Title: MIC31100-VRE Screen Type: Laboratory Services Program SOP Policy Number:

Issuing Authority: Director, Health Services

Next Review Date: Date Approved:

> PROGRAM Standard Operating Procedure – Laboratory Services Title: MIC31100 - VRE Screen Policy Number: Program Name: Laboratory Services Applicable Domain: Lab, DI and Pharmacy Services Additional Domain(s): Next Review Date: Effective Date: Date Approved: Issuing Authority: Director, Health Services Accreditation Canada Applicable Standard: N/A

#### **GUIDING PRINCIPLE:**

Specimens are submitted to identify carriers of vancomycin-resistant Enterococcus faecium and/or Enterococcus faecalis (VRE). Swabs may be submitted from any body site (other than nasal and axilla), but most commonly are collected from the rectum and are used to aid in the prevention and control of VRE in healthcare settings

## **PURPOSE/RATIONALE:**

This standard operating procedure describes the screening for Vancomycin Resistant Enterococci (VRE) on admission and as part of Multi-Resistant Organism (MRO) screens.

## SCOPE/APPLICABILITY:

This procedure applies to Medical Laboratory Technologists (MLTs) processing specimens for VRE screen.

#### **SAMPLE INFORMATION:**

Туре	<ul><li>Swab</li><li>Amie's with or without charcoal</li></ul>		
Source	<ul><li>Rectum</li><li>Stool</li><li>MRO screen: any site</li></ul>		
Stability	<ul> <li>If the sample is received in the laboratory and processed greater than 48 hours from collection:</li> <li>Add specimen quality comment: "Delayed transport may adversely affect pathogen recovery"</li> </ul>		
Storage Requirements	Room temperature		

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Policy Number: Date Approved: Page 1 of 7 Title: MIC31100-VRE Screen

Issuing Authority: Director, Health Services

Next Review Date:

Type: Laboratory Services Program SOP
Policy Number:
Date Approved:

Criteria for	2.	Specimen container label does not match patient identification on requisition
rejection	3.	Duplicate specimens obtained with same collection method from same collection location within 24 hours

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## **REAGENTS and/or MEDIA:**

- VRESelect agar (VRE), Blood agar (BA) and Mueller Hinton agar (MH)
- Identification reagents: gram stain, catalase and vancomycin E-test

## **SUPPLIES:**

- Disposable inoculation needles
- Wooden sticks

## **EQUIPMENT:**

- Biosafety cabinet
- 35° ambient air incubator
- Microscope
- Vitek 2 and supplies

#### **SPECIAL SAFETY PRECAUTIONS:**

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potentially infectious materials or cultures:

- Ensure that appropriate hand hygiene practices be used
- · Lab gown must be worn when performing activities with potential pathogens
- Gloves must be worn when direct skin contact with infected materials is unavoidable
- Eye protection must be used when there is a known or potential risk of exposure of splashes
- All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC)
- The use of needles, syringes and other sharp objects should be strictly limited

All patient specimens are assumed to be potentially infectious. Routine Practices must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

#### **QUALITY CONTROL:**

- Refer to MIC60040-Culture Media Quality Control procedure
- Refer to Test Manual for reagent quality control procedures

# **PROCEDURE INSTRUCTIONS:**

Step	Action
Proce	essing swabs for VRE screen
1	Monday to Sunday VRF swabs are processed by 15:00

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Policy Number: Date Approved: Page 2 of 7

Title: MIC31100-VRE Screen

Issuing Authority: Director, Health Services

Next Review Date:

Type: Laboratory Services Program SOP
Policy Number:
Date Approved:

2	<ul> <li>In the biosafety cabinet:</li> <li>Inoculate VRESelect agar with the swab</li> <li>Ensure all surfaces of the swab make contact with the agar</li> <li>Streak for isolated growth using a disposable inoculation needle</li> </ul>
3	Label the VRE plate with: R (Date + 2 days).
4	Incubate the media:  • Place VRE in the O₂ incubator in appropriate tray

#### **INTERPRETATION OF RESULTS:**

TILLEVI	PRETATION OF RESULTS			
Step	Action			
1	<ul><li>Observe VRE plate at 1</li><li>Examine for blue or pire</li></ul>	18 to 24 hours and 38 to 48 hours nk colonies		
2	<ul> <li>Record observations in</li> </ul>	are seen at 18 to 24 hours: the LIS introduction in the "Old urine culture" shelf		
3	<ul> <li>If no blue or pink colonies are seen at 38 to 48 hours:</li> <li>Record observations in the LIS</li> <li>Workup complete, VRE not isolated</li> </ul>			
	<ul><li>If blue colonies are seen:</li><li>Record observations in the LIS</li><li>Perform catalase test:</li></ul>			
	IF	THEN		
	Catalase POSITIVE	<ul> <li>Record observations in the LIS</li> <li>Workup complete, VRE not isolated</li> <li>Re-incubate plate for additional 38 to 48 hours if required</li> </ul>		
4	Catalase NEGATIVE	<ul><li>Record observations in the LIS</li><li>Subculture to BA plate</li></ul>		
	From the BA sub plate:  • Perform gram stain			
	IF THEN			
	Gram stain NOT GPC	<ul> <li>Record observations in the LIS</li> <li>Workup complete, VRE not isolated</li> </ul>		
	Gram stain GRAM POSITIVE COCCI	<ul><li>Perform GPI</li><li>Set up vancomycin E-test</li></ul>		
	If pink colonies are seen: Record observations in Subculture colonies to From the BA sub plate	the LIS BA sub plate		
5	IF	THEN		
	Gram stain NOT GPC Gram stain	<ul> <li>Record observations in the LIS</li> <li>Workup complete, VRE not isolated</li> <li>Perform GPI</li> </ul>		
	GRAM POSITIVE COCCI	<ul><li>Perform GPI</li><li>Set up vancomycin E-test</li></ul>		

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Policy Number: Date Approved: Page 3 of 7

Title: MIC31100-VRE Screen

Issuing Authority: Director, Health Services

Next Review Date:

Type: Laboratory Services Program SOP

Policy Number: Date Approved:

## **REPORTING INSTRUCTIONS:**

IF	REPORT		
No Blue or Pink colonies	Report: "No Vancomycin Resistant Enterococci (VRE) isolated"		
Blue and/or Pink colonies - Vitek ID: E.gallinarum and/or E.casseliflavus	<ul> <li>Verify the organism ID</li> <li>Suppress GPI result in the isolates tab:         <ul> <li>Change the Isolate # to a letter</li> <li>Verify the result</li> </ul> </li> <li>Enter and verify vancomycin E-test result</li> <li>Keep vancomycin E-test result suppressed</li> <li>Report: "No Vancomycin Resistant Enterococci (VRE) isolated"</li> </ul>		
	IF	REPORT	
Blue and/or Pink colonies - Vitek ID: E.faecalis and/or E.faecium	Vancomycin E-test MIC = <4 μg/mL	<ul> <li>Verify the organism ID</li> <li>Suppress GPI result in the isolates tab:         <ul> <li>Change the Isolate # to a letter</li> <li>Verify the result</li> </ul> </li> <li>Enter and verify vancomycin E-test result</li> <li>Keep vancomycin E-test result suppressed</li> <li>Report:         <ul> <li>No Vancomycin Resistant Enterococci (VRE) isolated"</li> </ul> </li> </ul>	
	IF	REPORT	
Blue and/or Pink colonies - Vitek ID: E.faecalis and/or E.faecium	Vancomycin E-test MIC = 4 μg/mL	<ul> <li>Re-incubate vancomycin E-test</li> <li>If after 48 hours MIC is still 4:</li> <li>Verify the organism ID</li> <li>Suppress GPI result in the isolates tab:         <ul> <li>Change the isolate # to a letter</li> <li>Verify the result</li> </ul> </li> <li>Enter and verify vancomycin E-test result</li> <li>Keep vancomycin E-test result suppressed</li> <li>Report:         <ul> <li>No Vancomycin Resistant Enterococci (VRE) isolated"</li> </ul> </li> </ul>	

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Policy Number: Date Approved: Page 4 of 7

Title: MIC31100-VRE Screen Type: Laboratory Services Program SOP

Issuing Authority: Director, Health Services Policy Number:

Next Review Date: Date Approved:

		<ul> <li>If after 48 hours MIC ≥8 μg/mL:</li> <li>Verify the organism ID</li> <li>List quantitation as "Isolated"</li> <li>Enter and verify vancomycin E-test result</li> <li>Keep vancomycin E-test result suppressed</li> <li>Report organism with isolate comment &amp;VRE1</li> <li>Add test ?REFD and finalize with "."</li> <li>In order entry, copy report to OCPHO (HPU1)</li> <li>In order entry, copy report to appropriate IPAC ward</li> <li>In order entry, add ESO code "VRE"</li> <li>Freeze and record in isolate log</li> <li>Forward isolate to DynaLIFE for vancomycin gene testing</li> </ul>
Blue and/or Pink colonies - Vitek ID: E.faecalis and/or E.faecium	Vancomycin E-test MIC = 8-16 µg/mL	<ul> <li>Repeat ID from vancomycin E-test plate</li> <li>Verify the organism ID</li> <li>List quantitation as "Isolated"</li> <li>Enter and verify vancomycin E-test result</li> <li>Keep vancomycin E-test result suppressed</li> <li>Report organism with isolate comment &amp;VRE2</li> <li>Add test ?REFD and finalize with "."</li> <li>In order entry, copy report to OCPHO (HPU1)</li> <li>In order entry, add ESO code "VRE"</li> <li>Freeze and record in isolate log</li> <li>Forward isolate to DynaLIFE for vancomycin gene testing</li> </ul>

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Policy Number: Date Approved: Page 5 of 7 Title: MIC31100-VRE Screen

Issuing Authority: Director, Health Services

Next Review Date:

Type: Laboratory Services Program SOP
Policy Number:
Date Approved:

	IF	REPORT
Blue and/or Pink colonies - Vitek ID: <i>E.faecalis</i> and/or <i>E.faecium</i>	Vancomycin E-test MIC = ≥32 μg/mL	<ul> <li>Verify the organism ID</li> <li>List quantitation as "Isolated"</li> <li>Enter and verify vancomycin E-test result</li> <li>Keep vancomycin E-test result suppressed</li> <li>The following isolate comment will be added: &amp;VRE</li> <li>In order entry, copy report to OCPHO (HPU1)</li> <li>In order entry, copy report to appropriate IPAC ward</li> <li>In order entry, add ESO code "VRE"</li> <li>Freeze and record in isolate log</li> </ul>

NOTE: STH IPAC ward is SIPAC. IRH IPAC ward is IIPAC

#### LIMITATIONS:

- 1. Organisms with atypical enzyme patterns may give anomalous results. The growth requirements of certain VRE can lead to their partial or total inhibition in culture.
- 2. Faecal specimens may cause some localized discolourization in the primary area of inoculation and should not be confused with a true chromogenic reaction wherein coloured colonies are visible. Interpret the colour of the isolate on well isolated colonies.
- 3. Strains of *E. faecalis* or *E. faecium* with intermediate resistance to vancomycin are infrequently encountered and may yield positive results.
- 4. Tightly clustered colonies of *Enterococcus gallinarum* and *Enterococcus casseliflavus* could appear as gray, slightly blue, or pink.
- 5. Leuconostoc, Pediococcus and Lactobacillus are intrinsically resistant to vancomycin, are inhibited or appear as colourless pinpoint colonies (blue or pink with tightly clustered colonies).
- 6. In some vancomycin-resistant *Enterococcus faecalis* strains such as ATCC 51299, which display low-level vancomycin resistance (MIC<32mg/L), the colonies may not develop the characteristic blue colour until 28h. If white colonies appear at 24h, a second reading must be performed at 28h.
- 7. Some Gram-negative bacteria may grow on VRESelect, but the size and colour of colonies enable differentiation from VREfm or VREfs.

# **CROSS-REFERENCES:**

- MIC36400-Referral of Category B Specimens to DynaLIFE
- MIC60040-Culture Media Quality Control

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Policy Number: Date Approved: Page 6 of 7

Title: MIC31100-VRE Screen Type: Laboratory Services Program SOP

Issuing Authority: Director, Health Services Policy Number:
Next Review Date: Date Approved:

#### **REFERENCES:**

1. Leber, A. (2016). *Clinical microbiology procedures handbook.* (4<sup>th</sup>ed.) Washington, D.C.: ASM Press

2. Jorgensen J.H., Pfaller M.A., Carroll K.C., Funke G., Landry M.L., Richter S.S., Warnock D.W. (2015). *Manual of Clinical Microbiology, 11<sup>th</sup> edition*. Washington, D.C: ASM Press

3. Bio-Rad. (2016/03). VRESelect package insert

APPROVAL:	
Date	

## **REVISION HISTORY:**

REVISION	DATE	Description of Change	REQUESTED BY
1.0	26 Apr 17	Initial Release	L. Steven
2.0	30 Nov 18	Updated to include new Vitek 2 instrument and two specimens per plate	L. Steven
3.0	30 Dec 21	Procedure reviewed and added to NTHSSA policy template	L. Steven
4.0	31 Aug 22	Updated to reflect new VRE agar VRESelect	L. Steven

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Policy Number: Date Approved: Page 7 of 7