Microbiology	Document No. MICR 6140 R
Planter	Page 1 of 7
Specimen Processing	Origination: 03/2007

POLICY STATEMENT	The initial processing of clinical specimen for bacteriology is a multifaceted endeavor involving a number of decision- making steps, including the need for processing the specimen for gram stains, aerobic, anaerobic, fungal, and mycobacterial cultures. These issues will determine whether the specimen requires any pretreatment before inoculation. First one must consider the specimen type and its anatomic origin. The second step is the selection of primary isolation media to be used for each specimen type. The final step is the selection of incubation temperature and atmosphere.
PURPOSE	This procedure provides technical instruction for the performance of the specimen processing.
SCOPE	This procedure applies to testing personnel authorized to perform testing. This group includes, but is not limited to Laboratory Technologists as well as leads and supervisory personnel.
RESPONSIBILITY	All the above personnel are responsible for following the specimen processing procedure without exception. In addition, testing personnel are also responsible for evaluating the results and taking proper remedial action.
RELATED DOCUMENTS	MICR 6140 Ja Transfusion Reaction Blood Cultures MICR 6140 Jb Set Up Chart MICR 6140 Jc Media Chart MICR 6140 Jd Micro Specimen Processing MICR 6370 J BAL Quantitative Cultures MICR 6200 R Gram Stain

SPECIMEN HANDLING

Proper Specimen collection is critical to isolating the causative agent of infection. See the Laboratory Test Directory and Laboratory Service Manual located on the SAINT for specimen collection and transport information.

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Specimen Processing	Origination: 03/2007

Specimen Aliquoting Requirements:

- Always utilize a fresh pipette any time material is aspirated.
- Never add a sample into an unlabeled container.
- Never mix sample types in one container.
- Never return an aliquot to the original container.
- The original container and the aliquot tube must be legibly initialed by the person performing the aliquoting.
- If a label is placed over an existing label, the new label must be initialed by the person performing the task.

QUALITY CONTROL

Quality Control is performed as outlined in CLSI Approved Standard M22: Quality Control for Commercially Prepared Microbiology Culture Media

PROCEDURE

<u>Media</u>

- Enrichment Media: Nutrients have been added to enhance the growth of bacteria.
- Differential Media: The media contains carbohydrates, indicators and chemicals that will differentiate different types of bacteria.
- Selective Media: This media contains certain carbohydrates, indicators and chemicals that are inhibitory to certain groups of bacteria.
- For appropriate selection of media see MICR 6140 Jb Set Up Chart and MICR 6140 Jc Media Chart

Inoculation

The streak plate method is a rapid and simple technique of mechanically diluting a relatively large concentration of microorganisms to a small, scattered population of cells. The goal is to obtain isolated colonies on a large part of the agar surface, so that desired species can then be brought into pure culture. Proper streaking of plates is an

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Specimen Processing	Origination: 03/2007		

indispensable tool in microbiology. In most cases a closed inoculating loop is used for streaking plates.

- All specimens are to be plated in the level II biological safety cabinet.
- Labeling:
 - Test request specimens through computer and obtain accession number for the cultures. See MICR 6140 Jd Micro Specimen Processing.
 - Place the small aliquot computer label on plates and tubes being careful not to cover the name of the media or expiration date.
 - Place string number on plates.
- <u>Urines cultures</u>, do not streak in quadrants. Instead use the appropriate calibrated loop (Inoculate with white - 0.001 ml or blue - 0.01 ml). Streak plate according to this pattern



Other Specimen types:

- If swabs are received, use one swab for the culture and the other swab for the gram stain. Discard the swab that was used for the gram stain. If only one swab is received, inoculate media first, and then make the smear.
- If a sterile body fluid is received and the volume is >1cc and clot is not present, centrifuge for 15 min at 2500 RPM's for the culture and the cytocentrifuge is used to make the direct slide for staining. If clot is present, break up the clot and set directly.

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Specimen Processing	Origination: 03/2007

- If a bronch wash is received and the volume is less than 10 ml set up the specimen directly. However, if the volume is greater than 10 ml, remove 2 ml, centrifuge for 15 min at 2500 RPM's and work with the sediment for the culture.
- If a BAL is received, set up the quantitative culture. See MICR 6370 J BAL Quantitative Cultures
- If a tissue is received, mince tissue into 1 mm. cubes and emulsify with tissue grinder in sterile saline.
- Inoculate the upper 1/3 of the solid media starting with the most nutritious media and least selective media (e.g. chocolate agar)
- o Method for streaking plates
 - Position the plate so that the spot of inoculum is nearest the hand not holding the loop (the opposite hand).
 - Use a sterile disposable loop, streak with gentle pressure. Use the method shown below to avoid tearing the agar.



http://web.indstate.edu/thcme/micro/basic.html

 Move the loop back and forth across the spot and then gradually continue toward the center of the plate as you sweep back and forth. Use a very gentle and even pressure.



http://web.indstate.edu/thcme/micro/basic.html

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Specimen Processing	Origination: 03/2007

- When creating the first quadrant, do not worry about keeping each pass across the plate separate from previous ones.
- When about 30% of the plate has been covered by the first streaking quadrant, rotate the plate and repeat the above procedure with the <u>same</u> loop for the second quadrant, but this time pick up some inoculum by crossing into the first quadrant 2-3 times and then not passing into it again.
- Repeat the same procedure for the third quadrant. After streaking the plate, discard the loop. Use a new loop for each plate to avoid carry over.



http://www.mansfield.ohio-state.edu/~sabedon/biol4035.htm

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- <u>Catheter tip cultures</u>:
 - Using sterile forceps transfer the catheter segment onto the surface of a BAP. If
 Tip is longer than 2 inches, using a sterile scalpel, cut off the tip to culture.
 - With light pressure from the forceps, roll the catheter back and forth across the plate at least four (4) times.
 - Return the catheter tip to the specimen container. Save with daily Specimens
- When multiple types of cultures are requested, bacterial cultures should be set up first.
- If fungus cultures are requested, inoculate Sabouraud Dextrose agar and Mycosel agar
- For Mycobacterial cultures put 5-10 ml of the specimen in a 50ml conical tube to be processed. See the procedure for Mycobacterial cultures.
- Incubate plates with the lid down in the designated 35°C CO2 incubator for 18-24 hours.
- Incubate fungal tubes in the 30°C incubator.
- Place anaerobic cultures in the appropriate environment by using the appropriate Gas Pak and place in the 35°C non CO2 incubator.
- Archive and save all specimens in the refrigerator for 1 week. Save all sterile body fluids and other irretrievable specimens for 4 weeks.

REPORTING

The interpretation of primary cultures requires considerable skill. From initial observations, the microbiologist must assess the colony growth and determine, depending on the source of the culture, whether any pathogens are present. Proper steps should be taken to quantitate, identify the microorganisms, and perform appropriate antimicrobial susceptibility testing. Report colony numbers according to following protocol:

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Specimen Processing	Origination: 03/2007

- Rare Growth = one to five colonies total
- Light Growth = colonies in first quadrant only ignoring a few colonies in second
- Moderate Growth = colonies up to the second quadrant ignoring a few colonies in third
- Heavy Growth = colonies up to the third quadrant
- Urine cultures = <10, 10-50, 50-100 or >100 CFU/ml
- Catheter tip cultures = <15 CFU or >15 CFU

REFERENCES

Isenberg, H. *Clinical Microbiology Procedure Handbook.* 2nd Edition, Volume 1, Section 3.3.1; Processing of Specimens, 2004

Bailey and Scott's Diagnostic Microbiology 11th edition, Mosby, Inc

BBL Quality Control and Product Information Manual for Plated and Tubed Media Becton Dickinson and Company, June 2009

CLSI Approved Standard M22-A3: Quality Control for Commercially Prepared Microbiology Culture Media

Transfusion Reaction – Blood Cultures

Blood Bank will bring 2 BacT/Alert bottles (aerobic and anaerobic), the unit to be cultured and the Meditech labels to Microbiology. If the unit itself is no longer available, the Blood Bank will submit 1 or 2 segments from the unit for culturing.

Receive the Blood culture in Meditech.

If direct gram stain is requested by a Pathologist, order GRAM

Source:	BBU = BLOOD BANK UNIT
Specimen description:	CRYO = CRYOPRECIPITATE GRAN = GRANULOCYTES P = PLASMA
	PT = PLATELETS RBC = RED BLOOD CELLS

Inoculate the blood bottles as follows:

- Label bottles.
- Inspect unit to determine amount of blood that can be removed via the 'pig-tail' of the unit. A pig-tail is the sealed, segmented tubing that is external to the blood unit
- Remove plastic covers from BacT/Alert bottles and clean the rubber stopper with an alcohol pad
- Place unit in such a manner that any product still inside is against the 'pig-tail' opening.
- Clamp unit approximately 1" below the tip of the 'pig-tail'.
- Wrap tip of 'pig-tail' with alcohol swab.
- Open syringe, insure needle is on tight, and express any air in syringe out.
- Carefully insert needle into tip of 'pig-tail'.
- Release clamp.
- Aspirate blood into syringe (should draw enough blood to place a minimum of 1.5cc in each bottle, if possible)
- Clamp off 'pig-tail' approximately 1" below the needle insertion site
- Remove syringe and inoculate BacT/Alert bottles
- Bend 'pig-tail' between tip and clamp.
- Slid metal crimp over fold
- Crimp, sealing unit
- Remove clamp

SOURCE	GRAM STAIN	CHOC	BAP	MAC	CNA	V/ML	THIO	CDC/KV	HE/SMAC/TCBS
Catherter Tip			Roll plate						
·									
CSF	Х	Х	Х				shunts only		
Sterile Body Fluids	Х	Х	Х	Х					
Joint Fluids	Х	Х	Х	Х			Peds bottle		
CC / CATH (0.001)			Х	Х					
CATH ≤12 yrs (0.01)			Х	Х					
Urine invasive (0.01)		Х	Х	Х					
Gential (cx,vaginal,urethral)		Х	Х	Х		Х			
Placenta		Х	Х	Х		Х			
GC ONLY		Х				ML			
Group B only		LIM Broth							
Sputum	Х	Х	Х	Х					
Endotracheal Aspirate									
Throat		Х	Х						
Nose/Nasopharyngeal		Х	Х						
Gastric	Х	Х	Х	Х					
BAL (0.01 & 0.001)	Х	Х	Х	Х					
Bronch Wash	Х	Х	Х	Х					
Stool			Х	Х					Х
Yersina		CIN							
Wound	Х	Х	Х	Х	Х				
Tissue	Х	X	X	X	X				
Abscess	Х	X	X	Х	Х				
Drainage	Х	X	X	<u> </u>	X				
Anaerobic		X	X					Х	
Actinomyces		<u> </u>	Х					X	
Grafts							Х		
Acinteobacter			X	X					
MRSA Culture		MRSA							
VRE		VRE							

	Name	Туре	Nutrients	Inhibotors	Indicators	Organisms Selected	Organisms Inhibited
BAP	Trypticase Soy with 5% Sheep Blood	Enrichment	Sheep blood				
CAMPY	Camplyobacter Agar	Selective	Sheep blood	Amphotericin B Cephalothin Trimethoprim Vancomycin Polymixin B		Campylobacter jejuni	Normal Fecal Flora
CDC	CDC with 5% Sheep Blood	Enrichment	Sheep blood			Anaerobes	Aerobes
СНОС	Chocolate II Agar	Enrichment	Hemoglobin IsoVitale X				
CIN	Cefsulodin-Irgasan-Novobicin	Selective Differential	Mannitol	Crystal Violet Na desoxycholate Cefsulodin Irgasan Novobicin	Neutral Red	Yersinia enterocolitica	gram neg & pos
CNA	Columbia CNA with 5% Sheep Blood	Selective	Sheep blood	Colistin Nalidixic Acid		Gram Positive	Gram Negative
HE	Hektoen Enteric	Selective Differential	Lactose Sucrose Salicin	Bile Salts	Acid Fuchsin Bromthymol Blue Ferric Ammonium Citrate Na Thiosulfate	Gram Negative	Gram Positive
ΚV	CDC Laked Sheep Blood w/ Kanamycin & Vancomycin	Selective	Laked Blood	Kanamycin Vancomycin		Gram Negative Anaerobes	Gram Pos Aerobes and Anaerobes
LIM	LIM Broth	Selective	Todd Hewitt Broth	Colistin Nalidixic Acid		Group B Strep	Gram Negative
MAC	MacConkey II	Selective Differential	Lactose	Crystal Violet Bile Salts	Neutral Red	Gram Negative	Gram Positive
ML	Martin Lewis	Selective	Hemoglobin IsoVitale X	Vancomycin Colistin Anisomycin Trimethoprim		N. gonorrhoeae	Gram Positive Gram Negative Yeast
SMAC	MacConkey II with Sorbitol	Selective Differential	Sorbitol	Crystal Violet Bile Salts	Neutral Red	Gram Negative	Gram Positive
TCBS	Thiosulfate Citrate Bile Salts Sucrose	Selective	Sucrose	Bile Salts	Bromthymol Blue Thyomol Blue Na Thiosulfate Ferric Citrate	Vibro	Normal Fecal Flora
THIO	Enriched Thioglycollate	Enrichment	Vitamin K1 Hemin	Na Thioglycollate		Anaerobes	
V agar	V Agar	Enrichment	Human Blood			G. vaginalis	

Specimen Processing and Edit

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2TY	SECOND ABO/RH TYPE	110.1001	В	Y		
3CEN	FISH, TRISOMY 3	910.2650	L	Y		
3TY	THIRD ABO/RH TYPE	110.1030	В	Y		
4TY	FOURTH ABO/RH TYPE	110.1040	В	Y		
5-OH-TRYPTAMINE	SEROTONIN, WHOLE BLOOD	1010.0535	L	Y		Mnem
5FC	ANTIFUNGAL, 5-FLUOROCYTOSINE	5000.2145	L	Y		Nam
5FC95316	5-FLUCYTOSINE, ANTIMICROBIAL	1010.0642	L	N		Numb
5FC95317	PEAK VALUE	1010.0644	L	N		Num
5FC95318	TROUGH VALUE	1010.0646	L	N		
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Enter search criteria into the textbox > Select the comment, once it is located.

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Select Enter/Edit Req

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🔄 LIS Specimen Desktop - SAH/ML	(Test) - Jane M Weiger				_	
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🔄 LIS Specimen Desktop - SAH/ML (SBA/SE	A.TEST5.65/SBA.TEST5.65 - Test) - Jane M Weiger	_	
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Enter a new account Facility = LAB Registration = REFS

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Enter required * information

Client Codes:

PHR = Pharmacy INF = Infection Control

LAB = Laboratory

leditech Health Care Information Syste	m			
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Enter required * information

Location Codes: PHR = Pharmacy **INF = Infection Control**

LABML = Main Laboratory

🛂 Meditech Health Care Inforn	nation System			
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SAVE - F12

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SAVE - F12
An Account Number starting with a "C" will be generated.



Continue order same as a patient

🖶 LIS Specimen Desktop - SAH/ML (SBA/	BA.TEST5.65/SBA.TEST5.65 - Test) - Jane M	Weiger		_ 🗆 ×
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Receive Specimens

From the Specimen Desktop Select Receive

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Select Specimen



Enter Specimen Numbers Select F12 when finished

🔄 LIS Specimen Desktop - SAH/ML (Test) - Jane M Weiger								_			
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Select F12 or Yes to finish



Print Labels

From the Specimen Desktop Select Labels

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Select Individual



Enter Specimen Numbers Select F12 when finished

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Enter Label Format e.g. Aliquot Select F12 when finished

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Select Printer > Change if necessary Select F12 when finished



Edit Specimens

From the Specimen Desktop Select Edit

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Select Specimen Data to Enter a Comment

🚰 LIS Specimen Desktop - SAH/ML (Test) - Jane M Weiger	
	%
Edit Specimen Data Sample Types Markers Codes Testing Receive Time LaB Timed Specime MIC Source/Spec Description MIC Source/Spec Description MIC Setup Times BBK Wristband BBK Specimen Expiration Cancel Product Order Uncancel Product Order	Single Image: Concelling and the second and the se
	? @ \$ A E

Enter Specimen Numbers Select F12 when finished



Enter Comment in Comments field Select F12 when finished

<mark>ች</mark> L	IS Specimen Desktop - SAH/ML (Test) - Jane M	Weiger			
0	Apple,Inpat 78/M 01/26/1932	SA0000002220 (SA00000194) Spec: 10:M0000009R	Status: RECD 4S 419-2 Dx/Rfv:	Dis:	2
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Select F12 or Close to File



Select Collection Data to Change the Collection Date

🚰 LIS Specimen Desktop - SAH/ML (Test) - Jane M Weiger		
	2	
Edit Specimen Data Collection Date Sample Types Markers Codes Testing Receive Time LAB Timed Specime MIC Source/Spec Description MIC Source/Spec Description MIC Setup Times BBK Wristband BBK Specimen Expiration Cancel Product Order Uncancel Product Order	Single Worklist Edit Enter/Edit Req Cancel Worksheets Enter Results Entry Screen Workcards Spreadsheet Inquiries Labels Collection Receive Site Batches Tasks Storage Change Site Tracking EMR	
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Enter New Collection Date > Specimen number Select F12 when finished

LIS Specimen Desktop - SAH/ML (Test) - Jane M Weiger			_	
			3	
New Collection Date 02/25/10 Co Specimen Tim Sts 10: M0000006R 1100 RECD APPL	Patient Name Account E,INPAT SA000002220	Coll Date 02/23/10	Single Worklist Edit Enter/Edit Req Cancel Worksheets Enter Results Entry Screen Workcards Spreadsheet	
Label Device Coll or Spec Label		Cancel Save	Inquiries Labels Collection Receive Site Batches Tasks Storage Change Site Tracking EMR	

Select MIC Source/Spec Description to Change the Source or Specimen Description

🚰 LIS Specimen Desktop - SAH/ML (Test) - Jane M Weiger		
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Edit Specimen Data Collection Date Sample Types Markers Codes Testing Receive Time MIC Source/Spec Description MIC Setup Times Bex. Wristband Bex. Specimen Expiration Cancel Product Order Uncancel Product Order	Single Worklist Edit Enter/Edit Req Cancel Worksheets Enter Results Entry Screen Workcards Spreadsheet Inquiries Labels Collection Receive Site Batches Tasks Storage Change Site Tracking EMR	

Enter the Specimen Number > Enter the New Source and/or New Spec Desc > Select F12 when finished

<mark>ች</mark> L	IS Specimen Desktop - SAH/ML (Test) - Jane M ¥	/eiger			
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Select F12 or Close to File



Specimen Inquiries

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# Select the type of Inquiry you wish to view

Inquiries         Print Specimens         Print Specimens (Internal)         Inquiry by Test View Group         Quick View         Bak Transfusions by MRI         Bak Transfusions by MRI         Bak View Available Products by Patient         Historical Results         Tracking	👺 LIS Specimen Desktop - SAH/ML (Test) - Jane M Weiger	_	
Inquiries         Print Specimens         Print Specimens         Print Specimens         Inquiry by Test View Group         Quick View         Review Results         BKK Product Inquiry         BKK Product Inquiry         BKK View Available Products by Patient         Historical Results         Tasks         Straage Til         Tasks         Tasks         BK Transfusions by MRI         BK Transfusions down         Review Results         Tasks         Tasks         Storage Til         Thange Sitce Storage         Tasks         Tasks		<u>ک</u>	
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# Enter the Patient's Name, Account Number or Specimen Number in the Specimen field. Select F12.

<mark>ሓ</mark> ∟	IS Specimen Desktop - :	SAH/ML (Test) - Jane M Weiger						_	
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## Select F12 to view the result.

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	Cancel OK		

## The report will display. Print Specimens

RUN DATE: 02/25/10 RUN TIME: 1437	Labor Saint Agnes H Department of 900 ( Baltin 41(	ratory *TEST* ospital Resul i Laboratory H Caton Avenue More, MD 21228 ) 368-3060	t Report Medicine }	PAGE 1
PATIENT: APPLE, INPAT REG DR: GALLAGER M.D., LAUR	ACCT #: AGE/SX: ENCE DOB: STATUS:	SA0000002220 78/M ADM IN	LOC: 4S ROOH: 419 HED: 2 TLOC:	<b>U #</b> :SA00000194 RBG:01∕26∕10 DIS:
SPEC #: 10:M0000009R SOURCE: LEG SPDESC: LEFT ORDERED: WOUND COMMENTS: Specimen receive	<b>COLL:</b> 02/25/10-1000 <b>RECD:</b> 02/25/10-1118 <b>ENTR:</b> 02/25/10-1135 d from the O.R.	STATUS: SUBM DR: OTHER DR:	RECD GALLAGER M.D.	REQ #: 00000519 ,LAURENCE R
Procedure	Result			
WOUND	PENDING			

## The report will display. Print Specimens (Internal)

DATE: 02/2 USER: JWEI	25/10 @ 1440 Laboratory *TEST* IGER MIC SPECIMEN INTERNAL INQUIRY	PAGE 1							
PATIENT: APPLE, INPATACCT: SA0000002220LOC: 4SU: SA00000194AGE/SX: 78/MROOM: 419REG: 01/26/10REG DR: GALLAGER M.D., LAURENCE RDOB: STATUS: ADM INBED: 2DIS:									
SPEC : 10	SPEC:         10:M0000009R         ORD FOR:         02/25/10-1000         STATUS:         RECD         REQ:         00000519           COLL:         02/25/10-1000         SUBM DR:         GALLAGER M.D., LAURENCE R           RECV:         02/25/10-1118         PT AGE AT COLL:         78								
SOURCE: ENTERED: COLL BY: LAST RPID: LAST ACT:	LEG SP DESC: LEFT 02/25/10-1135 ENT BY: JWEIGER OTHR DR: RCV BY: JWEIGER : 02/25/10-1437 WKID FN: 02/25/10-1406 BAR CD: 000516								
ORDERED: SETUP: COMMENTS:	ORDERED: WOUND SETUP: (NO PROCEDURES SETUP) COMMENTS: Specimen received from the O.R. @Source changed from ARM to LEG by JWEIGER. @Spec Desc changed from RIGHT to LEFT by JWEIGER.								
COL CATEG: ORD SITE: RECV SITE: PERFORM SI	: ML TRANSIT SITE: : ML ITE: ML 02/25/10-1118 JMW AT SITE: ML 02/25/10-	-1118 JMW							
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# **Cancel Specimens**

## From the Specimen Desktop Select Cancel to cancel a specimen

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Specimen	Patient	Time	BV	Rec Date	Rec Time	Rec Bv	EQ CD	Sts	Collection
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	oduro	D.	ate		Time		BV		(EMP

# **Enter Specimen Number**

#### Enter Cancellation Comment Select F12 when finished

<mark>₩</mark> ,L	S Specimen Desktop - SAH/ML (Test) - Jan	e M Weiger			_ 🗆 🗙
0	<b>Apple,Inpat</b> 78/M 01/26/1932	SA0000002220 (SA00000194) Spec: 10:M0000011R	Status: RECD 4S 419-2 Dx/Rfv:	Dis:	k
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	Cancel Spec Wkld				Tasks 🔡 Storage 🎧
					Change Site 📑
					Tracking 🍄
				Cancel Save	

# Select F12 or Yes to Cancel

	IS Specimen Desktop - SAH/ML (Test) - Jane M \	Veiger				Ľ
0	<b>Apple,Inpat</b> 78/M 01/26/1932	SA0000002220 (SA00000194) Spec: 10:M0000011R	Status: RECD 4S 419-2 Dx/Rfv:	Dis:	<i>i</i> ×	
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					Site Batches	
	Cancer Spec wkiu				Change Site	
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#### IMPORTANT:

## This following process is for canceling an ENTIRE test requisition.

## Select Requisition Desktop



## **Select Cancel Req**

LIS Requisition Desktop - SA	H/ML (Test) - Jane M Weiger				
	Orders Doctors	Specimens) (Addi	tional		
Patient Req Number		•			
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					Edit LIS Pt Data 😭 Edit MRI Markers 🐼
	BBK History Product View Group	Test/Proc Notes		Cancel Save ★ Ø	?@&@=

# Enter the Patient's Name or Account Number in the Patient field. Patient Demographics will appear on the top of the screen.

Tion Desktop - SLO/ML (Test) - Daniel Weiger					
<b>Apple,Orange</b> 21/M 06/09/1988	LOOOOOOOOO59 (SAOOOO0109)	Dr: PINTO M.D.,DOUGLAS	DBR REG CLI		×
Patient APPLE, ORAN	GE				
Req Number			<b>•</b>		Single 🛛 🖻
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					Edit MRI Markers 📾
Cancel Req Wkld					
			Cano	el Save	? 🗣 🛇 👚 🗉

### Select F9 in the Req Number field. A new window will display.

W LIS Requisition Desktop - SLO/ML (Test) - Daniel Weiger						<u>_   ×</u>
<b>Apple,Orange</b> 21/M 06/09/1988	LO0000000059 (SA00000109)	Dr: PINTO M.D.,DOUGLAS	DBR REG CLI			
Patient APPLE, ORA	NGE					
Req Number					Single	₿
					Worklist	12
Specimen Number	Status	Coll Date	Time		Enter/Edit	
					Enter/Edit/Res	EV
			<b>_</b>		Inquiry	R
					Cancel Req	Ex
	Orders	5			Uncancel Req	E?
					Move to Acct	GÓ
			<b></b>		Billing Inquiry	(6)
					Billing Edit	<u> (</u> 72
	Cancellation C	omments			Demand Bill	Ð
					Edit ICD Codes	88.8
					Reprint Waiver	89
					Edit LIS Pt Data	
					Edit MRI Markei	rs 📾
Cancel Req Wkld						
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### Select the Requisition radio button. Select OK.

😽 Requisition Lookup		
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	Deguisition	
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	O Order Group	
	<ul> <li>Specimen</li> </ul>	
	<ul> <li>Last Specimen</li> </ul>	
	<ul> <li>On/Before Date</li> </ul>	
ſ		
	Cancel	OK

## Select the requisition you wish to cancel.

Kequisition Lookup				_ IX
		00000050	ADDLE ODANCE 21/M	
Number	Chatria			
	Status			
0 00000145	ENTERED	01/20/10-1641	CBC, PT, PTT, BMP, UA, MAFP	
Submitted by: PIN		۵S		
Specimens: 0120:	C00014R RECD	0120:GS00004R R	ECD. 0120:H000078 RECD	
		Prior	Next	Cancel
				× )

#### All ordered tests will display in the middle of the screen. Enter cancellation comments.

Time Requisition D	Desktop - SLO/ML (Test) - Daniel Weige	er				
<b>Apple,O</b> 21/M 06/	<b>range</b> /09/1988	LO0000000059 (SA00000109) Req: 00000145	Dr: PINTO M.D.,DOUGLAS	DBR REG CLI		×
	Patient APPL Reg Number 0000	LE,ORANGE				Single
				_		Worklist
	Specimen Nu LAB 0120:H00007R	mber Status RECD	Coll Date 01/20/10 1	641 A		Enter/Edit
	LAB 0120:HC00003R	RECD	01/20/10 1	641		Enter/Edit/Res 🕅 Inquiry 🔍
		Order	з			Cancel Req 🗈
	CBC, PT, PTT, BMP, U	JA, MAFP		<b>^</b>		Move to Acct 🖂
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						Edit ICD Codes 🔤 Reprint Waiver 🔊
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	Cancel Reg Wikid					Edit MRI Markers 📾
				Cancel	Save √	? 🖻 🛇 🕆 🗉

Select F5 to search for canned comments. Enter the comment mnemonic in the textbox. If unknown, select F9 to view the full list of comments.



#### Enter search criteria into the textbox. Select the comment, once it is located.

Mnemonic	Name
OC1	
0010	
0011	CONTROL ONS
0C2	REPEAT/OC OUT OF RANGE
OC3	OC REPEATED OUT OF RANGE
0C4	TROUBLESHOOTING COMPLETED
QC5	QC RUN POST INSTRUMENT SERVICE
QC6	ASSAY RECALIBRATION
QC7	NEW CONTROL LOT
QC8	NEW REAGENT LOT
QC9	INCORRECT QC
<b>QCKIT</b>	QC KIT LOT # EXPIRATION DATE
QNS	QNS - PLEASE RESUBMIT
QRN	QUESTIONABLE RESULT

## Select Yes to confirm the cancellation.



## Select F12.

#### The entire requisition has been successfully cancelled.

