PROGRAM Standard Operating Procedure – Laboratory Services				
Title: MIC20130 – Kinyoun Stain	Policy Number:			
Program Name: Laboratory Services				
Applicable Domain: Lab, DI and Pharmacy Services				
Additional Domain(s):				
Effective Date:	Effective Date:			
Issuing Authority:	Date Approved:			
Director, Health Services				
Accreditation Canada Applicable Standard: N/A				

### **GUIDING PRINCIPLE:**

The kinyoun stain is used in the microscopic detection of acid-fast microorganisms such as Mycobacterium. Acid-fast organisms such as Mycobacterium have cell walls that are resistant to conventional staining by aniline dyes such as the Gram stain. Nonmycobacterial organisms with various degrees of acid-fastness include Rhodococcus species, Nocardia species, Legionella micdadei, and the cysts of Cryptosporidium, Isospora, Cyclospora and microsporidia.

### **PURPOSE/RATIONALE:**

This standard operating procedure describes how to perform the kinyoun stain.

## SCOPE/APPLICABILITY:

This procedure applies to Medical Laboratory Technologists (MLTs) performing the kinyoun stain.

### SAMPLE INFORMATION:

TypeSputum samples with STAT AFB stain requested

## **REAGENTS and/or MEDIA:**

- Kinyoun Carbol-Fuchsin
- Carbol Fuchsin Decolorizer
- Carbol Fuchsin Counterstain (Methylene Blue)

### SUPPLIES:

- Glass microscope slide
- QC slide
- Immersion oil
- Slide storage tray

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## EQUIPMENT

• Microscope

## **SPECIAL SAFETY PRECAUTIONS:**

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potential infectious materials or cultures.

- Ensure that appropriate hand hygiene practices be used.
- Lab gown must be worn when performing activities with potential pathogens.
- Gloves must be worn when direct skin contact with infected materials is unavoidable.
- Eye protection must be used when there is a known or potential risk of exposure of splashes.
- All procedures that may produce aerosols or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC).
- The use of needles, syringes and other sharp objects should be strictly limited.

All patient specimens are assumed to be potentially infectious. Routine Practices must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

# **QUALITY CONTROL:**

- Quality control is performed as tested
- A TQC order is automatically generated when test is ordered to record the QC results
- Refer to MIC60060-Microbiology Stain Quality Control

# **PROCEDURE INSTRUCTIONS:**

Step	Action				
Perfo	Performing the kinyoun stain				
1	Prepare a smear of the specimen to be stained. Refer to MIC10000- Microbiology Specimen Handling for slide preparation instructions.				
2	Place slide on the slide warmer in the BSC until dry.				
3	Flood the entire slide with Kinyoun Carbol Fuchsin Stain for 2 minutes and rinse thoroughly with water.				
4	Flood the slide with Carbol Fuchsin Decolorizer and decolorize until no more color drains from the slide (approx. 3 to 5 seconds). Rinse the slide thoroughly with water and shake off any excess moisture.				
5	Flood the slide with Carbol Fuchsin Counterstain (Methylene Blue) and allow the slide to stain for 30 seconds. Rinse thoroughly with water and allow to air dry. Do not blot.				
6	Examine the smear microscopically under a 100x oil immersion objective.				

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### **INTERPRETATION OF RESULTS:**

Step	Action
1	<ul> <li>Acid-fast mycobacteria will appear as dark pink to red bacilli</li> <li>Background and non-acid fast organisms will appear blue</li> </ul>
2	When a carbol fuchsin smear is read a minimum of 300 fields should be examined before the smear is reported as negative

## **REPORTING INSTRUCTIONS:**

IF	REPORT			
	<ul> <li>Result the test using the "STKIN" keypad</li> <li>Select Key N }AFBN (No AFB seen)</li> <li>III Tests (1)  Isolates (0)  Isolates (0)</li></ul>			
No Acid-Fast Bacilli seen on Kinyoun- stained smear	Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: Cancel Test     Image: Cancel Test       Image: Add Test     Image: Cancel Test     Image: C			
	2 )AFB2 (1-9/10 field) 3 )AFB3 (1-9/field) 4 )AFB4 (>9/field)			
<ul> <li>Result the test using the "STKIN" keypad</li> <li>Select Key 1 if 1-9 AFB seen per 100 fields</li> <li>Select Key 2 if 1-9 AFB seen per 10 fields</li> <li>Select Key 3 if 1-9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> <li>Select Key 4 if &gt;9 AFB seen per field</li> </ul>				
stained smear	#         TestID           1         STKIN			

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### NOTE:

- Refer to Reportable Diseases Public Health Act as of September 2009 for reporting to HPU1
- Refer to LQM70620-Laboratory Critical Results List-Microbiology for results that need to be phoned to ordering location
- Refer to MIC36100-Nosocomial Infection Notification Job Aid to determine if organism needs to be copied to Infection Prevention and Control
- Refer to MIC36200-Referral of Category A Specimens to APL for sending category A isolates to APL

### LIMITATIONS:

- 1. Rapidly growing mycobacteria may vary in their ability to retain acid-fast dyes and may fail to stain
- 2. Be aware of adequate safety precautions and procedures required when handling specimens that are submitted for mycobacterial evaluation
- 3. Mycobacterial staining should always be used as an adjunct to culture methods since culture techniques are much more sensitive than all acid-fast staining procedures

## **CROSS-REFERENCES:**

- 17-02-V1: Specimens Containing Suspected Risk Group 3 Pathogens
- LQM70620-Laboratory Critical Results List-Microbiology
- MIC10000-Microbiology Specimen Handling
- MIC36100-Nosocomial Infection Notification Job Aid
- MIC36200-Referral of Category A Specimens to Alberta Precision Laboratories
- MIC60060-Microbiology Stain Quality Control

## **REFERENCES:**

- 1. Clinical Microbiology Procedures Handbook, 4<sup>th</sup> edition, ASM Press, 2016
- 2. Dalynn Biologicals. Kinyoun Carbol Fuchsin Stain package insert, 2014

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# **APPROVAL:**

Date

#### **REVISION HISTORY:**

REVISION	DATE	Description of Change	REQUESTED BY
1.0	07 Feb 17	Initial Release	L. Steven
2.0	31 Mar 22	Procedure reviewed and added to NTHSSA policy template	L. Steven

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