	Stanton Territorial Hospital	Document Number: MIC10315	
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Health and Social Services Authority	YELLOWKNIFE NT X1A 2N1	Distribution:	
		Microbiology Specimen Processing	
Services Authority		Effective: 12 May, 2017	
Document Name: Water Testing – HPC Unit Dose SimPlate		Date Reviewed: 12 May, 2017	
Method		Next Review: 12 May, 2019	
Approved By: Jennifer G. Daley Bernier, A/Manager, Laboratory Services		Status: APPROVED	

PURPOSE: The presence of heterotrophic bacteria in dialysate waters can lead to the development of a Gram negative toxin mediated pyrogenic reaction, bacteremia, and chronic inflammatory response syndrome. Hot tub waters are screened for the presence of *Pseudomonas aeruginosa*, the causative agent of a superficial skin infection known as "Hot tub folliculitis". The HPC SimPlate method utilizes enzyme technology to target the most common enzymes of waterborne bacteria. The by-product of the enzymatic reactions can be seen as fluorescence using a UV light. Additionally, hot tub waters have a MacConkey plate inoculated to aid in the detection of *Pseudomonas aeruginosa*.

SAMPLE INFORMATION:

Туре	Hot tub water, Dialysate water	
Volume	10mL +/- 0.2mL	
Storage	Refrigerated	
Requirements	Reingerated	
Criteria for rejection	<10 mL water received	
and follow up action	Received >48 hours after collection	

REAGENTS and/or MEDIA:

Туре	SimPlate for HPC kit from IDEXX	
Storage	Store at 2-30°C away from light	
Requirements	Expiry date is printed on the box of media tubes	

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SUPPLIES:

10mL syringe

• UV light

• 35°C incubator

SPECIAL SAFETY PRECAUTIONS:

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potential infectious materials or cultures.

- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used when there is a known or potential risk of exposure of splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes and other sharp objects should be strictly limited.

All patient specimens are assumed to be potentially infectious. Universal precautions must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

QUALITY CONTROL:

Performed per shipment or lot number:

- Refer to QC Procedure MIC60500 Water quality control.
- A TQC order is automatically generated when new kit is entered into TQC to record the QC results.

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PROCEDURE INSTRUCTIONS:

Action
rming HPC Water Testing
Accession waters and select source. If source is HOT TUB or SPA WATER – HPC
will automatically order and a MacConkey plate label will generate. Labels for total
coliform testing will also print out for these waters.
Label requisition and sample containers.
Label sterile media tube (green top), SimPlate and MAC plate (where required)
Mix water. Use a 10 mL syringe to remove 10 mL of water from sample container.
Add to green top tube and shake. Allow powder in tube to dissolve
Remove SimPlate lid and pour contents of green top tube onto the center of the plate
base
Replace the lid and gently swirl to distribute the sample. Note: air bubbles do not
interfere with test
Tip the plate at a 90° angle so the excess water will drain into the absorbent pad at the
bottom of the plate
Invert the plate onto the plastic lid. On lid write R: (Date + 2 day) and time incubated
If water source is HOT TUB, perform total coliform testing as well using the Colilert-18
kit and processed as above.
If water is HOT TUB, aliquot off an additional 1mL of water and flood a labeled MAC
plate - incubate along with the SimPlate (35°C for 48hrs)

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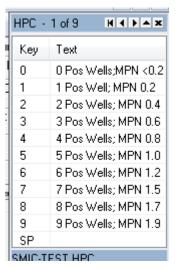
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INTERPRETATION AND REPORTING:

While viewing under the **UV lamp** - count the number of wells showing fluorescence.

Using the HPC keypad, select the number of wells that show fluorescence. This will automatically calculate the MPN (most probably number) of heterotrophic plate count bacteria in the original sample.



Report this number as the HPC.

HOT TUB/SPA WATERS:

3

2

- Look for growth of Pseudomonas aeruginosa Non-lactose fermenter, oxidase positive – send to VITEK for GNI
- Report as: "Isolated: Pseudomonas aeruginosa"
- If MAC plate is negative- Report as: "No Pseudomonas aeruginosa isolated"

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APPENDIX:

Unit-Dose SimPlate For HPC Most Probable Number (MPN) Table

# Positive	MPN	95% confidence limits	
Wells		lower	upper
0	<0.2	< 0.03	<1.4
1	0.2	0.0	1.4
2	0.4	0.1	1.6
3	0.6	0.2	1.9
4	0.8	0.3	2.2
5	1.0	0.4	2.5
6	1.2	0.6	2.7
7 8	1.5	0.7 0.8	3.0 3.3
9	1.7 1.9	1.0	3.5
10	2.1	1.1	3.9
11	2.1	1.1	4.2
12	2.6	1.5	4.5
13	2.8	1.6	4.8
14	3.0	1.8	5.1
15	3.3	2.0	5.4
16	3.5	2.2	5.8
17	3.8	2.3	6.1
18	4.0	2.5	6.4
19	4.3	2.7	6.7
20	4.5	2.9	7.0
21	4.8	3.1	7.4
22 23	5.1	3.3	7.7 8.0
24	5.3 5.6	3.5 3.8	8.4
25	5.9	4.0	8.7
26	6.2	4.2	9.1
27	6.5	4.4	9.4
28	6.8	4.7	9.8
29	7.1	4.9	10.2
30	7.4	5.1	10.6
31	7.7	5.4	10.9
32	8.0	5.6	11.3
33	8.3	5.9	11.7
34	8.6	6.2	12.1
35	9.0	6.4	12.6
36 37	9.3 9.7	6.7	13.0 13.4
38	10.0	7.0 7.3	13.4
39	10.4	7.6	14.3
40	10.8	7.9	14.8
41	11.2	8.2	15.2
42	11.6	8.5	15.7

# Positive	MPN	95% confidence limits	
Wells		lower	upper
43	12.0	8.8	16.2
44	12.4	9.1	16.7
45	12.8	9.5	17.3
46	13.2	9.8	17.8
47	13.7	10.2	18.3
48	14.1	10.6	18.9
49	14.6	10.9	19.5
50	15.1	11.3	20.1
51	15.6	11.7	20.7
52	16.1	12.1	21.3
53	16.6	12.5	22.0
54	17.1	13.0	22.7
55	17.7	13.4	23.4
56	18.3	13.9	24.1
57	18.9	14.4	24.9
58	19.5	14.9	25.7
59	20.2	15.4	26.5
60	20.9	15.9	27.3
61	21.6	16.5	28.2
62	22.3	17.1	29.2
63 64	23.1 23.9	17.7 18.3	30.2 31.2
65	23.9	19.0	32.3
66	25.7	19.7	33.5
67	26.6	20.4	34.7
68	27.6	21.2	36.1
69	28.7	22.0	37.5
70	29.9	22.9	39.0
71	31.1	23.8	40.7
72	32.4	24.8	42.5
73	33.9	25.8	44.4
74	35.5	27.0	46.6
75	37.2	28.2	49.1
76	39.2	29.6	51.9
77	41.4	31.1	55.1
78	44.0	32.8	58.9
79	47.0	34.8	63.6
80	50.7	37.1	69.5
81	55.5	39.8	77.5
82	62.3	43.2	89.9
83	73.8	47.6	114.6
84	>73.8	>47.6	>114.6

MPN is per ml of the 10 ml sample added to the media tube (pour-off is accounted for).

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REFERENCES:

 SimPlate for HPC Water Testing Product Information. (n.d.). Retrieved from www.idexx.ca/view/xhtml/en_ca/water/simplate.jsf.

REVISION HISTORY:

REVISION	DATE	Description of Change	REQUESTED BY
1.0	31Dec2013	Initial Release	A.Darrach
1.1	31March2016	Reviewed – No Changes	C. Russell
2.0	12-May-2017	Reviewed and revised. Safety precautions and reagent storage requirements added; Updated format; New document number (old number MIC52615)	L. Steven

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