

## TRAINING UPDATE

**Lab Location:** SGAH  
**Department:** Micro

**Date Distributed:** 1/1/2014  
**Due Date:** 1/31/2014  
**Implementation:** 1/15/2014

### DESCRIPTION OF PROCEDURE REVISION

**Name of procedure:**

**Gastrocult SGAH.M37.1**

**Description of change(s):**

<b>Section</b>	<b>Reason</b>
4.2	Added additional storage requirements
16	Removed date of Gastrocult pkg insert, added form

**This revised SOP will be implemented on January 15, 2014**

**Document your compliance with this training update by taking the quiz in the MTS system.**

**Approved draft for training (version 1)**

Technical SOP

<b>Title</b>	<b>Gastrocult</b>	
<b>Prepared by</b>	Hollie Genser	Date: 2/15/2012
<b>Owner</b>	Ron Master	Date: 11/26/2013

<b>Laboratory Approval</b>		<b>Local Effective Date:</b>
<b>Print Name and Title</b>	<b>Signature</b>	<b>Date</b>
<i>Refer to the electronic signature page for approval and approval dates.</i>		

<b>Review</b>		
<b>Print Name</b>	<b>Signature</b>	<b>Date</b>

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**1. TEST INFORMATION**

<b>Assay</b>	<b>Method/Instrument</b>	<b>Local Code</b>
Gastrocult Slide Test	Change in pH	GOBL

<b>Synonyms/Abbreviations</b>
None

<b>Department</b>
Microbiology

Form revised 3/31/00

## 2. ANALYTICAL PRINCIPLE

The Gastrocult slide includes both a specially buffered guaiac test for occult blood and a pH test based on the principle that certain dyes change color with changes in hydrogen ion concentration. This test is designed to be used with gastric samples. The occult blood test is not affected by normal therapeutic concentrations of cimetidine (Tagamet), iron or copper salts. Also, interferences from plant peroxidases are significantly reduced. Most guaiac-based products designed for use with fecal specimens are affected by these interferences, which are commonly encountered in gastric samples.

When a gastric specimen containing blood is applied to Gastrocult test paper, the hemoglobin from lysed blood cells in the sample comes in contact with the guaiac. Application of Gastrocult Developer (a buffered, stabilized hydrogen peroxide solution) creates a guaiac/oxidase-like reaction, which turns the test paper blue if blood is present. As with any occult blood test, results with the Gastrocult test cannot be considered conclusive evidence of the presence or absence of upper gastrointestinal bleeding or pathology. The Gastrocult test is designed for use as a preliminary screening aid and is not intended to replace other diagnostic procedures such as gastroscopic examination or X-ray studies (See Limitations of Procedure).

## 3. SPECIMEN REQUIREMENTS

### 3.1 Patient Preparation

Component	Special Notations
Fasting/Special Diets	N/A
Specimen Collection and/or Timing	It is recommended that samples be tested immediately after collection if possible. If this is not possible, the following procedure will yield satisfactory results:  Apply the sample in the pH Test Area and Gastrocult Test Area. Read the pH within 30 seconds after sample application. The Gastrocult Test Area may be developed immediately or up to 4 days, at room temperature 15-30°C, after sample application.
Special Collection Procedures	N/A
Other	N/A

### 3.2 Specimen Type & Handling

Criteria	
Type -Preferred	Gastric aspirate (nasogastric intubation or vomitus)

<b>Criteria</b>	
<b>-Other Acceptable</b>	None
<b>Collection Container</b>	Clean sealed container (plastic or glass)
<b>Volume - Optimum</b>	Sufficient amount of test material for application to the reaction area.
<b>- Minimum</b>	1 ml
<b>Transport Container and Temperature</b>	Clean sealed container (plastic or glass). Same as below.
<b>Stability &amp; Storage Requirements</b>	Samples may be stored, prior to application, at room temperature 15-30°C for 24 hours, or refrigerated at 2-8°C for 5 days. Frozen: Unacceptable
<b>Timing Considerations</b>	N/A
<b>Unacceptable Specimens &amp; Actions to Take</b>	Specimens that are unlabeled, improperly labeled, or those that do not meet the stated criteria are unacceptable. Request a recollection and cancel the test with the appropriate LIS English text code for “test not performed” message. Example: Wrong collection-UNAC. Document the request for recollection in the LIS.
<b>Compromising Physical Characteristics</b>	N/A
<b>Other Considerations</b>	N/A

#### 4. REAGENTS

Refer to the Material Safety Data Sheet (MSDS) supplied with the reagents for complete safety hazards. Refer to the section in this procedure covering “SAFETY” for additional information.

##### 4.1 Reagent Summary

<b>Reagents / Kits</b>	<b>Supplier &amp; Catalog Number</b>
Gastrocult Kit	Beckman Coulter Inc. Cat # 66040
Gastrocult Developer	Beckman Coulter Inc. Cat # 66115

##### 4.2 Reagent Preparation and Storage

**NOTES: Date and initial all reagents upon opening. Each container must be labeled with (1) substance name, (2) lot number, (3) date of preparation, (4) expiration date, (5) initials of tech, (6) any special storage instructions; check for visible signs of degradation.**

**Refer to the Material Safety Data Sheet (MSDS) for a complete description of hazards. If a specific hazard is present, it will be noted in this procedure when the hazard is first encountered in a procedural step.**

**Gastrocult Developer is an irritant. Avoid contact with skin. DO NOT USE IN EYES. Should contact occur, rinse promptly with water.**

<b>Assay Kit</b>	
<b>Reagent</b>	Gastrocult Developer, in a 10 ml vial and ordered separately. A developing solution containing a stabilized mixture of less than 2.9% hydrogen peroxide and 30% denatured ethyl alcohol in a citrate-buffered aqueous solution.
<b>Component</b>	Test slides for inoculation (40 tests per kit)
<b>Storage</b>	<ul style="list-style-type: none"> <li>Developer: Room temperature. Protected from heat and the bottle kept tightly capped when not in use. It is flammable and subject to evaporation.</li> <li>Slides: Protect slides from open air. Keep flap of slide sealed until ready to use. Store box containing slides at controlled room temperature 15-30°C in plastic storage pouch provided.</li> <li>Do not refrigerate or freeze</li> <li>Do not store slides or developer near volatile chemicals (e.g., iodine, chlorine, bromine, or ammonia)</li> <li>Protect from heat and light</li> </ul>
<b>Stability</b>	Stable until expiration on vial label or card label.
<b>Preparation</b>	The developer is ready to use. Slides are ready for use.

**5. CALIBRATORS/STANDARDS**

Not applicable

**6. QUALITY CONTROL**

**6.1 Controls Used**

<b>Controls</b>	<b>Supplier and Catalog Number</b>
Buffer Solution pH 2.00	Manufacturer: Lab Chem Inc Cat #:LC12220-1
Buffer Solution pH 7.00: Traceable to NIST	Manufacturer: La-Mar-Ka, Inc. Cat #: 0225

**6.1.1 PH:**

6.1.1.1 External Controls:

6.1.1.1.1 Apply one drop of pH 7 to pH test circle, interpret pH of sample within 30 seconds.

6.1.1.1.2 Apply one drop of pH 2 to pH test circle, interpret pH of sample within 30 seconds.

**6.1.2 Occult Blood:**

6.1.2.1 External Controls:

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6.1.2.1.1 Apply one drop of Camco Positive Control Solution to occult blood test area. Apply two drops of Gastrocult Developer directly over the sample in the occult blood area and interpret within 60 seconds. The positive internal control contains a hemoglobin-derived catalyst, which will turn blue within 10 seconds after applying developer.

6.1.2.1.2 Apply one drop of de-ionized water to occult blood test area. Apply two drops of Gastrocult Developer directly over the sample in the occult blood area and interpret within 60 seconds. The negative internal control contains no catalyst and should not turn blue after applying developer.

6.1.2.2 Internal Controls:

6.1.2.2.1 Add one drop of Gastrocult Developer between the positive and negative Performance Monitor areas. Interpret the Performance Monitor results. A blue color will appear in the positive Performance Monitor area within 10 seconds. The color will remain stable for at least 60 seconds. No blue should appear in the negative Performance Monitor area when developer is added. Note: If the sample is applied in such a way that it contacts the Performance Monitor areas, the negative Performance Monitor area may appear positive. This should be avoided.

**6.1.3** Results for the positive and negative internal and external controls for each patient test result must be documented on the log indicating the positive and negative internal controls were in range.

**6.2 Control Preparation and Storage**

**NOTE: Date and initial all controls upon opening. Each container should be labeled with (1) substance name, (2) lot number, (3) date of preparation, (4) expiration date, (5) initials of tech, and (6) any special storage instructions; check for visible signs of degradation.**

<b>Controls</b>	Buffer Solution pH 2.00 Buffer Solution pH 7.00: Traceable to NIST
<b>Control</b>	Camco Positive Control Solution 5 mg/dL bovine hemoglobin in an aqueous buffered solution with preservative
<b>Control</b>	De-ionized Water
<b>Storage</b>	Store at controlled room temperature 15-30°C
<b>Stability</b>	Stable until expiration on vial label or card label.
<b>Preparation</b>	All controls are ready to use.

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**6.3 Frequency**

Quality control, both internal and external, should be performed with each patient tested.

**6.4 Tolerance Limits**

If results are not as expected (see section 6.1), record results on Corrective Action Log along with the resolution; notify a Supervisor and repeat the test. Do not report out test results until the quality control issue is resolved.

**6.5 Review Patient Data**

Review patient results for any unusual patterns or trends. Contact your Supervisor if any are noticed.

**6.6 Documentation**

Refer to local policies and procedures for quality control documentation and to Quest Diagnostics records management program for record retention requirements.

**6.7 Quality Assurance Program**

- Each new lot number of reagent or new shipment of the same lot of reagent must be tested with external control materials. Performance of the new lot must be equivalent to the previous lot.
- Training must be successfully completed and documented prior to performing this test. This procedure must be incorporated into the departmental competency assessment program.
- The laboratory participates in CAP proficiency testing. All proficiency testing materials must be treated in the same manner as patient samples.
- Consult the Laboratory QC program for complete details.

**7. EQUIPMENT and SUPPLIES**

**7.1 Assay Platform**

Not applicable

**7.2 Equipment**

None

**7.3 Supplies**

None

**8. PROCEDURE**

**NOTE: For all procedures involving specimens, buttoned lab coats, gloves, and face protection are required minimum personal protective equipment. Report all accidents to your supervisor.**

**The package insert for a new lot of kits must be reviewed for any changes before the kit is used. A current Package Insert is included as a Related Document.**



**Important Note:** This test requires Gastrocult Developer, which is provided as part of this product. **Do not use Hemocult Developer or any other developing solution.**

Step	Action
1	Open slide.
2	Apply one drop of gastric sample to pH test circle and one drop to occult blood test area.
3	Determine pH of sample by visual comparison of test area to pH color comparator. This must be done within 30 seconds after sample application.
4	Apply two (2) drops of Gastrocult Developer directly over the sample in the occult blood test area. <b>Important Note:</b> Occasional gastric samples may be highly colored and appear as blue or green on the test area. Test results should only be regarded as positive if additional blue is formed after developer is added.
5	Read occult blood results within 60 seconds. The development of any trace of blue color in the occult blood test area is regarded as a positive result. Record results.
6	Add one (1) drop of Gastrocult Developer between the positive and negative Performance Monitor areas.
7	Interpret the Performance Monitor results

## 9. CALCULATIONS

Not applicable

## 10. REPORTING RESULTS AND REPEAT CRITERIA

### 10.1 Interpretation of Data

- A. A blue color will appear in the positive Performance Monitor area within 10 seconds. The color will remain stable for at least 60 seconds.
- B. No blue should appear in the negative Performance Monitor area when developer is added. **Note:** If the sample is applied in such a way that it contacts the Performance Monitor areas, the negative Performance Monitor area may appear positive. This should be avoided.
- C. Any blue originating from the Performance Monitor areas should be ignored when reading the specimen test results.
- D. Neither the intensity nor the shade of the blue from the positive Performance Monitor area should be used as a reference for the appearance of positive test results.

### 10.2 Reporting Results

- 1. Occult blood: Positive or negative
- 2. Ph: 1-10

**10.3 Units of Measure**

Not applicable

**10.4 Clinically Reportable Range (CRR)**

Not applicable

**10.5 Repeat Criteria and Resulting**

IF the result(s) are ...	THEN...
Inconclusive	Repeat the test(s). Notify a supervisor.

**11. EXPECTED VALUES**

**11.1 Reference Ranges**

1. Occult Blood: Negative

2. Ph: <4

**11.2 Critical Values**

None established

**11.3 Priority 3 Limit(s)**

None established

**12. CLINICAL SIGNIFICANCE**

The identification of occult blood can be useful in the early detection of gastric trauma or deteriorating gastric condition, while pH may be of use in evaluating antacid therapy.

**13. PROCEDURE NOTES**

- **FDA Status:** Approved/Cleared
- **Validated Test Modifications:** None

A. Because this test is visually read and requires color differentiation, it should not be interpreted by the visually impaired.

**B. Gastrocult Developer is an irritant. Avoid contact with skin. DO NOT USE IN EYES. Should contact occur, rinse promptly with water.**

**14. LIMITATIONS OF METHOD**

A. As with any occult blood test, the results of the Gastrocult test cannot be considered conclusive evidence of the presence or absence of upper gastrointestinal bleeding or pathology.

- B. Note: Many foods (e.g., incompletely cooked meat, raw fruits and vegetables, etc.) have peroxidase activity, which can produce a positive Gastrocult test result. Thus, a positive test result does not always indicate the presence of human blood.
- C. Gastrocult tests are designed as an aid to diagnosis, and are not intended to replace other diagnostic procedures such as gastroscopic examination or X-ray studies. There is disagreement in the literature regarding the exact therapeutic significance of varying levels of upper gastrointestinal bleeding.
- D. Gastrocult test results should be used only in conjunction with other information relevant to the clinical status of the patient. A positive test result may suggest the need for more careful monitoring of the patient.

**14.1 Analytical Measurement Range (AMR)**

N/A

**14.2 Precision**

N/A

**14.3 Interfering Substances**

Many foods (e.g., incompletely cooked meat, raw fruits and vegetables, etc.) have peroxidase activity, which can produce a positive Gastrocult test result.

**14.4 Clinical Sensitivity/Specificity/Predictive Values**

50 µg/ml of hemoglobin

**15. SAFETY**

The employee has direct responsibility to avoid injury and illness at work. Nearly all harmful exposures to infectious substances and chemicals, and other injuries, can be avoided with effective training and consistent safe work practices.

Become familiar with the Environmental Health and Safety (EHS) Manual to learn the requirements on working safely and protecting the environment from harm.

Although lab work typically focuses on the hazards of working with specimens and chemicals, we must also control other important hazards.

- Slips, trips, and falls cause many serious injuries. Please ensure that spills are cleaned quickly (to avoid slippery floors) and that you can see and avoid obstacles in your path.
- Ergonomic injuries result from performing tasks with too much repetition, force, or awkward position. Ergonomic injuries include strains and back injuries. Learn about ergonomic hazards and how to prevent this type of injury.
- Scratches, lacerations, and needlesticks can result in serious health consequences. Attempt to find ways to eliminate your risk when working with sharp materials.

Report all accidents and injuries immediately to your supervisor or the business unit Environmental Health and Safety Manager or Specialist.

**16. RELATED DOCUMENTS**

1. Laboratory Quality Control Program
2. Laboratory Safety Manual
3. Current Product Insert for Gastrocult
4. [Gastrocult Blood QC Log \(AG.F157\)](#)

**17. REFERENCES**

1. Rosenthal, P., Thompson, J. and Singh, M: Detection of Occult Blood in Gastric Juice. *J. Clin. Gastroenterol.* 6:119-121, 1984.
2. Data on file, Product Development Department, Beckman Coulter, Inc.
3. Norfleet, R.G., Rhodes, R.A. and Saviage, K: False-positive “Hemocult” reaction with cimetidine. *N. Engl. J. Med.* 302:467, 1980.
4. Gastrocult, *Test For Gastric Occult Blood And pH*. Beckman Coulter, Inc. September 2001.
5. GASTROCCULT procedure, MI162.004, Quest Diagnostics Nichols Institute, Chantilly

**18. REVISION HISTORY**

Version	Date	Section	Reason	Reviser	Approval
000	11/26/13		Update owner	L. Barrett	R. Master
000	11/26/13	4.2	Added additional storage requirements	R. Master	R. Master
000	11/26/13	16	Removed date of Gastrocult insert, added form	R. Master	R. Master
000	11/26/13	Footer	Version # leading zero's dropped due to new EDCS in use as of 10/7/13.	L. Barrett	R. Master

**19. ADDENDA**

None