Stanton Terri	Stanton Territorial Hospital	Document Number: MIC52400	
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NORTHWEST TERRITORIES		Distribution: Microbiology Test Manual	
Health and Social YELLOWKNIFE NT X1A 2N1 Services Authority	TELEOWKINIFE INT XIA ZINI		
	Effective:		
Document Name: Urea Test Approved By:		Date Reviewed:	
		Next Review:	
		Status: DRAFT	

**PURPOSE:** The urea test is used to determine the ability of an organism to split urea by the action of the enzyme urease forming two molecules of ammonia with resulting alkalinity.

## **SAMPLE INFORMATION:**

Туре	One, well isolated colony

## **REAGENTS and/or MEDIA:**

• Urea tube

## **SUPPLIES:**

- Disposable inoculation needles
- 35° ambient air incubator

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## **SPECIAL SAFETY PRECAUTIONS:**

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potentially infectious materials or cultures.

- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used where there is a known or potential risk of exposure to splashes.
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes, and other sharp objects should be strictly limited.

All patient specimens are assumed to be potentially infectious. Universal precautions must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

## **QUALITY CONTROL:**

- Inspect the media for cracks, freezing, contamination and bubbles when received into the laboratory. Refer to MIC60090 – Entering New Media and Reagents into TQC for recording media acceptance.
- This medium is considered exempt as per CLSI document M22-A3.

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# **PROCEDURE INSTRUCTIONS:**

Step	Action				
Perfo	Performing the urea test				
1	Remove the urea tube from the refrigerator and bring to room temperature.				
2	Using a disposable inoculating needle, touch the center of a well-isolated colony.				
3	Inoculate the agar slant surface. Do not stab the butt.				
4	Loosely replace the cap on the tube. DO NOT tighten the cap.				
5	Incubate in the O <sub>2</sub> incubator.				
6	Check for color change to bright pink after 3 hours and again at 18 to 24 hours. For				
	slower growing organisms such as <i>Nocardia</i> , the color change can take up to 7 days.				

## **INTERPRETATION OF RESULTS:**

IF	THEN	
Intense pink or red colour	Urea = Positive	
No colour change or pale yellow colour	Urea = Negative	
Uninoculated or Negative		

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#### PROCEDURE NOTES:

- The urea test is used as part of the identification of several genera and species of Enterobacteriaceae, including *Proteus, Klebsiella* and some *Yersinia* and *Citrobacter* species, as well as some *Corynebacterium* species, *Brucella, Helicobacter pylori* and many other bacteria that produce the urease enzyme.
- The test can also be used as part of the identification of *Cryptococcus* species.

## LIMITATIONS/PRECAUTIONS:

- 1. *Proteus* spp. will show positive reactions within 1 to 6 hours.
- 2. The cap of the urea tube must not be tightened.
- 3. Urea-positive, oxidase-positive, Gram-negative coccobacilli that are isolated from the urinary tract may be *Oligella ureolytica*.
- 4. Some organisms rapidly split urea (Brucella and H.pylori) while others react slowly.
- 5. When performing overnight tests from medium that contains peptone, the alkaline reaction may be due not to urease but to hydrolysis of peptone.
- 6. Urea is light sensitive and can undergo auto hydrolysis. Store at 2°C to 8°C in the dark.
- 7. Urea-positive, oxidase-positive, Gram-negative coccobacilli that do not grow on MacConkey agar in 24 hours are presumptively identified as *Brucella*, unless they are isolated from urine. *Immediately transfer cultures to a biosafety cabinet.*

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#### **REFERENCES:**

- Oxoid Urea Agar product details, 12/30/2018
- Clinical Microbiology Procedures Handbook, 4<sup>th</sup> edition, ASM Press, 2016
- CLSI. Quality Control for Commercially Prepared Microbiological Culture Media; Approved Standard—Third Edition. CLSI document M22-A3. Wayne, PA: Clinical and Laboratory Standards Institute; 2004

## **REVISION HISTORY:**

REVISION	DATE	Description of Change	REQUESTED BY
1.0	8 APR 19	Initial Release	L. Steven

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