#### TRAINING UPDATE

Lab Location: Department:

GEC, SGMC & WAH Core

Date Distributed:
Due Date:
Implementation:

7/15/2016 8/9/2016 **8/9/2016** 

#### **DESCRIPTION OF PROCEDURE REVISION**

# Name of procedure:

# Reagent Parallel Testing SGAH.QA37 v2

# **Description of change(s):**

Header: add other sites

Section 5.1, 5.2.1: specify 3 samples across AMR

Section 5.3.3: add reference to addendum

Section 9: add addendum

This revised SOP will be implemented on August 9, 2016

Document your compliance with this training update by taking the quiz in the MTS system.

# Non-Technical SOP

Title	Reagent Parallel Testing	
Prepared by	Jean Buss, Robert SanLuis	Date: 4/7/2011
Owner	Cynthia Bowman-Gholston	Date: 4/7/2011

Laboratory Approval			
Print Name and Title	Signature	Date	
Refer to the electronic signature page for approval and approval dates.			
Local Issue Date:	Local Effective Date:		

Review:			
Print Name	Signature	Date	

# **TABLE OF CONTENTS**

1.	PURPOSE	2
2.	SCOPE	2
	RESPONSIBILITY	
	DEFINITIONS	
	PROCEDURE	
	RELATED DOCUMENTS	
	REFERENCES	
	REVISION HISTORY	
9.	ADDENDA AND APPENDICES	7

#### 1. PURPOSE

This procedure defines the process for validation of new lots, new shipments, and new batches of reagent before or concurrent with use for patient testing.

#### 2. SCOPE

This SOP applies to all personnel who perform assay/sample testing.

#### 3. RESPONSIBILITY

The Supervisor is responsible for ensuring compliance with this SOP and that documentation of results is filed appropriately in each department.

The senior QA specialist is responsible for content and review of this procedure.

#### 4. **DEFINITIONS**

New Lot to Old Lot comparison: New reagent lots are checked against prior lots or known standards before or concurrent with being placed in use.

Same Lot comparison: Same lots received on different dates checked against prior lot or known standard before or concurrent with being placed in use.

TEa: Total Allowable Error; TEa is the amount of error that can be tolerated without invalidating the medical usefulness of the analytical result.

#### 5. PROCEDURE

#### **5.1** General Information

- A. The verification of reagent performance is required for all new shipment dates (even if the lot number is the same) and every time the lot number changes. Reagent parallel testing must be documented.
- B. A comparison of kits is performed to ensure that a new kit lot does not alter the performance of the tests.
  - 1. For qualitative tests, this is accomplished by running kit controls to confirm the manufacturer's ranges, and by running External Controls to check for bias between the kits.
  - 2. For quantitative tests, this is accomplished by running appropriate quality control to confirm the manufacturer's ranges, by running External Controls to check for bias between the kits and by testing at least 2 3 patient samples run simultaneously with both lots. The patient samples should span the analytical range (AMR) for the assay.

#### C. Documentation

- 1. Comparisons for quantitative tests are recorded on the New Reagent Lot or Shipment Comparison Study (Quant) Form. Results are recorded on the Comparison (TEA/4) tab. The form will perform all calculations.
- 2. The Lot to Lot Cross Check Log is used for qualitative test comparisons.
- 3. An alternative log may be used as specified in the test SOP (i.e. PFA-100, Coag lot conversion).

#### 5.2 Reagent Testing

5.2.1 Parallel testing when a new lot of reagent is received in the laboratory All new lot of reagents, kits and controls must be labeled with an orange "Do Not Use This Lot Number" sticker and be logged on the electronic receipt log.

#### A. For Qualitative Analysis Reagents / Kits / Controls

- 1. For kits that contain the controls
  - a. Run the controls from the old kit on the new reagent and the controls from the new kit on the old reagent
  - b. Label the kits with the green "This Lot is Ready to Use" sticker.
- 2. For kits with controls external to the kit
  - a. Run the external positive and negative controls on both the old and new kit.
  - b. Label the kits with the green "This Lot is Ready to Use" sticker.
- 3. For test with 'onboard' or 'built in controls'
  - a. Run the external positive and negative controls.

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- b. Positive and negative external and 'onboard' controls must be definite positive or negative.
- c. Label the kits with the "This Lot is Ready to Use" stickers.

#### 4. Controls

- a. Controls will be run with reagents that have already been tested.
- b. Positive and negative controls must be definite positive and negative. Any borderline results are not accepted.
- c. Label with the green "This Lot is Ready to Use" sticker.

## **B.** For Quantitative Analysis Reagents / Controls

#### 1. Reagent

- a. Reagent validation for those analytes that are new lot numbers will be performed in conjunction with Calibration.
- b. Calibration material (not otherwise in use as a calibrator), control material of known performance, and patient samples may be used for crosschecking reagents. A minimum of 3 patient samples must be tested. Ideally, the samples should span the AMR.
- c. See the Calibration procedure for each analyzer for the detailed procedure.
- d. Label reagent with the green "This Lot is Ready to Use" sticker.
- 2. Controls are run as per the Quality Control Program policy.

# 5.2.2 Reagents / Kit / Control of the same lot number that is presently being used with different ship and/or receipt dates

Reagents/Kit/Control of the same lot number with different ship and/or receipt date must be labeled with both a blue sticker and an orange date label, and must be logged on the electronic receipt log.

#### A. For Qualitative Analysis Reagents / Kits / Controls

- 1. For kits that contain the controls
  - a. Run the controls from the old kit on the new reagent and the controls from the new kit on the old reagent
  - b. Label the kits with the green "This Lot is Ready to Use" sticker.
- 2. For kits with controls external to the kit
  - a. Run the external positive and negative controls on both the old and new kit.
  - b. Label the kits with the green "This Lot is Ready to Use" sticker
- 3. For test with 'onboard' or 'built in controls'
  - a. Run the external positive and negative controls

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- b. Positive and negative external and 'onboard' controls must be definite positive or negative.
- c. Label the kits with the green "This Lot is Ready to Use" stickers.

#### 4. Controls

- a. Controls will be run with reagents that have already been tested
- b. Positive and negative controls must be definite positive and negative. Any borderline results are not accepted.
- c. Label controls with the green "This Lot is Ready to Use" stickers.

# **B.** For Quantitative Analysis Reagents / Controls

## 1. Reagent

- a. Run the current lot of controls using the new reagent.
- b. The control values should be within the 2SD limit for that analyte.
- c. Label the reagent with the green "This Lot is Ready to Use" sticker.

#### 2. Controls

- a. Run all levels of the control with the current lot of reagent.
- b. The control values should be within the 2SD limit for that analyte.
- c. Label the controls with the green "Ready to Use" stickers.

# 5.3 Analyze Data

- **5.3.1** Quantitative results of reagent cross checks must be evaluated according to statistical methods. The New Reagent Lot or Shipment Comparison Study (Quant) Form performs the calculations.
  - Individual difference between results with old and new reagents must not exceed half the allowable total error (TEa/2) for the assay.
  - Estimate of bias
    - o Mean value for the new reagent must not vary from the old reagent by more than one fourth of the TEa (TEa/4).
    - o The technical supervisor or above may approve a variation between old and new lot of one third of the TEa (TEa/3).
    - The technical director or above may approve a variation between old and new lot of one half of the TEa (TEa/2).
    - Only the medical director can approve the variation at straight TEa.
    - o **Note**: Data entered on the TEA/4 tab of the comparison form will pre-fill the other tabs (TEA/3, TEA/2 and TEA).
  - QC results for new reagent lot should fall within range for old lot of reagent.

**5.3.2** Positive controls must be Positive and Negative controls must be Negative. Any borderline results are not acceptable and must be repeated. Patient results should match prior results.

#### **5.3.3** Corrective action for failures

A failed lot-to-lot requires additional troubleshooting including but not limited to the following:

- How is the current reagent lot QC running against the peer?
- Was there a performance shift with the implementation of the current lot?
- Was there a recent calibration performed with a new lot of calibrator that caused a shift in performance?
- Is the same lot of calibrator being used or is it a new lot number?
- Has one of the other system labs already worked through this new lot implementation with or without issue?
- How does the current lot perform instrument to instrument how is the current QC performance instrument to instrument?

After considering the above questions, if all looks well then recalibrate and repeat the lot-to-lot comparison.

If the above criteria are not met, then perform an extended lot-to-lot comparison as well as an instrument to instrument comparison as needed utilizing our instrument to instrument criteria to decide our next steps.

- If the extended comparison passes and the instrument to instrument is acceptable, then QC ranges may be adjusted slightly as needed.
- If the extended studies fail, then proceed to extended troubleshooting and contact the manufacturer to work through the problem.

#### Note:

Refer to addendum for an example for troubleshooting lot to lot failures for the Vista analyzer. This example is intended to promote a train of thought; it is not all inclusive nor is it meant to resolve all potential problems.

#### 6. RELATED DOCUMENTS

Analyte/Test technical SOPs
Appropriate Analyzer Calibration procedures
Quality Control Program, QA procedure
Policy for Reagent Labeling and Handling, QA procedure
Coagulation Reagent Lot Conversion
Lot to Lot Cross Check Log (AG.F104)
New Reagent Lot or Shipment Comparison Study (Quant) Form (AG.F217)
SOP specific Cross Check forms

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Germantown Emergency Center

# 7. REFERENCES

Department-specific Checklists, College of American Pathologists, Laboratory Accreditation Program, Northfield, IL 60093

# 8. REVISION HISTORY

Version	Date	Reason for Revision	Revised By	Approved By
		Supersedes SOP L048.001		
000	5/2/16	Section 3: update job titles	L Barrett	C Bowman
		Section 4: add TEa		
		Section 5.1: add documentation description		
		Section 5.2: specify wording on green label		
		Section 5.2.1: add use of orange label		
		Section 5.2.2: add use of blue & orange date labels		
		Section 5.3: add calculations done by quant form,		
		add corrective action		
		Section 6: add forms and Coag SOP		
		Footer: version # leading zero's dropped due to new		
		EDCS in use as of 10/7/13		
1	6/28/16	Header: add other sites	R. SanLuis	C. Bowman
		Section 5.1, 5.2.1: specify 3 samples across AMR	L. Barrett	
		Section 5.3.3: add reference to addendum		
		Section 9: add addendum		

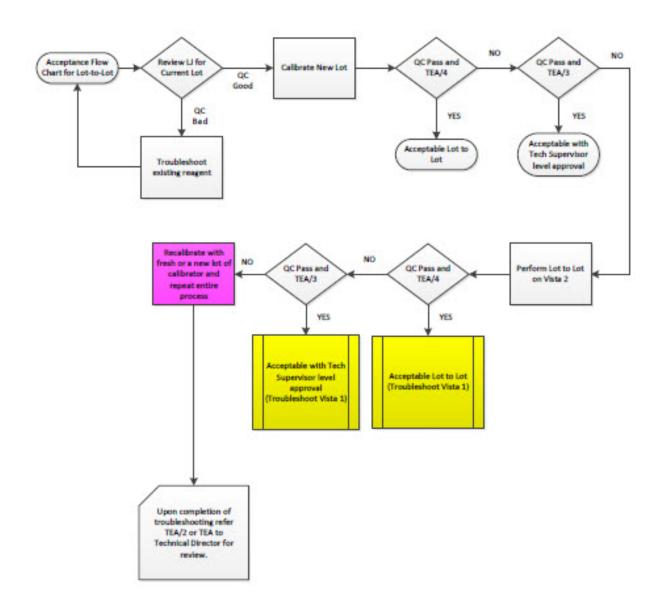
# 9. ADDENDA AND APPENDICES

Vista Lot to Lot Troubleshooting

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SOP version # 2 Page 7 of 8

# **ADDENDUM**

# **Vista Lot to Lot Troubleshooting**



Note: The samples being tested should span the range as much as possible. Consideration should be given to adding additional specimens to provide a clearer picture of the analyte performance.

SOP ID: SGAH.QA37 CONFIDENTIAL: Authorized for internal use only. Page 8 of 8