	Stanton Territorial Hospital P.O. Box 10, 550 Byrne Road YELLOWKNIFE NT X1A 2N1	Document Number: MIC70300	
NORTHWEST TERRITORIES Health and Social Services Authority		Version No: 1.0	Page: 1 of 8
		Distribution:	
		Microbiology Instrumentation Manual	
Services Authority		Effective: 06 November, 2017	
Document Name:		Date Reviewed: 06 November, 2017	
BACTEC FX Instrument Procedures		Next Review: 06 November, 2017	
Approved By:		Status: APPROVED	
Jennifer G. Daley Bernier, A/ Manager, Laboratory Services			

PURPOSE:

Blood cultures are collected from patients with suspected sepsis or bacteremia. The isolation of any organism(s) from a blood culture must be considered significant and correlated with the clinical picture. Although primarily directed towards the processing of blood cultures, occasionally other specimen types (sterile fluids, abscess material, bone marrow, etc.) are received in blood culture vials. These bottles may be processed in the same manner as blood cultures.

The BACTEC FX instrument continuously monitors routine blood cultures for evidence of growth for 5 days. Negative results are auto verified as follows:

- No growth in 48 hours (preliminary)
- No growth in 5 days (final) •

SAMPLE INFORMATION:

Туре	Blood culture vial
Source	Blood or sterile fluid

REAGENTS and/or MEDIA:

	 BACTEC[™] Plus Aerobic/F Culture Vials (blue top) 	
Туре	 BACTEC[™] Lytic/10 Anaerobic/F Culture Vials (purple top) 	
	 BACTEC[™] Peds Plus[™]/F Culture Vials (pink top) 	
Source	BD	
Volume	1 vial	
Stability	Stable until date of expiration indicated on vial	
Storage	Bottle storage before blood collection:	
Requirements	Room temperature	
Critoria for rejection	Do not use if:	
and follow up action	The expiration date has passed	
and tonow up action	There are other signs of deterioration	

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SUPPLIES:

- BACTEC Plus Aerobic/F Culture Vials (blue top)
- BACTEC Lytic/10 Anaerobic/F Culture Vials (purple top)
- BACTEC Peds Plus™/F Culture Vials (pink top)
- BACTEC FX instrument

SPECIAL SAFETY PRECAUTIONS:

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potential infectious materials or cultures.

- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used when there is a known or potential risk of exposure of splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes and other sharp objects should be strictly limited.

All patient specimens are assumed to be potentially infectious. Universal precautions must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

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PROCEDURE INSTRUCTIONS:

Step	Action		
Loadi	ng Vials onto the BACTEC FX		
1	Open the drawer. Ensure that the STATUS screen is displayed and the barcode scanner turns on.		
2	Scan the BACTEC barcode first, then the LIS barcode.		
3	Place the vial into next available station that displays a solid green indicator.		
4	Scan and place any other bottles that need to be loaded.		
5	Close drawer when finished		
6	 Delayed entry of bottles may lead to delayed results. Best practices dictate that bottles should be placed into the BACTEC as soon as possible after collection. If a delay was identified: Visually inspect the bottle. If growth is apparent, treat as presumptively positive. Do not place bottle into the BACTEC. Subculture and make a smear for Gram-staining. 		
	 If the Gram-stain is negative and the bottle has been kept at room temperature for upt to 48 hours, it can be placed into the BACTEC. 		

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Step	Action			
Exten	Extending Incubation Time of Vials to 10 days			
	From the Status display on the BACTEC screen:			
1	Tap the "Drawer view" button			
	Select the desired station and tap "OK"			
	• Select "Modify" next to the Protocol field and change to 10 days, "Save".			
	From the LIS: Result Entry → Scan the order number to access the plate log/test comments:			
	• If a CXSET requires extended protocol (two vials under one order number), must click on the			
	AE Culture (CXBAE) in Test Comment area to populate the plate log below with the Aerobic			
	culture media and follow steps 3 & 4 below. THEN click on the AN culture (CXBAN) in the			
	Test Comment area to populate the plate log below with the Anaerobic culture media and			
	repeat steps 3 & 4.			
	In plate log (Media Comments), look for the "EXT" media ID.			
	M. Add Media M. Result Media M. Cancel Media 💮 Delete Media M. Media C			
2	1 EXT			
	Follow EXT keypad (instructions are written in the keypad):			
	• Double click in the box under "R" so a check mark appears, or click the "Result Media"			
	button (see below). This alerts the LIS to stop the 5 day reporting and change it to 10			
	day auto-reporting.			
	M. Add Media M. Cancel Media Ø Delete Media M Media Comments # Media ID Media Comment R 2 KT I IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII			
	 Save the culture to save these changes to the plate log 			
3	Repeat steps $1 \rightarrow 4$ for each vial.			
	If culture is negative:			
	A 48 hour negative preliminary report will be automatically released by LIS.			
4	 A 10 day no growth final report will automatically be released. 			
	Manual reporting negative bottles on extended protocol is not required.			

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	Positive vial:
	 Touch screen → Status display → Look for button on either Drawer A/ B (see example
	below; Positive Vial in Drawer A)
Red	
glow	 Open appropriate Drawer → Look for station(s) with a FLASHING Red LED then follow the
	steps below to resolve:
	a) Remove vial from station
	b) Scan bottle barcode
	c) Positive vials may be returned to BACTEC FX if no bacteria seen in Gram up to 5
	hours after the vial have been removed.
	Negative vial:
	 Touch screen → Status display → Look for button on either Drawer A/ B (see example
	below; Negative Vials in both Drawer A and B)
	Status Reports Maintenance Configuration
0	
Green	
glow	
	• Open appropriate Drawer \rightarrow Look for station(s) with a FLASHING Green LED then follow
	the steps below:
	a) Remove all vials from stations with flashing Green LED's
	b) Discard in a large yellow Biohazard sharps container

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lf:	Then:			
Identifyir	Identifying Anonymous Vial			
	Check the Status screen to see which Drawer houses the Anonymous Vial.			
	• Example below: 1 Anonymous Vial is in Drawer B, zero Anonymous Vials in Drawer A.			
1	Status Reports Maintenance Configuration No. 1 No. 1 No			
2	Open Drawer and locate station(s) with a flashing Yellow LED			
-	 If the station flashes YELLOW and RED, then the vial is also POSITIVE. 			
	Remove vial from station \rightarrow touch screen displays the message:			
3	"ID01: Positive- Anonymous pulled. Scan sequence (message truncated) remove".			
°	(Message instructs the user to scan the bottle barcode & accession number)			
	Barcode scanner activates automatically when vial is removed.			
	 Scan bottle barcode 1st. 			
	Scan accession/order number barcode			
4	 After scanning the barcodes, the user has two options: 			
	i. Place the bottle back into the BACTEC or			
	ii. Press the SAVE button on touch screen to remove bottle from BACTEC and clear the			
	station (DO THIS FOR POSITIVE VIALS).			
	If vial was flashing yellow ONLY:			
	Replace vial back into station (FLASHING Green LED)			
5	If vial was flashing yellow AND red:			
	Press the SAVE button on the screen (this removes the positive vial from BACTEC and			
	clears the station).			
7	If there are more flashing Yellow or Yellow/Red stations, repeat steps 2 \rightarrow 5			
	If there are no more Anonymous Vials \rightarrow close Drawer.			
8	Check Status screen. Question mark will be grayed out if no more Anonymous Vials are			
	detected			

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REFERENCES:

• BACTEX FX Instrument User Manual

REVISION HISTORY:

REVISION	DATE	Description of Change	REQUESTED BY
1.0	06-Nov-17	Initial Release	L. Steven

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