

LRN Protocol Change Bulletin 11/1/18

Overview of *Brucella* spp. Isolation and Identification

- Acceptable Samples types updated, Section 2.a.1 (pg 1)

Acceptable samples

- (1) Appropriate clinical specimens (e.g. Blood, serum, bone marrow, joint fluid, cerebrospinal fluid, abscess fluid, tissue masses, breast milk)
- (2) Environmental/nonclinical samples
- (3) Culture isolates

- Table 1 updated to include RB51 (pg 2)

Table 1. Confirmation of *Brucella* spp. by biochemical and molecular tests

Strain	H ₂ S production	Urease		Gel formation	Lysis by Tbilisi		Real-time PCR (2 of 3 targets)
		<5 min	>5 min		RTD	RTD x10 ⁴	
<i>B. melitensis</i>	No	Variable	Yes	No	No	No	Positive
<i>B. abortus</i>	Variable	No	Yes	No	Yes	Yes	Positive
<i>B. abortus</i> RB51	Yes (may be weak)	No (may be weak)	Yes	No	No ¹	No ¹	Positive
<i>B. suis</i> ²	Variable	Yes	Yes	No	No	Yes	Positive
<i>B. canis</i>	No	Yes	Yes	Yes	No	No	Positive

¹Different from *B. melitensis*, there is no "ghost lysis"

²Some *B. suis* demonstrate no lysis (i.e. resistance) at the RTD x 10⁴ concentration of Tbilisi phage

Isolation of *Brucella* spp. from Human, Animal and Environmental Samples

- Breast Milk added to sample type, Section 2.a.1.b (pg 1)
- Updated tissue processing- now use tissue grinder, Section 5.b. (pg 3)

Gel Formation Test (*Brucella canis*)

- Older culture works better should use a 48-72 hr old culture for test, Section 2.a. (pg 1)
- Phenolized Buffered Saline now stored at room temperature and will be good for 5 years, Section 3.a.
- Changed control to *B. canis* from NVSL order number BRU-NS, Section 4.a. (pg 2)

LRN Protocol Change Bulletin 11/1/18

Hydrogen Sulfide Production

- **Updated QC**
Quality control Section 4.a.1,2 (pg 2)
 - a. Controls
 - (1) Positive control: *B. abortus*, strain 19 or *Escherichia coli* (ATCC 25922) or equivalent
 - (2) Negative control: *Pseudomonas aeruginosa* (ATCC 27853) or equivalent
 - b. Quality control testing
 - (1) Test each new lot of lead acetate paper with positive and negative control strains.
 - (2) Use both a negative and a positive control organism with each run.

Susceptibility to *Brucella* phage Tbilisi

- Changed controls used, Section 3.b.(2).a,b,c (pg1)
- Changed protocol – Now use 0.5McFarland's inoculum, Section 5.a.(2) & 5.b.(1)
- Changed observation period/intervals, Section 5.b.(7) (pg 3)
 - Examine at 48 hrs and daily thereafter for up to 4 days.

Urease Test (Christensen's Method)

- Added a note to positive control reaction, Section 6.a.
 - NOTE: All *Brucella* spp. except *Brucella ovis* should produce a positive reaction after overnight incubation.

Slide Agglutination for Detection of *F. tularensis* Cellular Antigens

- Updated control that is used Section 3.2.a.
 - *Francisella tularensis* LVS cells

Antimicrobial Susceptibility Testing of *Yersinia pestis* by Etest

- Changed Etest order numbers Section 3.b.a-d
 - Reagents
 - (1) Etest strips (bioMérieux) for the following antimicrobial agents
 - (a) Ciprofloxacin (catalog #412310)
 - (b) Gentamicin (catalog # 412367)
 - (c) Levofloxacin (catalog #412392)
 - (d) Tetracycline (catalog #412470)
- Store Etests according to manufacturer's recommendations. Do not use after the expiration date.

LRN Protocol Change Bulletin 11/1/18

The Non-variola *Orthopoxvirus* Real-time PCR Assay

- Has 510(k) clearance by the FDA as a Class II medical device
- Added RP positive control to table 1, Section 4. Table 1

Table 1. Use of primer and probe sets with samples and controls

		VAC1	RP	16S
1	Clinical Specimen DNA	Yes	Yes	Optional (See section 6.a.(4))
2	Viral culture lysate DNA	Yes	No	Yes
3	No Template Control (NTC)	Yes	Yes <i>If DNA from clinical specimens is included in the run</i>	Yes <i>If DNA from viral cell culture lysates is included in the run</i>
4	Agent Positive Control	Yes	No	No
5	RNase P Positive Control	No	Yes <i>If DNA from clinical specimens is included in the run</i>	No

- RP positive control should be <30 CT, Section 4(5) (pg 7)
- Quanta mastermix is now approved details on pages 9-13
- Flow chart added for interpretation of results page 19
- Added RP Positive requirements samples must be <40 CT page 22

Note: This assay can't be run until a validation is complete