

<b>NORTHWEST TERRITORIES</b> <b>Health and Social Services Authority</b>	<b>Laboratory</b> <b>Stanton Territorial Hospital</b> P.O. Box 10, 550 Byrne Road YELLOWKNIFE NT X1A 2N1	<b>Document Number:</b> MIC30300	
		<b>Version No:</b> 5.0	Page: 1 of 7
<b>Document Name:</b> <b>Genital Culture – Bacterial Vaginosis</b>		<b>Distribution:</b> <b>Microbiology Culture Manual</b>	
<b>Approved By:</b> Jennifer G. Daley Bernier, A/manager, Laboratory Services		<b>Effective:</b> 30 January, 2017 <b>Date Reviewed:</b> 30 January, 2017 <b>Next Review:</b> 30 January, 2019	
		<b>Status:</b> <b>APPROVED</b>	

**PURPOSE:** To determine the presence or absence of Bacterial Vaginosis, Trichomonas vaginalis and Candida spp. in vaginal specimens.

**SAMPLE INFORMATION:**

<b>Type</b>	Swab <ul style="list-style-type: none"> <li>• Amie's swab</li> </ul>
<b>Source</b>	<ul style="list-style-type: none"> <li>• Posterior vaginal vault or vaginal orifice</li> <li>• Only performed on patients ≥ 13yrs of age</li> <li>• If specimen received on patient &lt;13yrs of age, process as Genital Culture</li> </ul>
<b>Stability</b>	<ul style="list-style-type: none"> <li>• Viability of Trichomonas vaginalis is significantly diminished if wet preps are not examined within 1h of collection.</li> <li>• If the sample is received in the laboratory greater than 24 hours from collection:             <ul style="list-style-type: none"> <li>➤ Add specimen quality comment: "Delayed transport may adversely affect pathogen recovery"</li> </ul> </li> <li>• If the sample is received in the laboratory and processed greater than 5 days from collection:             <ul style="list-style-type: none"> <li>➤ Add order comment: "Smear for Bacterial Vaginosis made at time of receipt not time of collection. This delay may alter the specimen results. Please interpret results with caution."</li> </ul> </li> </ul>
<b>Storage Requirements</b>	Room temperature
<b>Criteria for rejection and follow up action</b>	<ol style="list-style-type: none"> <li>1. Unlabeled/mislabeled swabs</li> <li>2. Dry swabs</li> </ol>

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**REAGENTS and/or MEDIA:**

- Sabouraud dextrose agar (SAB)

**SUPPLIES:**

- Microscope slides and coverslips
- Test tubes
- Sterile saline
- Biosafety cabinet
- Gram stain reagents
- Immersion oil
- 35° ambient air incubator

**SPECIAL SAFETY PRECAUTIONS:**

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potential infectious materials or cultures.

- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used when there is a known or potential risk of exposure of splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes and other sharp objects should be strictly limited.

All patient specimens are assumed to be potentially infectious. Universal precautions must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

**QUALITY CONTROL:**

Refer to Quality Control manual for reagent quality control procedures

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**PROCEDURE INSTRUCTIONS:**

Step	Action
<b>Processing vaginal swabs for Bacterial Vaginosis</b>	
<b>1</b>	<p><b>Double check patient age</b></p> <ul style="list-style-type: none"> <li>• If patient is &lt;13yrs of age, specimen should be processed as a genital culture. Refer to MIC30375.</li> <li>• If patient ≥13yrs of age proceed to step 2.</li> </ul>
<b>2</b>	<ul style="list-style-type: none"> <li>• <u>If Trichomonas is requested:</u> <ul style="list-style-type: none"> <li>➤ Perform microscopic examination for <i>Trichomonas vaginalis</i>.</li> </ul> </li> <li>• <u>If Genital culture is requested AND relevant clinical information is provided:</u> <ul style="list-style-type: none"> <li>➤ Perform direct smear and vaginal culture.</li> <li>➤ Relevant clinical information/diagnosis includes: Post-surgical, Toxic shock syndrome, pregnant, intrapartum, postpartum, post hysterectomy, pessary in place, D&amp;C.</li> </ul> </li> <li>• <u>If Genital culture is requested and NO relevant clinical information is provided:</u> <ul style="list-style-type: none"> <li>➤ Process as BV only and add order comment: “Routine genital cultures will not be performed unless testing is requested AND relevant clinical history is provided”.</li> </ul> </li> <li>• <u>If Yeast culture is requested AND relevant clinical information is provided:</u> <ul style="list-style-type: none"> <li>➤ Perform yeast culture.</li> <li>➤ Relevant clinical information/diagnosis includes: Chronic or recurrent vaginal candidiasis, treatment failure or immunosuppression</li> </ul> </li> <li>• <u>If Yeast culture is requested and NO relevant clinical information is provided:</u> <ul style="list-style-type: none"> <li>➤ Process as BV only and add order comment: “Routine yeast cultures will no longer be performed unless there is a clinical history of recurrent vaginal candidiasis, treatment failure and/or immunosuppression, particularly HIV infection. Microscopic examinations will continue for the detection of yeast and, if seen, will be reported”.</li> </ul> </li> </ul>

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<b>3</b>	<ul style="list-style-type: none"> <li>• <u>If Trichomonas is requested:</u> <ul style="list-style-type: none"> <li>➤ Label test tube with wet prep label.</li> <li>➤ In the biosafety cabinet add approximately 0.5 mL saline.</li> <li>➤ Label slide with patient’s last name, accession number and BV to identify test required.</li> <li>➤ Inoculate slide with swab, ensuring all surfaces of the swab make contact with the slide.</li> <li>➤ Place on hotplate to allow slide to dry.</li> <li>➤ If yeast is requested, inoculate SAB, streak for isolation and incubate at room temperature on the wound bench for 48 hours.</li> <li>➤ Place swab into wet prep tube and mix by flicking the tube. Incubate at 35° for 20 minutes.</li> </ul> </li> </ul>
<b>4</b>	<ul style="list-style-type: none"> <li>• <u>If Trichomonas is not requested,</u> <ul style="list-style-type: none"> <li>➤ Wet prep does not need to be performed.</li> <li>➤ Label slide with patient’s last name, accession number and BV to identify test required.</li> <li>➤ Inoculate slide with swab, ensuring all surfaces of the swab make contact with the slide.</li> <li>➤ Place on hotplate to allow slide to dry. If yeast is requested, inoculate SAB, streak for isolation and incubate at room temperature on the wound bench for 48 hours.</li> </ul> </li> </ul>
<b>5</b>	Allow smear to dry and perform gram stain. Refer to MIC20115 – Gram stain procedure.
<b>6</b>	If applicable, examine wet prep under 40x magnification as per MIC52700 – Wet Preparation using Saline.

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**INTERPRETATION OF RESULTS:**

Step	Action
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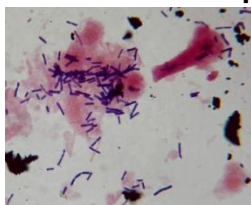
**Interpretation of Bacterial Vaginosis specimens**

1	<p><u>Trichomonas testing:</u></p> <ul style="list-style-type: none"> <li>• After incubation dab the culture swab onto the microscope slide then return the swab to its original transport tube.</li> <li>• Place the coverslip on the slide. Allow the slide to “settle”</li> <li>• Place the slide under the microscope. Examine under 40X for the presence of yeast and trichomonas.</li> </ul> <p>TIP: Decrease the light of a light microscope by closing the diaphragm or use a phase contrast microscope.</p>
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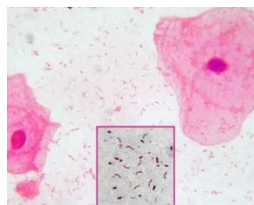
2	<p><u>Bacterial vaginosis testing:</u></p> <ul style="list-style-type: none"> <li>• Examine the gram stain under 10X magnification scan 5-10 fields and quantitate the number of epithelial cells, white blood cells and clue cells seen.</li> <li>• Add one drop of immersion oil to the slide and under 100X magnification scan 5-10 fields and quantitate the number of Lactobacilli, Gram variable bacilli and pleomorphic bacilli (Mobiluncus). Record numbers of Yeast and Trichomonas, if present.</li> <li>• Calculate the N-Score as in the table below:</li> </ul>
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Quantitation of bacterial morphotype	Points scored per morphotype				
	None	1+	2+	3+	4+
<b><i>Lactobacillus spp.</i></b> : med/large Gram positive rods	4	3	2	1	0
<b><i>Gardner Ella vaginalis</i></b> : Small Gram-negative or variable rods <b>Bacteroides</b> : Gram-negative rods	0	1	2	3	4
<b><i>Mobiluncus sp.</i></b> : Curved gram neg or –variable rods	0	1	1	2	2

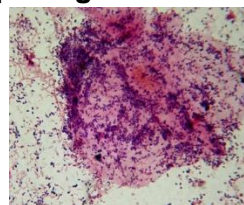
**Gram stain examples of relevant bacterial pathogens**



*Lactobacillus sp. vaginalis*



*Mobiluncus sp.*



*Gardnerella vaginalis*



*Trichomonas*

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**REPORTING OF RESULTS:**

If:	Then:								
<b>Trichomonas reporting:</b>									
Trichomonas seen (parasite must be motile)	<ul style="list-style-type: none"> <li>In SCWPT select <b>“Trich – Seen”</b></li> <li>Finalize SCWPT</li> </ul>								
Trichomonas not seen	<ul style="list-style-type: none"> <li>If sample is &lt; 24 hours old, at SCWPT select: <b>“Trichomonas – Not seen.”</b></li> <li>If sample &gt;24h and ≤ 72h old select: <b>“Trichomonas - Not Seen</b> <b>NOTE: The presence of Trichomonas cannot be ruled out since there was a delay &gt;24hrs in transport and/or processing of this specimen.”</b></li> <li>If sample &gt;72h old, specimen is unsuitable for Trichomonas analysis and the test needs to be cancelled. Click on <b>Cancel Test</b> and insert cancelation comment <b>TRICH:</b> <b>“Specimen in transit greater than 72h. Vaginal swabs should be submitted to the laboratory optimally within 24 hours for Trichomonas examination. Unsuitable for Trichomonas analysis.”</b></li> </ul>								
<b>Bacterial vaginosis reporting:</b>									
Report BV Smear results by N-Score	<table border="1"> <thead> <tr> <th style="background-color: #00b050; color: white;">N-SCORE</th> <th style="background-color: #00b050; color: white;">REPORT</th> </tr> </thead> <tbody> <tr> <td>0 – 3</td> <td><b>Smear NEGATIVE for Bacterial Vaginosis</b></td> </tr> <tr> <td>4 - 6</td> <td> <b>Gram stain shows altered vaginal flora. Results are indeterminate for Bacterial Vaginosis.</b>                      ➤ If clue cells are present, report:  <b>Presence of Clue cells suggests transition of vaginal flora towards Bacterial Vaginosis. Repeat testing of another vaginal smear is recommended.</b> </td> </tr> <tr> <td>7 – 10</td> <td><b>Smear consistent with BACTERIAL VAGINOSIS</b></td> </tr> </tbody> </table>	N-SCORE	REPORT	0 – 3	<b>Smear NEGATIVE for Bacterial Vaginosis</b>	4 - 6	<b>Gram stain shows altered vaginal flora. Results are indeterminate for Bacterial Vaginosis.</b> ➤ If clue cells are present, report: <b>Presence of Clue cells suggests transition of vaginal flora towards Bacterial Vaginosis. Repeat testing of another vaginal smear is recommended.</b>	7 – 10	<b>Smear consistent with BACTERIAL VAGINOSIS</b>
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Add comments if appropriate	Patient is >55 yrs.	<b>“Results may not be reliable in post-menopausal women. Correlate with the clinical picture”.</b>
	Yeast cells/pseudo hyphae seen.	<b>“Yeast Seen. Candida species are normal flora in the genital area of 30-40% of women. The presence of yeast must be correlated with clinical picture”</b>
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	3+ or 4+ WBC's	<b>“Purulence suggests the presence of another infection and/or inflammatory condition. Correlate with the clinical picture. Testing for <i>N. gonorrhoeae</i> and <i>C. trachomatis</i> may be indicated”.</b>
	Multiple anaerobic organisms seen	<b>“The presence of mixed anaerobes seen intracellularly may be more compatible with cervicitis or pelvic inflammatory disease than bacterial vaginosis. Clinical correlation required.”</b>
	Scant or no cells, or excess amorphous material	<b>“Insufficient sample to assess for vaginitis. Recollection required”.</b>

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**REFERENCES:**

- Clinical Microbiology Procedures Handbook, 4<sup>th</sup> edition, ASM Press, 2016
- Jorgensen J.H., Pfaller M.A., Carroll K.C., Funke G., Landry M.L., Richter S.S., Warnock D.W. 2015. Manual of Clinical Microbiology, 11<sup>th</sup> edition, ASM Press, Washington, D.C.

**REVISION HISTORY:**

<b>VERSION</b>	<b>DATE</b>	<b>Description of Change</b>	<b>REQUESTED BY</b>
1.0	03-Dec-2010	Initial Release	M-L Dufresne
2.0	30-Apr-2012	Update and Review	C.Russell
3.0	01-Mar-2013	Revision	S.Webber
4.0	21-Oct-2013	LIS updates/document number change	A.Darrach
5.0	06-Dec-2016	Update to reflect pre-analytical and analytical requirements	L.Steven

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