



University of Washington Medical Center 1959 NE Pacific Street. Seattle, WA, 98195 Transfusion Services Laboratory Policies and Procedures Manual	Original Effective Date: 02-11-2016	Number: PC-0020.02
	Revision Effective Date: 06-20-2019	
TITLE: ABO/Rh Manual Tube Method		

PURPOSE:

Provide instructions for determining the ABO and Rh type of patient using a manual tube technique

PRINCIPLE & CLINICAL SIGNIFICANCE:

Principle

Plasma and cells from patients are tested with commercial reagents to determine if the A, B and/or D antigens are present on the red cells and if the corresponding A and B antibodies are present in the plasma

Clinical Significance

Transfusion of ABO/Rh-incompatible blood can be associated with acute intravascular hemolysis, renal failure and death. Likewise, transplantation of ABO-incompatible organs is associated with acute humoral rejection. Because of clinical consequences associated with ABO incompatibilities, ABO/Rh typing is the foundation of pretransfusion testing and important before transplantation

POLICY:

- Weak D testing (one time) is required when initial typing with Anti-D is
 - Nonreactive for the following patient categories:
 - Cord blood and neonate specimens (for maternal Rh immune globulin eligibility)
 - Samples from donors or potential donors of blood or stem cell components (this does not include red blood cell component type confirmation testing)
 - Discrepant or discrepancies exists between the current and historical ABO/Rh. These discrepancies must be resolved before issuing type specific blood components (refer to SOP *ABO and Rh Discrepancy*)
- Plasma components and platelets may be released after one ABO/Rh performed by the University of Washington Transfusion Service Laboratory (UWTSL) is on record
- Historical ABO/Rh testing from Bloodworks Northwest (BWNW) shall not be used for compatibility testing or allocating any blood components to patients

SPECIMEN REQUIREMENTS:

EDTA is preferred and if not tested soon after collection, should be stored at 1-6°C
 Red top clotted blood samples are also acceptable
 See SOP *Specimen Acceptability*

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REAGENTS/SUPPLIES/EQUIPMENT:

Reagents:	Supplies:	Equipment:
<ul style="list-style-type: none"> • anti-A • anti-B • anti-D • A₁ cells • B cells • ABO + Rh Control • Blood Bank Saline 	<ul style="list-style-type: none"> • 12 x 75 glass tubes • Blood bank transfer pipettes 	<ul style="list-style-type: none"> • Calibrated serologic centrifuge • Calibrated cell washer • Agglutination viewer

QUALITY CONTROL:

Quality Control is performed daily

INSTRUCTIONS:

TABLE OF CONTENTS:

- [Adult ABO/Rh Manual Testing](#)
- [Neonate ABO/Rh Manual Testing](#)

Adult ABO/Rh Manual Testing

STEP	ACTION
1	Label 7 test tubes per SOP <i>Labeling for Manual Testing</i> <ul style="list-style-type: none"> • 4 forward type • 2 reverse type • 1 patient cell suspension
2	Add 1 drop of reagent anti-A, anti-B, anti-D and ABO/Rh control to the respectively labeled tubes for forward type
3	Add 2 drops of patient plasma/serum to tubes labeled for reverse type
4	Prepare an approximate 3-4% cell suspension using patient red cells in the respectively labeled tube
5	Add 1 drop of the 3-4% patient's red cell suspension to tubes labeled for the forward type
6	Add 1 drop of reagent A ₁ cells and reagent B cells to the respectively labeled tubes for the reverse type
7	Mix gently and centrifuge for time posted
8	Resuspend the cell button and examine for agglutination and/or evidence of hemolysis
9	Read, grade and record the reactions (refer to SOP <i>Grading Reactions</i>)
10	Go to section Interpreting & Reporting Results

Neonatal ABO/Rh Manual Testing

STEP	ACTION
1	Label 5 test tube per SOP <i>Labeling for Manual Testing</i> <ul style="list-style-type: none"> • 3 forward type • 1 patient cell suspension
2	Add 1 drop of reagent anti-A, anti-B, anti-D and ABO/Rh control to the respectively labeled tubes for forward type.
3	Prepare an approximate 3-4% cell suspension of patient cells in the respectively labeled tube.
4	Add 1 drop of the 3-4% patient's red cell suspension to tubes labeled for forward type.
5	Mix gently and centrifuge
6	Shake gently to resuspend the cell buttons, and examine for agglutination and/or evidence of hemolysis.
7	Read, grade and record the reactions (refer to SOP <i>Grading Reactions</i>)
8	Go to section Interpreting & Reporting Results

CALCULATIONS/INTERPRETATIONS/RESULTS REPORTING/NORMAL VALUES/CRITICAL VALUES

Interpretation

Table 1: ABORh Result Interpretation

ABORh Result interpretation							
Forward grouping		Rh D Typing		Reverse grouping**		Resulting in SQ	
Anti-A	Anti-B	Anti-D*	Control	A ₁ cells	B cells	Interp.	Sunquest Entry
0	0	+	0	+	+	O POS	O,P
0	0	0	0	+	+	O NEG	O,N
+	0	+	0	0	+	A POS	A,P
+	0	0	0	0	+	A NEG	A,N
0	+	+	0	+	0	B POS	B,P
0	+	0	0	+	0	B NEG	B,N
+	+	+	0	0	0	AB POS	L,P
+	+	0	0	0	0	AB NEG	L,N

ND = not done
 * Perform Weak D testing prior to reporting ABO/Rh result if reaction with anti-D is:

- negative on a cord blood or neonate samples
- Negative on a donor sample

 ** Reverse grouping is not performed on neonatal samples or patients < 4 months old and is resulted as INT (infant not tested) in the testing grid

Results Reporting in Sunquest

STEP	ACTION															
1	Select the test in “Blood Order Processing” (BOP)															
2	Enter the reactions in the ABO/Rh grid															
3	<p>Complete the ABO/Rh interpretation except for the following circumstances:</p> <table border="1" data-bbox="302 541 1419 1373"> <thead> <tr> <th data-bbox="302 541 537 642">If the specimen is from a(n)</th> <th data-bbox="537 541 846 642">And</th> <th data-bbox="846 541 1419 642">Then</th> </tr> </thead> <tbody> <tr> <td data-bbox="302 642 537 810">Any patient</td> <td data-bbox="537 642 846 810">Reactions do NOT match the ABO/Rh Interpretation Tables and/or are discrepant from previous results</td> <td data-bbox="846 642 1419 810"> <ul style="list-style-type: none"> • Click <Accept> • Resolve discrepancy before entering the ABORH interpretation (See SOP <i>ABO/Rh Discrepancy</i>) </td> </tr> <tr> <td data-bbox="302 810 537 947">Cord Blood specimens</td> <td data-bbox="537 810 846 947">Rh D negative</td> <td data-bbox="846 810 1419 947"> <ul style="list-style-type: none"> • Click <Accept> • Perform Weak D testing before entering interpretation (refer to SOP <i>Weak D Testing</i>) </td> </tr> <tr> <td data-bbox="302 947 537 1062">All SCCA & all female patients < 50 y.o.</td> <td data-bbox="537 947 846 1062">1+ Rh D Reactivity</td> <td data-bbox="846 947 1419 1062"> <ul style="list-style-type: none"> • Report as Rh negative • Add a PB comment: WKDP = Weak D positive </td> </tr> <tr> <td data-bbox="302 1062 537 1373">Prenatal specimens (PREN)</td> <td data-bbox="537 1062 846 1373">1+ Rh D Reactivity</td> <td data-bbox="846 1062 1419 1373"> <ul style="list-style-type: none"> • Report as Rh negative • Add a PB comment: WKDP = Weak D positive • For new weak D positive results on prenatal samples, add a BBC comment: RHREC = Patient is a candidate for Rh Immune globulin. Further D antigen characterization can be obtained with Rh D genotyping. </td> </tr> </tbody> </table> <p>NOTE: If patient has a previous history of Weak D testing documented in the LIS, repeat weak D testing is not required.</p>	If the specimen is from a(n)	And	Then	Any patient	Reactions do NOT match the ABO/Rh Interpretation Tables and/or are discrepant from previous results	<ul style="list-style-type: none"> • Click <Accept> • Resolve discrepancy before entering the ABORH interpretation (See SOP <i>ABO/Rh Discrepancy</i>) 	Cord Blood specimens	Rh D negative	<ul style="list-style-type: none"> • Click <Accept> • Perform Weak D testing before entering interpretation (refer to SOP <i>Weak D Testing</i>) 	All SCCA & all female patients < 50 y.o.	1+ Rh D Reactivity	<ul style="list-style-type: none"> • Report as Rh negative • Add a PB comment: WKDP = Weak D positive 	Prenatal specimens (PREN)	1+ Rh D Reactivity	<ul style="list-style-type: none"> • Report as Rh negative • Add a PB comment: WKDP = Weak D positive • For new weak D positive results on prenatal samples, add a BBC comment: RHREC = Patient is a candidate for Rh Immune globulin. Further D antigen characterization can be obtained with Rh D genotyping.
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NOTES AND LIMITATIONS:

- 3-4% suspension of RBCs may be prepared by one of the following:
 - Using volume estimation with comparison to the reagent red cells for visual verification
 - Adding one drop of packed RBCs to approximately 1-1.5 mL of blood bank saline
- Positive reactions characteristically show 3+ to 4+ agglutination by reagent ABO antibodies; reactions between test serum and reagent red cells are often weaker. Any discrepancy between the results of the tests with serum or plasma and red cells must be resolved before an interpretation is recorded for the patient’s or donor’s ABO group.
- Mixed-field agglutination should be investigated for possible cause prior to interpreting results.
- Positive ABO/Rh control invalidates test. Refer to SOP *ABO/Rh Discrepancy*

REFERENCES:

- Technical Manual. Bethesda, MD: AABB Press, current edition
- Standards for Blood Banks and Transfusion Services. Bethesda, MD: AABB Press, current edition
- Reagent Manufacturer's Insert; current version

RELATED DOCUMENTS:

SOP Specimen Acceptability
SOP Quality Control of Manual Bench Reagents
SOP Labeling for Manual Testing
SOP Grading Reactions
SOP ABO/Rh Discrepancy
SOP Weak D Manual Tube Testing

APPENDICES:

NA

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UWMC SOP Approval:	
UWMC CLIA Medical Director	_____ Date _____
	Mark H. Wener, MD
Transfusion Service Manager	_____ Date _____
	Nina Sen
Compliance Analyst	_____ Date _____
	Christine Clark
Transfusion Service Medical Director	_____ Date _____
	Monica Pagano, MD
UWMC Biennial Review:	
	_____ Date _____
	_____ Date _____

06/20/19 – Revised as part of Ortho Gel Method implementation to add a control tube to all ABO/Rh to be standard with the gel method and result entry in the LIS.