STANTON TERRITORIAL HEALTH AUTHORITY

Yellowknife, Northwest Territories

TITLE: Wet Preparation using Saline	**Revision Date:** 20-April-2018	**Issue Date:** 20-April-2016
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Distribution: Microbiology Test Manual	**Page:** 1 of 4	
Approved by: S. Asmussen, Manager of Diagnostic Services	**Signed by:** 	

PURPOSE:

- To identify the presence of yeast and/or trichomonas in vaginal swabs
- To confirm the presence of yeast in culture.
- To determine the motility characteristics of some bacteria.

INTRODUCTION:

Wet-preps are performed directly from submitted vaginal swabs on all such specimens to screen for yeast and *Trichomonas vaginalis.* A wet preparation is also useful in confirming the presence of yeast colonies in culture as it is quicker than a Gram stain. Wet-preps can help determine the motility characteristics of some bacteria.

NOTE: Saline is suitable for wet-mount preparations of Gram-negative bacteria. For other bacteria broth mediums must be used. This method will not cover wet-preparations for motility where medium other than saline is required.

SUPPLIES:

- Swab or Culture Plate
- Microscope
- Glass slides and cover slips
- Test tubes
- Sterile Saline

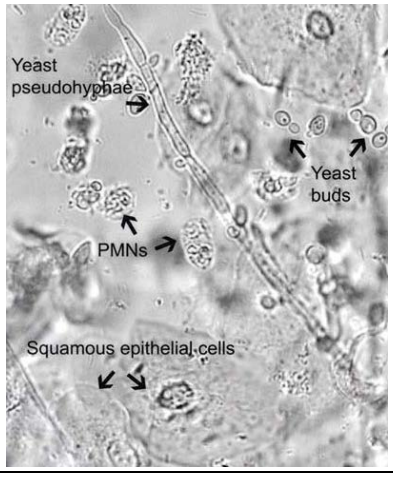
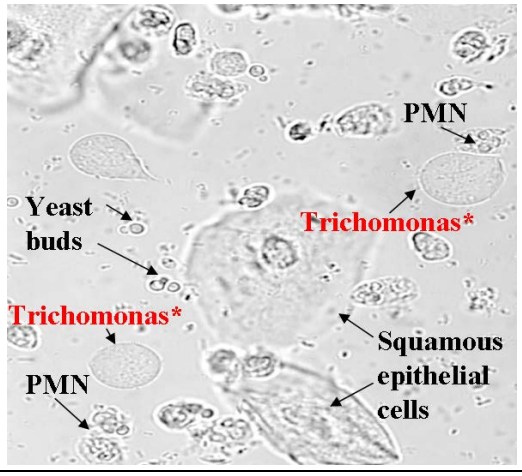
SPECIAL SAFETY PRECAUTIONS:

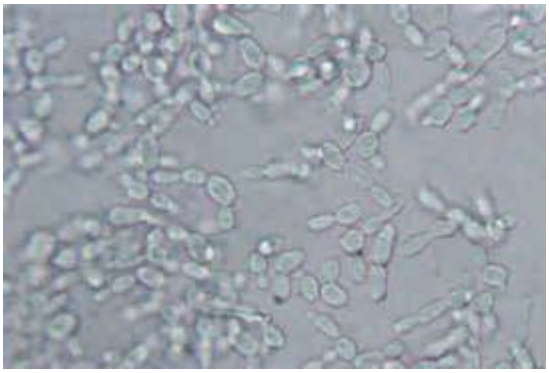
Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potentially infectious materials or cultures.

- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used where there is a known or potential risk of exposure to splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes, and other sharp objects should be strictly limited.

PROCEDURE INSTRUCTIONS:

Step	Action
Setting up a Wet Prep from a Swab	
1	Label the test tube with the accession number and patient demographics.
2	Place approximately 0.5 mL of saline into a test tube.
3	Place the culture swab in the saline and “flick” the tube to loosen the swab material.
4	Incubate for 15 minutes at 35ºC.
5	After incubation dab the culture swab onto the microscope slide then return the swab to its original transport tube.
6	Place the coverslip on the slide. Allow the slide to “settle”
7	Place the slide under the microscope.
8	Examine under 40X for the presence of yeast and trichomonas. TIP: Decrease the light of a light microscope by closing the diaphragm or use a phase

	contrast microscope.  

Step	Action
Setting Up a Wet Prep from Culture	
1	**LIS CODE: ^WP** Place a drop of saline onto a glass slide
2	Using a sterile loop or wooden stick pick the desired colony from the plate and make a smooth suspension in the saline
3	Place the cover slip on the slide. Allow the slide to “settle”
4	Place the slide under the microscope.
5	Examine under 40X for the presence of yeast or to observe motility. TIP: Decrease the light of a light microscope by closing the diaphragm or use a phase contrast microscope. 

INTERPRETATION OF RESULTS:

- In order to call yeast the cells must be budding
- In order to call Trichomonas the flagella must be seen and motile
- If wet mount is for motility and motility is negative by initial wet mount the organism cannot be classified as non-motile until more in-depth motility tests are performed.

REFERENCES:

- American Society For Microbiology. (2004). Clinical Microbiology Procedures Handbook. In H. D. Isenberg (Ed.), *Motility Tests* (p. 3.17.31.2).
- Murray, P., Baron, E. J., Jorgensen, J., Landry, M. L., & Pfaller, M. (2007). *Manual of Clinical Microbiology* (Vol. 1). Washington, DC, USA: ASM Press.

REVISION HISTORY:

REVISION	DATE	Description of Change	REQUESTED BY
1.0	03-NOV-2010	Initial Release	M-L Dufresne
2.0	31Dec2013	LIS updates/pictures	A.Darrach
3.0	31March16	Update of “Special Safety Precautions” to reflect risk assessment recommendations.	C. Russell