

Document Name: Ear Culture

Approved By:

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Status: **APPROVED**

PURPOSE:

To determine the presence or absence of bacterial pathogens in ear specimens.

SAMPLE INFORMATION:

Type	<ul style="list-style-type: none"> Swab, Amie's with or without charcoal
Source	<ul style="list-style-type: none"> External auditory canal (outer ear)
Stability	<p>If the sample is received in the laboratory and processed greater than 48 hours from collection:</p> <ul style="list-style-type: none"> Add specimen quality comment: "Delayed transport may adversely affect pathogen recovery"
Storage Requirements	Room temperature. If transport is > 2 hours, swabs should be refrigerated.
Criteria for rejection and follow up action	<ol style="list-style-type: none"> Unlabeled/mislabeled swabs Dry swabs

***Note: Tympanocentesis fluid is obtained for the diagnosis of otitis media (middle ear infection). These specimens are to be handled as sterile body fluids. (Refer to MIC34100)**

REAGENTS and/or MEDIA:

- Blood Agar (BAP), Chocolate Agar (CHOC) and MacConkey Agar (MAC)
- Identification reagents: catalase, rapid Staph, rapid Strep, oxidase, etc.

SUPPLIES:

- Wooden sticks
- Disposable inoculation needles
- Microscope slides
- Biosafety cabinet
- 35° ambient air and 37° CO₂ incubators
- Vitek 2 Compact and supplies

SPECIAL SAFETY PRECAUTIONS:

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potential infectious materials or cultures.

- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used when there is a known or potential risk of exposure of splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes and other sharp objects should be strictly limited.

All patient specimens are assumed to be potentially infectious. Universal precautions must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

QUALITY CONTROL:

Refer to Quality Control manual for reagent quality control procedures.

PROCEDURE INSTRUCTIONS:

Step	Action
Processing Swabs for Ear Culture	
1	In the biosafety cabinet, inoculate Blood Agar, Chocolate Agar and MacConkey Agar from the swab. Make gram stain.
2	<p>Streak for isolated growth using a disposable inoculation needle.</p> <div data-bbox="743 638 980 877" style="text-align: center;"> </div> <p>Streak out to cover the whole plate.</p>
3	Place the MAC plate in the O ₂ incubator. Place BAP and CHOC plates in the CO ₂ incubator.
4	Allow smear to dry and perform Gram Stain. Gram stain must be read before culture plates.
5	Examine plates after 24 hour incubation. Record your observations in the LIS.
6	Re-incubate CO ₂ plates for an additional 24 hours. Discard O ₂ plate.

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Pathogens	Normal Flora
<ul style="list-style-type: none"> • <i>Staphylococcus aureus</i> • <i>Streptococcus pyogenes</i> • Other β-hemolytic <i>Streptococci</i> • <i>Streptococcus pneumoniae</i> • <i>Haemophilus influenzae</i> • <i>Moraxella catarrhalis</i> • <i>Pseudomonas aeruginosa</i> • Non-fermentative Gram-negative bacilli • Enterobacteriaceae spp. • <i>Candida spp.</i> • Fungi 	<ul style="list-style-type: none"> • Acinetobacter spp. • Aerococcus spp. • Bacillus spp. • Coagulase negative Staphylococcus • Corynebacterium • Micrococcus spp. • Neisseria species • Anaerobes • Viridans Streptococcus

INTERPRETATION OF RESULTS:

Step	Action
Interpretation of Otitis externa specimens	
1	<p>Confirm gram stain has been read prior to reading culture plates. Ensure growth on culture media correlates with gram stain results. If discordant results are found:</p> <ul style="list-style-type: none"> • Re-examine smear and culture plates. • Check for anaerobic growth. • Re-incubate culture to resolve. • May need to inoculate special selective media. • Consider re-smearing or re-planting specimen to exclude the possibility of error.
2	<p>Observe plates at 24 hours and 48 hours for growth of enteric Gram-negative bacilli, <i>Pseudomonas aeruginosa</i>, Yeasts, <i>Vibrio alginolyticus</i>, <i>Streptococci</i>, <i>Staphylococcus aureus</i>, <i>Haemophilus influenzae</i>, <i>Moraxella catarrhalis</i> and Fungi.</p> <p>Usually only one pathogen is responsible for otitis externa. Mixed cultures should be minimally identified.</p>
3	<p>Skin flora such as coagulase negative staphylococci and coryneforms are normal in the external ear canal and should not be further evaluated.</p>
4	<p>Mixed cultures of Gram-negative rods should be minimally identified.</p>

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Otitis externa pathogens identification			
ORGANISM	GRAM STAIN	IDENTIFICATION	SUSCEPTIBILITY
<i>Staphylococcus aureus</i>	Gram-positive cocci, in clusters	<ul style="list-style-type: none"> Catalase: Positive Slide coagulase: Positive Tube coagulase: Positive 	Vitek AST-GP67
<i>Streptococcus pyogenes</i>	Gram-positive cocci, in chains	<ul style="list-style-type: none"> Catalase: Negative PYR: Positive Step grouping: A positive 	KB as per ASTM
Other beta-hemolytic Streptococci	Gram-positive cocci, in chains	<ul style="list-style-type: none"> Catalase: Negative PYR: Negative Strep grouping: B, C or G positive 	KB as per ASTM
<i>Streptococcus pneumoniae</i>	Gram-positive cocci, in pairs	<ul style="list-style-type: none"> Alpha hemolytic Catalase: Negative Optochin: Sensitive Vitek GPI 	KB and E-test as per ASTM
<i>Haemophilus influenza</i>	Gram-negative coccobacilli	<ul style="list-style-type: none"> Gram-negative coccobacilli Growth on CHOC, no growth on BA except as satellitism ALA: Negative 	Not performed. Add isolate comment: &HAEM
<i>Moraxella catarrhalis</i>	Gram-negative diplococci	<ul style="list-style-type: none"> Gram-negative diplococci Oxidase: Positive Catarrhalis disk: Positive 	Beta-lactamase (only report if positive)
<i>Pseudomonas aeruginosa</i>	Gram-negative bacilli	<ul style="list-style-type: none"> Oxidase: Positive Characteristic odor and pigment 	Vitek AST-N213
Other Gram-negative bacilli	Gram-negative bacilli	<ul style="list-style-type: none"> Vitek GN or API 	Vitek AST-N213
<i>Vibrio alginolyticus</i>	Gram-negative bacilli	<ul style="list-style-type: none"> Oxidase: Positive Indole: Positive Grows well on MAC Vitek GNI 	Vitek AST-N213
Candida species	Yeast	<ul style="list-style-type: none"> Wet prep: Yeast Vitek YST ID 	Not performed

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REPORTING RESULTS:

No growth after 2 days	<ul style="list-style-type: none"> Report “No Growth after 2 days”
Mix of skin flora (CNS, diptheroids, Viridans strep, Bacillus spp., Micrococcus spp.)	<ul style="list-style-type: none"> Report “Mixture of skin flora” List quantitation.
Mix of enteric Gram-negative bacilli	<ul style="list-style-type: none"> Report “Mixture of coliform organisms” List quantitation.
Growth or mix of other non-pathogenic organisms	<ul style="list-style-type: none"> Report “Commensal flora” List quantitation.
Growth of anaerobe	<ul style="list-style-type: none"> Report “Gram Negative Bacilli Anaerobic” or “Gram Positive Cocci Anaerobic” List quantitation.
Growth of mixed anaerobe	<ul style="list-style-type: none"> Report “Mixture of anaerobic organisms” List quantitation.
Growth of fungus	<ul style="list-style-type: none"> Report “Fungal species” with isolate comment &REF2 to state: “This organism has been referred for further identification.” List quantitation. Add test ?REFE. Send to Prov. Lab for identification as per MIC10500 – Referral of Category B specimens to Provincial Laboratory. Freeze specimen and enter into specimen isolate log.
Predominant or pure growth of yeast (if not predominant or pure report as part of commensal flora)	<ul style="list-style-type: none"> If Vitek identified as Candida albicans, report. If not Candida albicans, report as YSTNOT (Yeast, not Candida albicans). List quantitation.
Growth of pathogens	<ul style="list-style-type: none"> Report organism identification. List quantitation. Report susceptibility results as per DynaLIFE ASTM.

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PROCEDURE NOTES:

- An external ear culture with a predominant Gram-negative bacillus, beta-hemolytic Streptococcus, *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Haemophilus influenzae* or *Moraxella catarrhalis* generally indicates infection with that organism.
- Historically, the nasopharynx was cultured in order to predict the pathogens in the middle ear. This practice is no longer recommended.
- Treatment of otitis externa usually consists of ear drops.

LIMITATIONS:

- False-negative cultures can result from overgrowth of the culture with normal skin flora.
- False-positive results can be caused by over-interpretation of culture results.

REFERENCES:

- Clinical Microbiology Procedures Handbook, 4th edition, ASM Press, 2016
- Jorgensen J.H., Pfaller M.A., Carroll K.C., Funke G., Landry M.L., Richter S.S., Warnock D.W. 2015. Manual of Clinical Microbiology, 11th edition, ASM Press, Washington, D.C.

REVISION HISTORY:

REVISION	DATE	Description of Change	REQUESTED BY
1.0	06-NOV-17	Initial Release	L. Steven

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