NORTHWEST TERRITORIES Health and Social	Stanton Territorial Hospital P.O. Box 10, 550 Byrne Road YELLOWKNIFE NT X1A 2N1	Document Number: MIC53400	
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		Distribution:	
		Microbiology Test Manual	
Services Authority		Effective:	
Document Name: A	erotolerance Test	Date Reviewed:	
		Next Review:	
Approved By:		Status: DRAFT	

PURPOSE: The aerotolerance test is used to designate an organism as an anaerobe.

SAMPLE INFORMATION:

Туре	Organisms growing on anaerobic culture media and not present on
туре	aerobic culture media.

REAGENTS and/or MEDIA:

- Chocolate agar
- Brucella agar

SUPPLIES:

- Disposable inoculation needles
- Glass microscope slides
- Anaerobic trays or jars
- Anaerobic packs and indicators
- 35° ambient air and 37° CO₂ incubators

SPECIAL SAFETY PRECAUTIONS:

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potentially infectious materials or cultures.

- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used when there is a known or potential risk of exposure to splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes and other sharp objects should be strictly limited.

All patient specimens are assumed to be potentially infectious. Universal precautions must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

QUALITY CONTROL:

Refer to MIC60010 – Microbiology Quality Control for anaerobic tray and jar quality control.

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PROCEDURE INSTRUCTIONS:

Step	Action				
Perfo	Performing the aerotolerance test				
1	Subculture a single, well-isolated colony of each morphological type suspected to be				
	an anaerobe. Using one colony ensures that the same organism goes onto both agar				
	plates and the slide.				
2	Touch each colony with a loop or sterile stick and subculture to Chocolate agar,				
	Brucella agar and a labelled, glass microscope slide.				
3	The Chocolate agar should be inoculated first, so that if only the Brucella agar plate				
	grows there is no question of not having enough organisms to initiate growth.				
4	Incubate the Chocolate agar in CO_2 incubator for 24 hours. If no growth is seen, re-				
	incubate for an additional 24 hours.				
5	Incubate the Brucella agar in anaerobic jar or tray for 48 hours along with original				
	anaerobic media.				

INTERPRETATION OF RESULTS:

IF	THEN
Isolate grows on Chocolate agar at	Organism is not a strict anaerobe and can be
24 or 48 hours	identified using routine methods.
AND	
Isolate grows on Brucella agar at	
24 or 48 hours.	
Isolate does not grow on Chocolate	Organism is a strict anaerobe.
agar at 24 or 48 hours	
AND	
Isolate does grow on Brucella agar at	
24 or 48 hours.	
Isolate does not grow on Chocolate	Test is inconclusive:
agar at 24 or 48 hours	Reasons: organism died or media not
AND	inoculated.
Isolate does not grow on	Repeat aerotolerance testing with colony on
Brucella agar at 24 or 48 hours.	original anaerobic media.

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LIMITATIONS:

- 1. Chocolate agar should be used for aerotolerance testing. *Haemophilus* spp. will grow anaerobically on Brucella agar and therefore will be mistaken for anaerobic Gramnegative rods if Chocolate agar is not used.
- 2. Some authorities have suggested performing aerotolerance testing in multiple environments (ambient air, CO₂, a microaerophilic environment) on problem isolates. This is not necessary for identification of the most commonly isolated anaerobes, but it may be considered for isolates when their exact atmospheric requirement is difficult to determine using the above procedure.
- 3. If anaerobic organism processing occurs on the open bench, all plates should be promptly incubated anaerobically as some clinical isolates may die after relatively short exposure to oxygen.

REFERENCES:

- Clinical Microbiology Procedures Handbook, 4th edition, ASM Press, 2016.
- Jorgensen J.H., Pfaller M.A., Carroll K.C., Funke G., Landry M.L., Richter S.S., Warnock D.W. 2015. Manual of Clinical Microbiology, 11th edition, ASM Press, Washington, D.C.

REVISION HISTORY:

REVISION	DATE	Description of Change	REQUESTED BY
1.0	23 MAR 19	Initial Release	L. Steven

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