoxime 10ug + AmpC inhibitor discs			
	Cartridge D: Cefpodoxime 10ug +ESBL inhibitor +AmpC			
	inhibitor discs			
Source	Alere Canada			
	Store at 2-8°C in the containers provided until the expiry date shown			
Storage	on the pack label. Allow to equilibrate to room temperature before			
Requirements	opening.			

MATERIALS:

- Sterile swabs, forceps, applicator sticks
- Normal Saline (0.9% Sodium Chloride)
- Densitometer
- 35°C ambient air incubator
- Mueller Hinton Agar
- Small metric ruler

TITLE: MAST DISC FOR ESBL	Revision Date:	Issue Date:	
	20-April-2018	20-April-2016	
Document Number: MIC50645	Status: Approved		
Distribution: Microbiology Test Manual	Page: 2 of 6		

QUALITY CONTROL:

QC must be performed upon receipt of a new lot or shipment, and weekly thereafter.

- Must be run with both Positive and Negative controls
- A TQC order will be generated upon batch receiving the new lot/shipment and automatically generated every week as per weekly KB QC. To result the TQC order, log into TQC→Resulting Worklist → MICS – All Scheduled Micro → Run Worklist →MAST
- New lot/shipment QC should be done upon receipt to the lab.
- Weekly MAST QC should be done when performing Vitek weekly QC.

POSITVE CONTROL (AmpC pos):

Enterobacter cloacae NCTC 13406 (non-ATCC strain)

NEGATIVE CONTROL (ESBL and AmpC neg):

E.coli ATCC 25922**

**Deterioration/malfunction check: Compare all the zone diameters in the negative control (compare zone diameters of disks A, B, C and D). All diameters should be within 2mm of each other or less (ideally the zone diameters from all disks should be equal). Any greater differences between the disks imply malfunction or deterioration and the MAST disks should not be used.

SPECIAL SAFETY PRECAUTIONS:

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potentially infectious materials or cultures.

• Lab gown must be worn when performing activities with potential pathogens.

TITLE: MAST DISC FOR ESBL	Revision Date:	Issue Date:	
	20-April-2016	20-April-2016	
Document Number: MIC50645	Status: Approved		
Distribution: Microbiology Test Manual	Page: 3 of 6		

- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used where there is a known or potential risk of exposure to splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes, and other sharp objects should be strictly limited.

PROCEDURE INSTRUCTIONS:

Step	Action
1	LIS CODE: MAST Using a pure, fresh culture of the test organism, prepare a suspension in saline equivalent to a 0.5 McFarland standard
2	Moisten a sterile swab with the suspension and spread evenly over a Mueller Hinton Agar plate, as for Kirby Bauer testing. Wait for moisture to absorb into agar, approx 3 minutes. Do not wait more than 15 minutes to apply disks.
3	Using sterile forceps (may sterilize with an alcohol pad) place one of each type of MAST disc onto the inoculated media, ensuring sufficient space between the discs to allow formation of clearly defined zones of inhibition.
4	Incubate the plate at 35-37°C for 18 to 24 hours.
5	Measure and record in mm the diameter of any zones of inhibition that are observed. Discs showing no zone of inhibition should be recorded at 6mm.

TITLE: MAST DISC FOR ESBL	Revision Date: 20-April-2018	Issue Date: 20-April-2016
Document Number: MIC50645	Status: Approved	
Distribution: Microbiology Test Manual	Page: 4 of 6	

INTERPRETATIONS OF RESULTS:

1. Manual Interpretation

Step	Action				
1	Compare the zone of inhibition of the Cefpodoxime disc A to the zones on inhibition of				
	each of the Cefpodoxime plus inhibitor disc	s B, C , and D .			
	IF	THEN			
	All zones are within 2mm of each other	Organism is NEGATIVE for both			
		ESBL and AmpC activity			
2	Subtract A from B ; C from D ; B from D; A	from C			
	IF	THEN			
	Each of B-A and D-C >=5mm AND	The organism demonstrates ESBL activity			
	Each of D-B and C-A <5mm	alone.			
	Each of B-A and D-C <5mm AND	The organism demonstrates AmpC			
	Each of D-B and C-A >=5mm activity alone.				
3	Subtract C from D and A from B				
	IF	THEN			
	D-C >=5mm BUT B-A <5mm	The organism demonstrates ESBL and			
		AmpC combined activity.			

2. Template Interpretation

Step	Action								
	Open up the	e ESBL calcu	ulator prog	ram					
	In the S: drive: LAB \rightarrow Bacteriology \rightarrow Quality Control \rightarrow ESBL calculator								
	A	В	С	D	E	F	G	AJ	AM
1	Mast Group		ESBL/An	npC	Calc	ulato	r	CJH v9.00 2012.04.10	
		LAB NUMBER	COMMENTS	A CPD (mm)	B CPD + ESBL INHIBITOR (mm)	C CPD+AMPC INHIBITOR (mm)	D CPD + ESBL + AMPC INHIBITORS (mm)	Intrepretation	Notes
				(Minir	ium Value mu	st be at least 6		AMPC	
	Doe, John	12345678 li	nput HCN here	15	15	25	25	AMILC	

TITLE: MAST DISC FOR ESBL	Revision Date: 20-April-2018	Issue Date: 20-April-2016
Document Number: MIC50645	Status: Approved	
Distribution: Microbiology Test Manual	Page: 5 of 6	

Input the following: • Patient Name

Lab Number

 Health Card Number
 Zone interpretation for Disks A, B, C, D

NOTE: for zone sizes where there is growth up to the disk input 6mm not 0mm
The system will calculate and interpret
SAVE the worksheet

ALTERNATIVELY A calculation program is available at <u>www.mastgrp.com</u>. This has been added to the desktop on the Urine Bench computer.

REPORTING RESULTS:

- In the plate log (Media field) add media "MAST". A keypad will appear in which you can enter the zone sizes for A, B, C and D discs, and the interpretation.
- Go to "Isolates" and add, using A or B, not a number, the isolate "ESBL Confirmed". This will mark the isolate for epidemiology reporting.
- From the AST Manual use the B-lactam charts to determine the comment to add to the isolate which is being reported. These are all in the comment keypad.
- Copy results to appropriate HPU.

LIMITATIONS:

- Use discs only from the same batch/pack. Do not use cartridges from different batches together; batches should never be mixed.
- Occasionally a pattern will occur where the "A" disc has no zone while the "C" disc does. This is an indeterminate result and the organism needs to be sent to DynaLife for further testing.

TITLE: MAST DISC FOR ESBL	Revision Date: Issue Date:		
	20-April-2018	20-April-2016	
Document Number: MIC50645	Status: Approved		
Distribution: Microbiology Test Manual	Page: 6 of 6		

RELATED DOCUMENTS:

• MIC51000: Kirby-Bauer

REFERENCES:

• Mast Group Ltd., MASTDISCS[™]ID test insert.

REVISION HISTORY:

REVISION	DATE	Description of Change	REQUESTED BY
1.0	310ct12	Initial Release	S.Webber
1.1	31Dec13	Reporting procedure	S. Webber
1.2	15Apr14	Weekly QC positive control (<i>E.cloacae</i>)	Driedger (L)
2.0	31Mar16	Update of "Special Safety Precautions" to reflect risk assessment recommendations.	C. Russell