

<b>PROGRAM Standard Operating Procedure – Laboratory Services</b>	
Title: MIC33400 – Genital Culture-IUD	Policy Number:
Program Name: Laboratory Services	
Applicable Domain: Lab, DI and Pharmacy Services	
Additional Domain(s): NA	
Effective Date:	Next Review Date:
Issuing Authority: Director, Laboratory and Diagnostic Imaging Services	Date Approved:
Accreditation Canada Applicable Standard: NA	

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**GUIDING PRINCIPLE:**

Genital colonization by *Actinomyces* spp. has been associated with the use of (IUDs). *Actinomyces* spp. may be seen in smears from secretions around the IUD.

**PURPOSE/RATIONALE:**

This standard operating procedure describes the screening for *Actinomyces* spp. in intra-uterine devices (IUD) specimens.

**SCOPE/APPLICABILITY:**

This standard operating procedure applies to Medical Laboratory Technologists (MLTs) processing specimens for IUD culture.

**SAMPLE INFORMATION:**

<b>Type</b>	<ul style="list-style-type: none"> <li>IUD in a dry, sterile container</li> </ul>
<b>Stability</b>	If the sample is received in the laboratory and processed greater than 24 hours from collection: <ul style="list-style-type: none"> <li>Add specimen quality comment: "Delayed transport may adversely affect pathogen recovery"</li> </ul>
<b>Storage Requirements</b>	Refrigerated
<b>Criteria for rejection</b>	1. Unlabeled/mislabeled swabs 2. Specimen container label does not match patient identification on requisition

**REAGENTS and/or MEDIA:**

- Brucella agar (BRU) and Thioglycollate broth (THIO)

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### SUPPLIES:

- Sterile red top vacutainer tube
- Sterile pipette
- Disposable inoculation needles
- Microscope slides
- Wooden sticks

### EQUIPMENT

- Biosafety cabinet
- Vortex
- Centrifuge
- 35° ambient air and 35° CO<sub>2</sub> incubators
- VITEK 2 and supplies

### SPECIAL SAFETY PRECAUTIONS:

Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potentially infectious materials or cultures:

- Ensure that appropriate hand hygiene practices be used
- Lab gown must be worn when performing activities with potential pathogens
- Gloves must be worn when direct skin contact with infected materials is unavoidable
- Eye protection must be used when there is a known or potential risk of exposure of splashes
- All procedures that may produce aerosols or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC)
- The use of needles, syringes and other sharp objects should be strictly limited

All patient specimens are assumed to be potentially infectious. Routine Practices must be followed. Since viable micro-organisms are used, all cultures must be handled with appropriate precautions. All equipment in contact with cultures should be decontaminated by appropriate methods.

### QUALITY CONTROL:

- Refer to Test Manual for reagent quality control procedures


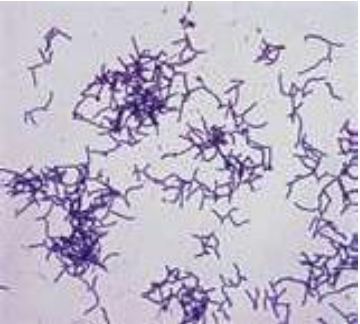
### PROCEDURE INSTRUCTIONS:

Step	Action
<b>Processing specimens for IUD culture</b>	
1	In the biosafety cabinet, add Thioglycollate broth to the specimen container containing the IUD and vortex for 30 seconds.
2	Using a sterile pipette, transfer the THIO broth to the red top vacutainer tube and centrifuge at 3500 rpm for 10 minutes.
3	After centrifugation is complete, remove the supernatant and transfer to a labeled red top tube. Place 1 to 2 drops of sediment onto BRU and add 2 to 5 drops into THIO broth. Streak the BRU for isolated growth using a disposable inoculation needle.

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<b>4</b>	<p>Incubate all media:</p> <ul style="list-style-type: none"> <li>Place specimen and supernatant tube in the O<sub>2</sub> incubator</li> <li>Label THIO with day 2 date, day 5 date and day 10 date and place in the THIO rack in the O<sub>2</sub> incubator</li> <li>Place BRU in anaerobic jar with anaerobic pouch and indicator as soon as possible after inoculation. Label jar with day 2 date and place in the O<sub>2</sub> incubator</li> </ul> <p><b>NOTE:</b> Anaerobes should not be exposed to air for 42 to 48 hours after inoculation</p>
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**INTERPRETATION OF RESULTS:**

Step	Action
<b>1</b>	<ul style="list-style-type: none"> <li>Observe BRU after 48 hours for growth of <i>Actinomyces</i> spp.</li> <li>If no growth suggestive of <i>Actinomyces</i> spp. is observed, re-incubate BRU for an additional 3 days</li> <li>If no growth suggestive of <i>Actinomyces</i> spp. is observed at 5 days, re-incubate for an additional 5 days</li> <li>After 10 days, examine plate for growth suggestive of <i>Actinomyces</i> spp.</li> <li>Colonies typical of <i>Actinomyces</i>: white, "molar tooth," pitting the agar</li> </ul> <div style="text-align: center;">  </div>
<b>2</b>	<ul style="list-style-type: none"> <li>Observe THIO after 48 hours for growth</li> <li>If no growth in THIO is observed, re-incubate THIO for an additional 3 days</li> <li>If no growth is present in THIO at 5 days, re-incubate for an additional 5 days</li> <li>If growth present, perform gram stain. If organisms resembling <i>Actinomyces</i> spp. are seen, culture broth to BRU agar and incubate anaerobically for 48 hours</li> <li>Gram stain of <i>Actinomyces</i> spp. is branching, gram-positive bacilli</li> </ul> <div style="text-align: center;">  </div>

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<b>3</b>	From growth of colonies suggestive of <i>Actinomyces</i> spp. on BRU, perform VITEK 2 ANC card to determine identification of organism.
<b>4</b>	From growth of subculture media from THIO, perform VITEK 2 ANC card to determine identification of organism.

**REPORTING INSTRUCTIONS:**

IF	REPORT
No <i>Actinomyces</i> spp. isolated	<ul style="list-style-type: none"> <li>Report: "<b>No Actinomyces isolated</b>"</li> </ul>
<i>Actinomyces</i> spp. isolated	<ul style="list-style-type: none"> <li>Add organism: "<b>Actinomyces spp.</b>"</li> <li>List quantitation as "<b>Isolated</b>"</li> </ul>

**CROSS-REFERENCES:**

NA

**REFERENCES:**

1. Leber, A. (2016). *Clinical microbiology procedures handbook*. (4<sup>th</sup>ed.) Washington, D.C.: ASM Press
2. Jorgensen J.H., Pfaller M.A., Carroll K.C., Funke G., Landry M.L., Richter S.S., Warnock D.W. (2015). *Manual of Clinical Microbiology, 11<sup>th</sup> edition*. Washington, D.C: ASM Press

**APPROVAL:**

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 Date

**REVISION HISTORY:**

REVISION	DATE	Description of Change	REQUESTED BY
1.0	20 Oct 17	Initial Release	L. Steven
2.0	22 Feb 21	Procedure reviewed and added to NTHSSA policy template	L. Steven
3.0	27 Feb 23	Procedure reviewed	L. Steven

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