#### TRAINING UPDATE

Lab Location:

SGMC and WAH

Date Implemented: 7.19.2017

Department: Phlebotomy

Due Date: 8.4.2017

#### **DESCRIPTION OF PROCEDURE REVISION**

# Name of procedure:

Malaria Smear Collection

# **Description of change(s):**

Formatting was updated to align with other phlebotomy procedures. No changes in content were made.

# **Electronic Document Control System**



Document No.: SGAH.P23[3]

Title: Malaria Smear Collection

**Owner: LESLIE BARRETT** 

Status INWORKS

Effective Date: 13-Aug-2017

**Next Review Date:** 

Non-Technical SOP

Title	Malaria Smear Collection	
Prepared by	Leslie Barrett	Date: 9/10/2009
Owner	Ron Master	Date: 9/10/2009

Print Name and Title	Signature	Date
Refer to the electronic signature page for approval and approval dates.		
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Review:			
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## TABLE OF CONTENTS

1.	PURPOSE	2
	SCOPE	
	RESPONSIBILITY	
	DEFINITIONS	
	PROCEDURE	
	RELATED DOCUMENTS	
	REFERENCES	
8.	REVISION HISTORY	6
9.	ADDENDA AND APPENDICES	6

# 1. PURPOSE

This procedure describes specimen collection for malaria testing using the finger stick method.

#### 2. SCOPE

This procedure applies to all phlebotomists collecting specimens for malaria smear testing.

# 3. RESPONSIBILITY

All phlebotomists must understand and adhere to this procedure when collecting and preparing malaria smears.

The Microbiology Director and Field Operations Manager/Supervisor are responsible for the content and review of the procedure.

# 4. **DEFINITIONS**

None

# 5. PROCEDURE

Step	Action
1	Introduce yourself to the patient using AIDET technique (Acknowledge, Introduce, Duration, Explanation, Thank you).
2	Wash your hands and don latex-free gloves.
3	Identify the patient per procedure.

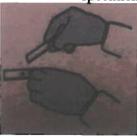
Step	Action				
4	Using a pencil, label 8 frosted slides with the following:				
	A. Patient's full name				
	B. Patient's medical record number				
	C. Date and time of collection				
	D. Collectors initials or identification code				
	E. Specimen accession number				
	Note: Do not use pen or marker to label slides. The staining process wil				
	remove pen or marker from the slide.				
	r				
5	Assemble the supplies necessary to perform a fingerstick collection per				
	procedure.				
	A. Latex-free gloves				
	B. Alcohol-prep pad (70% isopropyl alcohol)				
	C. 2x2 sterile gauze				
	D. Lancet				
	E. Warming device, if required				
	F. Band-Aid or tape				
	G. Biohazard sharps container				
	H. Frosted slides				
6	Determine the site used for blood collection.				
	A. For infants, puncture the heel.				
	B. For adults, use the middle or ring finger.				
	RED				
7	Warm the area to be punctured.				
8	Clean the puncture area with 70% isopropyl alcohol and allow the area to dry.				
	Be sure the finger is completely dry before pricking.				
	18				
9	Using an approved lancet, puncture the ball of the finger (or heel for infants).				
	Immediately discard the lancet in a biohazard sharps container.				
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	1000				
10	Wipe away the first drop of blood with clean gauze.				

11 Prepare four thin smears first.

A. Touch the drop of blood to a clean, labeled slide. Do not use blood from a lavender tube to prepare slides.



B. Obtain a clean, spreader slide, held at a 45° angle, toward the drop of blood on the specimen slide.



C. Wait until the blood spreads along the entire width of the spreader slide.



D. While holding the spreader slide at the same angle, push it forward rapidly and smoothly.



Strive for a thin smear that is rounded, feathered, and progressively thinner toward the center of the slide.

Step	Action
12	Prepare four thick smears next.
	A. Touch the drop of blood to a clean, labeled slide. Do not use blood from a lavender tube to prepare slides.
	B. Using the corner of a clean slide, spread the drop of blood in a circle the size of a dime (diameter 1-2 cm). Do not make the thick smear too thick, or it will fall off the slide (you should be able to read newsprint through it).
13	After collection, apply pressure to the puncture site with sterile gauze for 3-5 minutes until bleeding stops. The patient can assist if he/she is able.
14	Cover the site with tape or Band-Aid after the bleeding has stopped.
15	Allow slides to air dry in a flat position in a cardboard slide holder. Do not cover until drying is complete.
16	Collect an EDTA specimen per venipuncture procedure.
17	Recheck the labeling by comparing the name and MRN on each slide and tube to the name and MRN on the patient wristband.
18	Place the specimen tube in a biohazard bag and seal.  A. Do not put more than one patient's specimens in a bag.  B. Never transport specimens that are not in a biohazard bag.
19	Complete the malaria history form by asking the patient what countries he/she has visited.
20	Clean the work area by discarding all used materials in the appropriate waste container. Do not leave any trash behind.

Step	Action
21	Thank the patient and wish him/her a good day.
22	Wash hands.
23	Receive the specimens in the laboratory computer and deliver the smears, specimen, and history form to microbiology.

# 6. RELATED DOCUMENTS

SOP: Patient Identification and Specimen Labeling

SOP: Venipuncture SOP: Fingerstick

SOP: Heelstick Specimen Collection

Form: Malaria History Form, AG.F289 (see Reference tab)

#### 7. REFERENCES

A. Giemsa Stain for Malaria, Babesia, and other Blood Parasites, Parasitology Subteam of the Microbiology Best Practice Team, ODMI709a, ver 3.0.

B. Centers for Disease Control and Prevention, Laboratory diagnosis of malaria: Preparation of blood smears.

# 8. REVISION HISTORY

Version	Date	Reason for Revision	Revised By	Approved By
		Supersedes SOP P008.001		
000	7/1/2014	Section 3: update titles, remove annual review Section 6: add revised form Section 9: remove outdated form Footer: version # leading zero's dropped due to new EDCS in use as of 10/7/13.	L. Barrett	R. Master, S.Khandagale
1	7/27/16	Header: add other site Section 5: Updated format and wording for clarity. Added pictures to the procedure and requirement to collect a venous EDTA sample in addition to slides. Section 9: Deleted the addendum.	S.Codina	R. Master
2	7/14/17	Section 5: Updated format to align with phlebotomy SOPs; moved supplies into procedure steps Section 6: Updated SOP titles	SCodina	RMaster

## 9. ADDENDA AND APPENDICES

None