



STANDARDIZED TESTING / OPERATING PROTOCOL REQUEST/ANNOUNCEMENT

Annual Stago Lot Conversion

Description:	The laboratories will convert to the following lot numbers of PT, PTT, FIBR and controls: STA-Neoplastine Cl Plus Lot 251218 Exp 10/31/2018 STA-PTT Automate Lot 251274 Exp 10/31/2018 STA-Fibrinogen Lot 251221 Exp 10/31/2018 STA-Coag N&ABN Plus Lot 251136 Exp 09/30/2018
Date:	July 26 th , 2017 @ 10:00 am
Performing Locations:	Click on the boxes that apply: ⊠ Alamance Regional □ Alamance Cancer Center ⊠ Annie Penn Hospital ⊠ Moses Cone Hospital ⊠ Med Center at High Point ⊠ Med Center at Mebane ⊠ Wesley Long Hospital ⊠ Women's Hospital
Affected Locations:	Click on the boxes that apply: ⊠ Alamance Regional □ Alamance Cancer Center ⊠ Annie Penn Hospital ⊠ Moses Cone Hospital ⊠ Med Center at High Point ⊠ Med Center at Mebane ⊠ Wesley Long Hospital ⊠ Women's Hospital

	Click on the boxes that apply:
Affected Departments:	 □Blood Bank □Cytology □Flow Cytometry □Histology □Microbiology □Phlebotomy □Point of Care ⊠Rapid Response Lab □Respiratory Therapy □Specimen Processing
Specimen Type:	Sodium citrate plasma
Updated Clinical Lab Procedures:	Greensboro/Reidsville Procedures: COAG-0540-CH Parallel Testing with Coagulation COAG-0716C-CH Stago Information Sheet COAG-0841C-CH Stago Reagent Chart QM-1735L-CH Reference Ranges Alamance Procedures: COAG-710 PT COAG-710 PTT
Retired Clinical Lab Procedures:	N/A

Notification to Client:	Click on the boxes that apply: Section Not Applicable Memo Needed Distribution of Memo: Medical Staff Allied Health Professionals (PA, Nurse Practioners) Anesthesia Annie Penn (Primary Source Physicians) Dentist Emergency Department/Urgent Care Centers Family Practice Infectious Docs #ID Docs (John Campbell, Robert Comer, Jeffrey Hatcher, Cynthia Snider, Kees Van Dam) OB/GYN Pathology Pediatricians Psych Radiology Surgery #Nursing Leadership (Directors, Asst. Directors, Clinical Nurse Manager) Pharmacy - Send to DeAnne Brooks & Jim Hasspacher #IIM Residents Kim Helsabeck Philebotomy Managers and Supervisors
	 Phlebotomy Managers and Supervisors Point of Care: Sheila, Kim & Marty
Accreditation Section:	 Click on the boxes that apply: Section Not Applicable □CAP Test menu change needed □CMS Analyte form change needed □Proficiency Testing surveys changes needed or ordered
Laboratory IT section:	Click box and type needed changes/additions: □ Section Not Applicable ⊠LIS changes ⊠Reference range change/addition

	[
	Update the	following reference intervals:	
	$PT \rightarrow 11.4$	– 15.2 s	
	$\mathbf{PTT} \rightarrow 24$	– 36 s	
	$\mathbf{FIBR} \rightarrow 21$	0 – 475 mg/dL	
	$INR \rightarrow Re$	move reference interval	
		ll Failure change/addition	
	Critical V	√alue change/add	
	\Box Text con	nments needed	
		n collection instructions	
	\Box Need to 1	monitor TAT	
	\Box CPT cod	e for tests(s)	
	Update con	ntrol code IDs in Sunquest for al	l Stagos:
	C-COAN =	= Q12373	
	C-COAP =	= Q12374	
	The Cone H	Health laboratories will convert to	he new lot of PT, PTT, FIBR, and
	QC on July	26, 2017 at 10 am.	
	(No	te: Moses Cone will not switch to	FIBR Lot 251221. They previously
	veri	fied Lot 251088 and will continue	to use it until supply is low.
	Clai	rityCor system correlation perform	ed with 251088 at Moses Cone to
	prov	ve no statistical uniference between	The sites. All results acceptable.)
	While no re	eference interval adjustments were	required based on the lot conversion
	studies, the	laboratories will standardize refer	ence ranges at midnight on 7/26/2017.
	New Refer	ence Intervals:	
	PT	→ 11.4 – 15.2 s	
		New Geometric Mean: 13.2 s	
		New ISI: <u>1.25</u>	
Technical Staff	INR	$A \rightarrow$ No reference interval reported	
Update:	PTI	$2 \rightarrow 24 - 36 \text{ s}$	
	FIB	$R \rightarrow 210 - 4/5 \text{ mg/dL}$	
	In addition	the laboratories (EXCEPT MedC	anter Mehane) will switch to the
	COAN and	COAP Plus controls These new	controls are stable on the instrument for
	24 hours.	Corrier Flus controls. These new	controls are stable on the instrument for
	Control	COAN/COAP (Mebane Only)	COAN/COAP (All Other Sites)
	Volume	1.0 mL	2.0 mL
	Stability	8 hours	24 hours
	On the day	of the lot conversion, each site mu	st do the following:
	1 I.m.	late the reference intervals in the T	ast Mathodology socion of the
	I. UPC	value the reference intervals in the I	est methodology section of the
	alla	192015 101 1 1, 1 1 1, and FIDK as It	yuncu.

	 All sites (except MedCenter Mebane) will need to adjust the QC files in the instrument to reflect the change to the new Plus controls. Each site must updated each Stago with <u>new ISI 1.25</u> and <u>new geometric mean 13.2 s</u>. Before reporting the first patient tested after the conversion, each site must calculate the INR manually, compare to printed analyzer INR and document in lot conversion notebook. After go live, sites should <u>perform QC every 4 hours for 5 days</u>. See attached detailed instructions for more information. If any problems occur during the lot conversion, please contact Jackie Hobbins at 336-832-8397 or 919-523-1611.
STOP Initiator:	Jackie Hobbins
Alamance Medical Director Signature:	Quality Department will obtain signature: Jana (Rubinas M. D. 7/24/17
Greensboro/Reidsville Medical Director Signature:	Quality Department will obtain signature:

Approved and current. Effective starting 6/14/2016. 1719F (version 1.0) QM-1719F-CH STOP Request. Annoucement Template Cone Health Laboratories QM-1719F-CH

Lot Conversion Tasks:

- Build QC files in Sunquest. <u>Use historic standard deviations.</u> Note: Alamance has been running the new lot of QC on their ARSTA1 since Monday. You can look at their LJs to see how they are recovering to help gauge where your mean should be.
- 2. Delete Patient files on the Stago.
 - a. Stago Compact Max: Test Panel → Patient Analyses → Patient files → Select Waste Basket/Delete → Select All files
 - b. Stago Compact: Files menu \rightarrow Delete Patient Files \rightarrow Press F4 to delete
- 3. Adjust reference ranges as applicable in instrument for PT, PTT, and FIBR. Print screenshots of changes and file under tab 2 in your notebook behind applicable page.
 - a. Stago Compact Max: Methodologies → Select Test → Modify → Select Page 3 → Edit Usual Values section to new reference ranges.
 - b. Stago Compact: Setup → Tests → Select Test → Page Down to page 3 → arrow over the Usual Values and make necessary changes → ESC and Save before Quitting
- 4. Adjust QC in analyzer for new 24 hour stability Print screenshots of changes and file under tab 2 in your notebook behind applicable page.
 - a. Stago Compact Max: Follow attached instructions for Stago Compact Max analyzer.
 - b. Stago Compact: Follow attached instructions for Stago Compact.
- 5. Load reagents and QC.

Print screenshots of calibration screens for PT, PTT, and FIBR and file under tab 2 in your notebook behind applicable page.

Refer to COAG-0540-CH Parallel Testing with Coagulation

6. Perform INR verification

Remember to use a <u>scientific</u> calculator. You can change the calculator on Windows PCs to be scientific by selecting View \rightarrow Scientific. File INR verification under tab 2 in your notebook behind applicable page. Approved and current. Effective starting 6/14/2016. 1719F (version 1.0) QM-1719F-CH STOP Request.Annoucement Template Cone₇ Health Laboratories QM-1719F-CH

Post Lot Conversion Tasks:

1. Perform QC every 4 hours for 5 days to assess control stability.

See attached schedules to post at bench to help your techs: AP, AR, MC, MHP, WH, and WL Stago Lot Conversion QC Sign Off MedCenter Mebane Stago Lot Conversion QC Sign Off

2. At the end of 5 days, print LJs and adjust means to reflect recovery. Fill out QM-1508F-CH Quality Control Lot Verification form. This is an assayed control.

File signed QM-1508F in notebook under tab 2.

Please make all necessary adjustments by August 3rd. I will audit on August 4th to verify this has been done to prevent some issues we've seen in the past.

3. Continue to monitor QC during weekly and monthly LJ review and adjust if necessary.

Stago Compact Max Instructions for QC Setups

How to update the test set up for the STA Coag N and ABN Plus

1. Access the patient files from the Test panel, click Patient analyses and then Patient files .

Click

- a. Window Delete Files appears
- b. To delete all the files, select ALL FILES
- c. Click Delete
- 2. Once all of the patient files have been deleted, from the Test panel, select Methodologies.
 - a. Double click on the methodology that needs to be modified.
 - b. Once you have opened up the test methodology, click on vor on the right side of the page.



c. Once on page 3 of the methodology, you will need to change the "**Identity**" of the QC that you will be using. (For the Coag N and ABN Plus, the new identity is 12373 for Coag N and 12374 for Coag ABN).

Quality controls Lev: Identity Key Name Period Stab Vol. Min: vol. (mi) 1 12349 7 STA-COAG CONT N 6 8 1.00 0.50 2 12353 4 STA-COAG ABN 6 8 1.00 0.50 3				/					PT	T: Prothr	ombin Tin	ie	
Lev. Identity Key Name Period (h) Stab. (h) Vol. (ml) Min.vol. (ml) 1 12349 7 STA-COAG CONT N 6 8 1.00 0.50 2 12353 4 STA-COAG ABN 6 8 1.00 0.50 3 STA-COAG ABN 6 8 1.00 0.50 3 STA-COAG ABN 6 8 1.00 0.50 3 .				(Quality contr	ols							6
1 12349 7 STA-COAG CONTIN 6 8 1.00 0.50 2 12353 4 STA-COAG ABN 6 8 1.00 0.50 3 3 3 6 8 1.00 0.50 Unit: Conversion factor Were Prince: Transmission rank Na 9 1 Minimum 11.00 sec None 1 Printout limits Minimum 10.00 sec		Identity											23
2 12353 1 STA-CCACABN 6 8 0.00 0.50 3 STA-CCACABN 6 8 0.00 0.50 3 0.50 3 0.50 3 0.50 10 <		12349	7				6			C	0.50		3
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d. Next, you will need to change choose the "Key" for the QC. Click the _____ and a dropdown box of options will appear. Choose the appropriate test option for the methodology you are in, i.e. Neoplastine CI Plus sec.

			BARCO	DE			
			Product reading key	12373			6
		Key	Key paramet	ers			C-s
	12373	0	2 STANEO CLISS			0.50	5
	12374	44	3 STA-NEO CI PLUS %			0.50	Co
			4 STA-NEO CI PLUS sec				
			5 STA-NEOPLASTIN R %	/			
1		Param	6 STA-NEOPLASTIN R sec		alues		
		sion factor	7 STA-PTT A		11.00	sec	
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Ref. T					0.00	SPC	
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		*	Quality controls						
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	12353	4		6	/8		0.50	3	
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- e. The **Name** should self-populate when you type in the identity.
- f. Next, adjust the Period, Stability, Volume and Min. Volume as needed.
 Period = 8 hours
 Stability = 24 hours
 Volume = 2 ml

Min. Volume = 0.5ml

g. Complete the same information for the Coag ABN Plus.



and save the Test Set Up changes.

METHO printou	DOLOGIE t/transmis	S – Quali sion	ty controls and				03/01/2016 09:23	2
						PT+ \$:	Prothrombin Time	C
			Quality controls	5				6
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	12374	ñ		8			0.50	C
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INR	<u>*</u>		e z .		Pri	ntout lim	its	
Ref.T	•		2		Minimum	0	.00 sec	
None	-				Maximum	120	.00 sec	
								E

It will tell you that the following actions will be performed and wants you to type "Yes" to accept.

1ETHO printo <u>ut</u>	DOLOGIE t/transmis	S – Qua sion	lity controls and				02/25/2016 07:03	2
			•			N	EW PT1: NEW PT	fi 🕕
			Quality control	s				ŏ
Lev.								23
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			IMPORTANT CHANGES D	ETECTED				
-	_		THE FOLLOWING ACTIO	NS WILL BE F	PERFORMED	1900		-
DAGA		Param	Calibration			104	es 	
Unit		sion lactor	Quality control		Delete	10	0.00 sec	
Raw (sec)			Please type YES to a	ccept:		20	0.00 sec	
None	•					lin	nits	
None	•		Confirm	Abo	ort		0.00 sec	
None	•				Maximum	30	0.00 sec	
								m

Click on Confirm.

(Note: verify that there are no controls on page 2 of the test setup. If there are controls defined on page 2 (calibration page you must change them as well to make them match the ID's above on third page of test setup)).

- 3. Make the same adjustments to each Methodology that will use the new STA Coag N and ABN Plus QC.
- 4. Load the new QC and run QC for each test.
 - a. If you are using the same lot of reagent, but changing QC only, you will need to "recalibrate". For this example, D-Dimer is selected.
 - b. From the Calibration Page, click on the Test ID of interest



c. From the Calibration page 1, select



d. Enter the Access/Password Code and confirm.

ACC	ESS CODE
lease enter your ac	cess code:
Confirm	Cancel

e. Calibration page 2 will pop up and you can then select the lot number you want to calibrate:

Identity	Name	Lo
12048	TAMP./BUF. D-DI	113823
		113823
		114105
		114477
12049	LATEX D-DI	114828

f. Once you have selected the correct lot, the drop down field will populate

	Reagents	
Identity	Nane	
12048	TAMP./BUF. D-DI	11447
12049	LATEX D-DI	11447

The instrument will run the new calibration with the new QC.

¹⁵ Stago Compact Classic Instructions for QC Set Up

How to update the test set up for the STA Coag N and ABN Plus -- Classic Compact



1. Access the patient files from the Test panel, click Files and then Delete Patient files.

 You will need to select the files to be deleted (all files), using F1 for the beginning of the list and F2 for the End of the list. Select F10 to Execute and type in YES when prompted.

1 lg. 4 *	I THE VELEC	TION THINKIN
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1	FILE SELECTION			
	From 1547 1		Filtering	
	F1 = Beginning of List	3) г	Prefix	File Status ((
	To 5462 2			Confirmed []Complete/Error
	FZ = End of List 3		4	[]]Complete []Incomplete 🔓
-	F3 = By Cursor	L		
	F10 Execute	Esc Qu	it 👔	🕴 Field Selection
ļ				



3. Once all of the patient files have been deleted, from the **Test panel**, select **Setup**, **Tests**

a. Double click on the methodology that needs to be modified. In these directions we will be looking at Fibrinogen.

		TEST SETUP	2
Fibringen	3		
PT	PS COAG		
PTT	HNF COLO		
FIB	HBPMCOLO		
TT	HNF COAG		
11	HBPM		
V	TP +		
UTI-X	• F8 1:20		
011	• F8 1:40		
×	• F8 1:80		
0111	• F9 1:20		
IX	• F9 1:40		
XI	• F9 1:80		
XII			
AT III			
PC COLO			
PC COAG			

			PRIM	NT QC			
Interpretation Interpretatio Interpretatio Interpretation Interpretation Inte	e t Test	F6: Print			TEST	SETUP - Page 1/3	
Esc: Quit				Late	est Modific	cation : 06 09 2007	A STREET
Abbreviation Name	FIB Fibrinoge	n 2mL	dentifi	ication M	lethod clo	nt-based	
Samp 1	e				Diluant		
Volume Inc	u. Dil	. ID	N	ame	Vial	Stab. Min.Volume	
100 11 240	sec 1/20	11361	Beagen	KOLLER ts	15 ml	144 h 0.90 ml	
ID	Name	Incub	Vol. V	ial Stab	Min.Volume	Washing	
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Rb		186					1
RC	MA DID O		-				in order to not waist
Rd 12215	STA-FIBZ		50 Result	2 192	0.50	no special	pouring off 5mls at a
Min. Time	4	Primary Un	its	mg∕dl	Min.	Max.	1 Make up Neo as us bottle.
Max. Time	80	Corrector	145445	1.000	60.0	0 1200.0	2. Pipet off 5mls of Ne
Rd Heating	30 No	Single/Dup Precision	licate	10.0 ×	Redil.	Condition (mazdl)	3. This Neo has an exp
Stirring	No		coef 1	0.00000	1/8	< 150.00	on the bottle the expi
Clot Type	low		coef2 offset	1.00000	1/40	> 800.00	4. Put a reducer and st of Neo then put it on t
Contra da		-	_				5. You must change th you put it on the analy.
						No. of Concession, Name	6. Once the Neo has be
						and the second	the 96hr expiration rem
							 Pour the remaining f bottle and put back on t forget to change the am

b. Once you have opened up the test methodology, in this case Fibrinogen, page down to page 2.

- c. On page 2 of Fibrinogen, you will need to change the "Identity", "key", "vial", "stability" and "min. vol." of the QC that you will be using.
- d. For the Coag N and ABN Plus, the new identity is 12373 for Coag N [Level 1] and 12374 for Coag ABN [Level 2]).

PgUp Previous PgDn Next Pag Esc Quit	s Page je		Test Setup - Page :	2/3
	C	ALIBBATION	TTO TOTTOGEN ZMI	
MOD	B Barcoded	mode	Scalı	
Vicw Poi	150.00 300.00 nts 600.00 800.00	mg/d1	Baw Data Concentration	Log
Control Leve	S ID 1 1 12349 1 2 12353	Key Name 4 STA-CDAG CONT P 7 STA-CDAG ABN	Uial Stab. Min.Uc ml h (ml N 1 B 0. 1 8 0.	lume) 50 50
Phase and a second seco				

e. Next, you will need to choose the "**Key**" for the QC. Click the **DEL** key and a drop-down box of options will appear. Choose the appropriate test option for the methodology, in this case, scroll down to #10 and Enter.



- f. Next, adjust the Stability, Volume and Min. Volume as needed. Stability = 24 hours
 Volume = 2 ml
 Min. Volume = 0.5ml
- g. Page down to page 3 of the test set up



- h. You will need to input the correct **ID** for the Coag N (12373) and Coag ABN (12374) and select the correct **KEY** just like on page 2.
- i. The Name should self-populate when you type in the identity.
- j. Next, adjust the **Period**, **Stability**, **Volume** and **Min. Volume** as needed.

Period = 8 hours Stability = 24 hours Volume = 2 ml Min. Volume = 0.5ml k. Here is how the new 24 hour Coag N and Coag ABN should look:

	Quality Control				Vial	Stab.	Min.Volume
	ID	Key	Name	h	nl	h	(ml)
Level 1 Level 2 Level 3	12373 12374	4	STA-COAG N+ STA-COAG ABN+	8 8	2.00 2.00	24 24	0.50 0.50

I. Click ESC save the Test Set Up changes, and enter your access code.





It will tell you that the following actions will be performed and wants you to type "**Yes**" to accept. (the screen will look very <u>similar</u> to this picture...)

1ETHO rintout	DOLOGIE /transmis	S – Qua sion		02/25/2016 07:03	2			
						NE	W PT I : NEW PT	
Ì			Quality controls					ŏ
Lev.			Name					23
1	LI		SETU	P			0.20	3
	L2		NEW PT	-1			0.20	3
31			NEW FI					
			IMPORTANT CHANGES DETE		CODMED			
		Param	THE FOLLOWING ACTIONS	WILL BE FER	FORMED	lue	20	
		r ar ar r	Calibration		~	10		
		~~~~~	Quality control		Jelete	20	).00 sec	
Raw (sec)			Please type YES to acce	pt:				
None			Confirm	Abort		lim	its	
None	<u> </u>					C	).00 sec	
None	·			ſ	1aximum	300	).00 sec	
								-

- 4. Make the same adjustments to each Methodology that will use the new STA Coag N and ABN Plus QC. PT and PTT may not have controls in page 2 of the test set up and that is ok...simply go to page 3 and proceed.
- 5. Load the new QC and run QC for each test.
  - a. If you are using the same lot of reagent, but changing QC only, you will need to "recalibrate".
  - b. From the Calibration page 1, select Fibrinogen using F1. Then select F10 to run the Calibration.



The instrument will run the new calibration with the new QC.

## AP, AR, MC, MHP, WH, WL

Stago Lot Conversion QC Sign Off						
	N/A	N/A	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	N/A	N/A	N/A
Tech						
*Shade	ed time	s indica	ate whe	en new	QC is t	o be m

# **COAN and COAP must be made with 2 mL** of reagent grade water.

# MedCenter Mebane

Stago Lot Conversion QC Sign Off						
	N/A	N/A	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	1400	1800	2200
Tech						
	200	600	1000	N/A	N/A	N/A
Tech						
*Shade	ed time	s indica	ate whe	en new	QC is t	o be ma



TO: Medical and Pharmacy Staff

FROM: John Patrick, MD, FCAP, FASCP

John Patrick, MD, FCAP, FASCP Greensboro and Reidsville Hospital Laboratories Medical Director, Cone Health Tara Rubinas, MD, FACP, FASCP Medical Director, Anatomic and Clinical Pathology Laboratories, Alamance Device Data (Rubinas MD 7/24/17)

DATE: July 20, 2017

The Cone Health hospital laboratories conducted new coagulation reagent correlation studies and found no clinically significant differences between the two lot numbers. While typically this would indicate no need for reference range updates, we are pleased to announce that minor reference range adjustments will be made to standardize the Greensboro/Reidsville and Alamance campuses.

The laboratory will update the ISI of the new PT reagent (Lot 251218) to 1.25 on July 26, 2017 at 10:00 am.

Assay	Current Range Greensboro	Current Range Alamance	Range on 7/26/2017	Units
Unfractionated Heparin	0.30 - 0.70	0.30 - 0.70	0.30 - 0.70	IU/mL
Low Molecular Weight Heparin	0.50 - 1.20	N/A Not Reported	0.50 - 1.20	IU/mL
Prothrombin Time (PT)	11.6 - 15.2	11.4 – 15.0	11.4 - 15.2	Seconds
Partial Thromboplastin Time (aPTT)	24 – 37	24 - 36	24 - 36	Seconds
Fibrinogen	204 - 475	210 - 470	210 - 475	mg/dL

The following assays are available to monitor anticoagulants:

Assay:	Anticoagulant:
Prothrombin Time (PT)	Warfarin (Coumadin)
Activated Partial Thromboplastin Time(aPTT)	*Unfractionated Heparin, Direct Thrombin Inhibitors (Bivalirudin and Argatroban)
Heparin Assay (Anti-Xa)	*Unfractionated Heparin
Low Molecular Weight Heparin (Anti-Xa)	Enoxaparin, Dalteparin, Tinzaparin

#### Recommendations for monitoring anticoagulant therapy:

For an anticoagulant naïve patient a baseline PT and aPTT should be performed before choosing anticoagulant therapies. If the baseline results are not within the normal range, it is recommended that a patient risk assessment be done before proceeding with therapy. Direct oral anticoagulants (apixaban, edoxaban, dabigatran or rivaroxaban) interfere with routine coagulation tests. Interpretation of lab results should be done with caution without knowing the time of when the patient's last dose was taken.

*Heparin levels are measured by an anti-Factor Xa assay and reported in IU/mL of activity. This is a direct measurement of the patient drug level and avoids the lack of specificity inherent in the aPTT assay. Studies indicate that monitoring of heparin therapy using anti-Xa levels, as opposed to the aPTT, can more quickly achieve patient therapeutic ranges and shorten hospital stays. If a patient heparin level is not in the expected range, and patient dosage has been confirmed, the Antithrombin III Assay is available to assess possible heparin resistance.

Monitoring of the Factor Xa Inhibitors Fondaparinux, Rivaroxaban, Apixaban, and Edoxaban appear to be unnecessary for most patients. Assay techniques and target ranges for FXa Inhibitors have not been rigorously standardized and there is very little information relating anti-Xa levels to clinical outcomes. Currently, Cone Health Laboratories do not perform specially-calibrated assays to monitor FXa Inhibitors.

If there are any questions, please contact Dr. John Patrick at 832-7531 / jdpatrick@auroradx.com or Dr. Tara Rubinas at 538-7832 / tara.rubinas@conehealth.com

	Alama	nce Region 1240 Huffr Burlingt	nal Medical Ce man Mill Rd. ton, NC 27215	enter		INTERIM RE	PORT	
NAME: MRN : ACCT:	TESTING,LAB 123456789 111	LOC: EDA MD: DEFAU	ULT, PROVIDER		AGE: 31Y DOB: 07/	SEX: 15/1986	Μ	
M275	COLL: 07/24/2017	11:50 RI	EC: 07/24/2017	11:51	PHYS: DE	FAULT, PROV	VIDE	
FIBI	RINOGEN	н	477	[2]	10-475]	mg/dL		$\{AH\}$
PRO: PI IN	THROMBIN TIME ROTHROMBIN TIME NR	н	16.2 1.97	[1]	1.4-15.2]	seconds	{AH}	{AH}
aPT	ſ	н	40	[24	4-36]	seconds		${AH}$
{ 2	AH} = Performed at	Alamance H	Hospital Lab,	1240 Hu	uffman Mi	11 Rd.,		

Burlington, NC 27215

Jula Pateral, M7 7/25/17

Jara (. Rubina, M.id. 7/24/17

TESTING, LAB PRINT DATE: 07/24/2017 END OF REPORT PRINT TIME: 12:01 PAGE 1

INTERIM REPORT Moses Cone Memorial Hospital 1200 N. Elm Street Greensboro, NC 27401 NAME: TESTING, LAB MRN : 123456789 AGE: 31Y SEX: M LOC: MAJO ACCT: 111 MD: DEFAULT, PROVIDER DOB: 07/15/1986 M274 COLL: 07/24/2017 11:49 REC: 07/24/2017 11:50 PHYS: DEFAULT, PROVIDE L 208 [210-475] mg/dL  $\{MC\}$ FIBRINOGEN PROTHROMBIN TIME [11.4-15.2] seconds H 15.5 {MC} PROTHROMBIN TIME {MC} 1.78 INR н 38 seconds {MC} [24-36] aPTT IF BASELINE aPTT IS ELEVATED, SUGGEST PATIENT RISK ASSESSMENT BE USED TO DETERMINE APPROPRIATE ANTICOAGULANT THERAPY.

{MC} = Performed at Moses Cone Hospital Lab, 1200 N. Elm St., Greensboro, NC 27401

Jule Paterne, MS 1/25/17

Jana C. Per MD 7124117

TESTING, LAB PRINT DATE: 07/24/2017 PRINT TIME: 12:00

END OF REPORT

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Ala	mance Regional Medical Ce 1240 Huffman Mill Rd. Burlington, NC 27215	nter INTERIM REPORT	2
NAME: TESTING,LAB MRN : 123456789 ACCT: 111	LOC: MUC MD: DEFAULT, PROVIDER	AGE: 31Y SEX: M DOB: 07/15/1986	
M276 COLL: 07/24/202	17 11:51 REC: 07/24/2017	11:52 PHYS: DEFAULT, PROVIDE	
FIBRINOGEN	306	[210-475] mg/dL {A	н}
PROTHROMBIN TIME PROTHROMBIN TIME INR	H 15.7 1.78	[11.4-15.2] seconds {U	{UC} C}
aPTT	35	$[24-36]$ seconds $\{A$	н}
<pre>{AH} = Performed at Burlington, {UC} = Performed at Blvd., Meban</pre>	Alamance Hospital Lab, 1 NC 27215 Mebane Urgent Care Cente e, NC 27302	.240 Huffman Mill Rd., er Lab, 3940 Arrowhead	

Julen Paterne, 100 7/25/17

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Spra (. Rulsmas M. D. 7/24/17

TESTING, LAB PRINT DATE: 07/24/2017 END OF REPORT PRINT TIME: 12:01

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