

The specimen should be collected into a clean, sterile, leak-proof centrifuge tube:

- Tube 2 is for microbiology culture
 - Tube 4 is for viral testing if requested
 - CSF samples should be:

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- Immediately transported to the laboratory
 - Should NOT be refrigerated
 - Insufficient volume for tests requested: contact the physician to prioritize requests
 - Leaking specimens should be processed. Add comment to order entry
- Improperly collected, labeled, transported, or handled specimens should be processed. Waver of responsibility form SCM40110 needs to be filled out by the responsible nurse
 - Accession the sample in SoftMic in Order Entry with test code "CXCSF"
 - In the Micro tab, fill in the Collected, Received and Plated rows and save
 - Media labels print out in the order of media inoculation:

1:BA-C=**Blood**, 2:CHO-C=**Chocolate**, 3:MAC-0=**MacConkey**, 4:CONC=**Supernatant**

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FILENAME: MIC35000-After-Hours CSF Culture Setup Job Aid

		Document Name:	Document Number:
	Stanton Territorial Hospital	After-Hours CSF Culture Setup Job Aid	MIC35000
NORTHWEST TERRITORIES Health and Social	P.O. Box 10, 550 Byrne Road YELLOWKNIFE NT X1A 2N1		
Services Authority		Distribution:	Date Issued:
		Microbiology Culture Manual	January 31, 2022

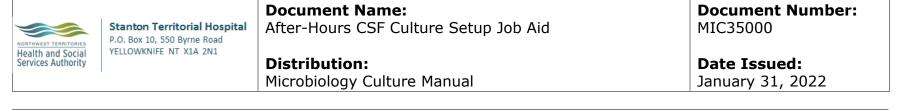
5	>1mL received	Centrifuge tube #2 at 3500 rpm for 10 minutes			
		 Transfer the supernatant to the CONC tube and discard 			
		• Mix remaining supernatant with sediment in tube and use to plate			
		and prepare the smear			
	<1mL received	Do not centrifuge			
		 Plate and prepare smear using the un-spun sample 			
	Inoculate media and slides				
	• Label two slides using a pencil with the accession number, patient's last name, and				
	"CSF"				

- In the biosafety cabinet, using a sterile pipette, aspirate fluid from the bottom of the sediment tube or collection tube
- Place 1 to 2 drops onto BA, CHOC and MAC. Streak for isolated growth using a disposable inoculation needle. Streak out to cover the whole plate
- Prepare smears by placing 1 or 2 drops of CSF on microscope slides. Allow the drop(s) to form one large drop. Do not spread the fluid
- Shunt fluids should be also planted to THIO broth

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After plating:

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- Place the BA and CHO plates in the top CO₂ incubator
- Place in the white tray on the bottom shelf labelled CSF
- Place MAC and sample tube in the O₂ incubator
 - > Place in the white tray on the middle shelf labelled CSF
- Close BSC and allow smears to dry in the BSC
- **9 Note:** The reading of CSF smears is to be done by a Microbiology Technologist
 - Leave requisition on the front bench of the microbiology lab

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