

# STANTON TERRITORIAL HEALTH AUTHORITY

# Yellowknife, Northwest Territories

TITLE: Kinyoun Stain	Revision Date:	Issue Date:
	07-April-2017	07-April-2015
Document Number: MIC81600	Status: Approved	
Distribution: Mycobacteria Manual	Page: 1 of 4	
Approved by:	Signed by:	
Gloria Badari, Director, Corporate Services	(Original Signed Copy in	
and Chief Financial Officer	Microbiology)	

# **PURPOSE:**

To standardize the staining of Mycobacteria smears using the Kinyoun stain kit. The Kinyoun stain is a "cold" ZN method; fixing smears does not require heat.

## **SPECIAL SAFETY PRECAUTIONS:**

- Handle all patient samples and testing reagent using "Routine Practices"
- Please refer to the Northwest Territories Infection Prevention and Control Manual,
   March 2012
- Prior to testing all patient are to be identified as per I-0500 Use of Two Patient Identifiers.

#### **REAGENTS and/or MEDIA:**

Stain	Information	
Stains for direct or culture smears:		
	Suppler: BD	
Kinyoun (cold) method (for direct smear confirmation)	Storage: Room Temperature	
	<ul> <li>Kit contains: Carbolfuchsin KF, Decolorizer,</li> </ul>	
	Brilliant Green K	

### **KINYOUN STAIN:**

#### **SUPPLIES:**

- White slide rack
- 4x 4 gauze
- Dedicated TB timer

- Alcohol wipes
- TB Stain Kit K, use before expiration date.

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FILENAME: MIC81600KinyounStainPRO.doc	PRINT DATE: 9 March 2015	

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Distribution: Mycobacteria Manual	Page: 2 of 4		

Step	Action		
Follo	Follow the steps below to stain the direct smear with Kinyoun Stain:		
1	It is acceptable to stain with Kinyoun directly over the A/R stain.		
	If you choose to do this, proceed to Step 3 below.		
2	If preparing a new smear for staining:		
	Refer to document: MIC80800 Mycobacteria Media Inoculation & Direct		
	Smear Preparation		
3	Completely flood all smears with Carbolfuchsin. Leave for 4 minutes		
4	Using gentle water pressure, rinse slides in running water.		
	Continue to rinse slides until all stain is removed; this may take several rinses.		
5	Decolorize smears one-by-one. Do not batch decolorize.		
	Smears are more sensitive to over-decolorization in the Kinyoun stain		
	compared to the A-R stain.		
	Flood one smear with Decolourizer.		
	<ul> <li>Leave on for 3→ 5 seconds.</li> </ul>		
	Using gentle water pressure, rinse slide in running water.		
	Repeat for all smears.		
6	Completely flood all smears at the same time with Brilliant Green.		
	Leave for 30 seconds		
7	7 Using gentle water pressure, rinse slides in running water.		
8	8 Place slides in a white slide holder lined with gauze to drip dry.		
	Place slides in the O2 incubator to speed up drying.		
	Unlike A/R stained slides, Kinyoun Smears do not need to be kept in the dark.		
	Expected Results:		
	Acid Fast Bacilli <mark>fuchsia</mark> / <mark>red</mark> bacilli, beading may be seen		
	Background pale blue/green		

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Document Number: MIC81600	- I -	Status: Approved	
Distribution: Mycobacteria Manual	Page: 3 of 4		

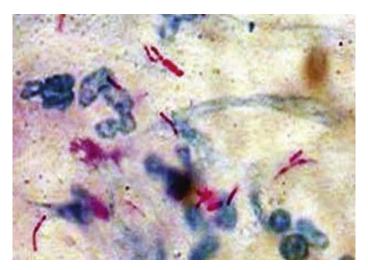


Image 1 (above): AFB seen in Kinyoun smear.

Source; http://www.clevelandclinicmeded.com/medicalpubs/diseasemanagement/infectious-disease/tuberculosis/

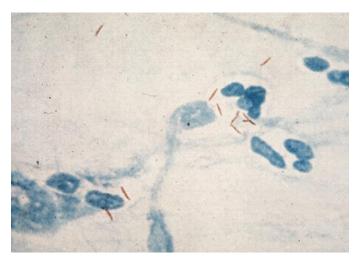


Image 2 (above): AFB seen in Kinyoun smear Source: http://itg.content-e.eu/Generated/pubx/173/tuberculosis/diagnosis.htm

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Document Number: MIC81600	Status: Approved	Status: Approved	
Distribution: Mycobacteria Manual	Page: 4 of 4		

### **RELATED DOCUMENTS:**

- MIC80800 Mycobacteria Media Inoculation & Direct Smear Preparation
- MIC81300 AFB Stain QC & Slide Prep

### **REFERENCES:**

Beckton Dickenson. (2014, August). TB Stain Kits and Reagents. Product Insert.
 United States: Beckton Dickenson.

# **REVISION HISTORY:**

REVISION	DATE	Description of Change	REQUESTED BY
1.0	31Jan15	Initial Release	L. Driedger
	03Feb2015	Review	S. Webber
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