



**Stanton Territorial Hospital**  
P.O. Box 10, 550 Byrne Road  
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**Document Name:**

After-Hours CSF Culture Setup Job Aid

**Distribution:**

Microbiology Specimen Processing Manual

**Document Number:**

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**Uncontrolled When Printed**

**What to do with CSF sample after-hours?**

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|----------|--|---|
| <b>1</b> | The specimen should be collected into a clean, sterile, leak-proof centrifuge tube: <ul style="list-style-type: none"> <li>• Tube 2 is for microbiology culture</li> <li>• Tube 4 is for viral testing if requested</li> </ul>   |   |
| <b>2</b> | CSF samples should be: <ul style="list-style-type: none"> <li>• Immediately transported to the laboratory</li> <li>• Should NOT be refrigerated</li> </ul>   |   |
| <b>3</b> | <ul style="list-style-type: none"> <li>• Insufficient volume for tests requested: contact the physician to prioritize requests</li> <li>• Leaking specimens should be processed. Add comment to order entry</li> <li>• Improperly collected, labeled, transported, or handled specimens should be processed. Waiver of responsibility form SCM40110 needs to be filled out by the responsible nurse</li> </ul>   |   |
| <b>4</b> | In the microbiology lab, open the sash on the BSC to turn the blower on. If the blower does not automatically start, press the "blower" button on the right-hand side panel. The BSC should run for at least 5 minutes prior to the inoculation of the CSF sample.   |   |
| <b>5</b> | <ul style="list-style-type: none"> <li>• Accession the sample in SoftMic in Order Entry with test code "CXCSF"</li> <li>• In the Micro tab, fill in the Collected, Received and Plated rows and save</li> <li>• Media labels print out in the order of media inoculation: <ul style="list-style-type: none"> <li>1:BA-C=<b>Blood</b></li> <li>2:CHO-C=<b>Chocolate</b></li> <li>3:MAC-0=<b>MacConkey</b></li> <li>4:SUP=<b>Red Top Supernatant Tube</b></li> </ul> </li> </ul> |   |
| <b>6</b> | > 1 mL received  | <ul style="list-style-type: none"> <li>• Centrifuge tube #2 at 3500 rpm for 10 minutes (program 2)</li> <li>• Transfer the supernatant to the SUP tube with a sterile pipette</li> <li>• Mix remaining supernatant with sediment in the collection tube and use to plate and prepare the smear</li> </ul> |
|          | < 1 mL received  | <ul style="list-style-type: none"> <li>• Do not centrifuge</li> <li>• Plate and prepare smear using the un-spun sample</li> </ul>   |

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| <b>7</b> | <p>Inoculate media and slides:</p> <ul style="list-style-type: none"><li>• Label two slides using a pencil with the accession number, patient's last name, and "CSF"</li><li>• In the biosafety cabinet, using a sterile pipette, aspirate fluid from the bottom of the collection tube</li><li>• Place 1 to 2 drops onto BA, CHOC, and MAC. Streak for isolated growth using a disposable inoculation needle. Streak out to cover the whole plate</li><li>• Prepare smears by placing 1 or 2 drops of CSF on microscope slides. Allow the drop(s) to form one large drop. Do not spread the fluid</li></ul> |
| <b>8</b> | <p>After plating:</p> <ul style="list-style-type: none"><li>• Place the slides on the drying rack in the BSC</li><li>• Place the BA and CHO plates in the top CO<sub>2</sub> incubator on the top shelf labelled "NEW WOUND"</li><li>• Place the MAC plate in the O<sub>2</sub> incubator on the top shelf labelled "NEW WOUND"</li><li>• Place the collection tube and supernatant tube in the bucket labelled "STERILE BODY FLUIDS"</li></ul>  |
| <b>9</b> | <ul style="list-style-type: none"><li>• Close the sash on the BSC and allow smears to dry in the BSC</li></ul> <p><b>NOTE: The reading of CSF smears is to be done by a Microbiology Technologist</b></p> <ul style="list-style-type: none"><li>• Leave requisition on the front bench of the microbiology lab</li></ul>   |

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