



University of Washington Medical Center 1959 NE Pacific Street. Seattle, WA 98195 Transfusion Services Laboratory Policies and Procedures Manual	Original Effective Date: 06-20-2019	Number: PC-0072.01
	Revision Effective Date:	
TITLE: Ortho Vision® Resource Management and Daily Quality Control		

PURPOSE:

To provide instruction for performing reagent quality control (QC) on the Ortho Vision® and the use and handling of reagents, diluents, MTS gel cards and other materials.

PRINCIPLE & CLINICAL SIGNIFICANCE:

A quality control program is established to ensure reagents; antisera and the Ortho Vision® perform as expected prior to reporting of patient or blood component test results. Validity of routine test results are evaluated by testing the reagents with a commercially available QC kit and samples prepared according to manufacturer's recommendations. Quality control testing is considered acceptable if valid positive and negative results are obtained for each reagent tested.

Resources used with the analyzer must be tracked, monitored and discarded as per manufacturer requirements and expiration dates. The Resources screen is used to monitor, discard, and replenish resources as needed.

POLICIES:

- Quality control (QC) testing is performed:
 - Each day of use
 - Whenever a new reagent lot number is placed into use – this includes MTS Gel Cards
 - After maintenance (daily, weekly, monthly, yearly)
 - After service or repair of the analyzer
- QC testing is performed daily on both analyzers in use. Quality control results of a reagent on one analyzer, is not acceptable for use on the 2nd analyzer unless QC is repeated on the 2nd analyzer prior to reporting of patient or blood component test results.
- Unexpected results are investigated and resolved prior to reporting patient or blood component test results
- Each lot of reagent and MTS gel card in use is quality control tested and must meet acceptance criteria prior to verification and reporting of patient or blood component testing. Patient and blood component samples may be run in parallel with the control samples as long as this requirement is met.
- All antisera and cellular reagents are stored per manufacturer instructions when not in use
- Preparation of ABO/Rh and antibody screen controls may be necessary if AlbaQ-Chek samples are unavailable, expired or at an inadequate volume
- **Table 1: MBC QC Test Profiles, Reagents, QC Samples** lists the QC Test Profiles required for daily QC and the MTS® Gel Cards, reagents, diluents and QC samples required for each. If circumstances prevent testing a QC Test Profile, patient and blood component samples will not be tested on the analyzer.
 - **DAILY:** The following Test Profiles are performed daily on each analyzer:

- Type and Screen
- Donor Rh Pos
- Donor Rh Neg
- DAT Poly
- DAT IgG
- **DAY OF USE:** Rh Phenotype profile is performed on day of use only. Rh Phenotype card QC is not programmed and must be run before or with patient/donor testing, results printed and attached to QC log
- **NO QC:** The following test profiles do not have a corresponding QC test profile. The reagents used are QC as part of the Type and Screen Profile which must be complete and acceptable prior to use of these test profiles:
 - Blood Type
 - Antibody screen
 - Cord Blood

Table 1: MBC QC Test Profiles, Reagents, QC Samples

QC Test Profile	MTS® Gel Card	Reagent Red Blood Cells	Diluents	QC Sample
Type and Screen	<ul style="list-style-type: none"> • MTS® A/B/D Monoclonal and Reverse Grouping Card • MTS® Anti-IgG Card 	<ul style="list-style-type: none"> • 0.8% AFFIRMAGEN® Reagent Red Blood Cells • 0.8% SURGISCREEN® Reagent Red Blood Cells 	MTS Diluent 2 Plus	<ul style="list-style-type: none"> • AlbaQ-Chek Level1-QC1 • AlbaQ-Chek Level2-QC2 • AlbaQ-Chek Level3-QC3
Donor Rh Pos	<ul style="list-style-type: none"> • MTS® A/B Monoclonal Grouping Card 	N/A	MTS Diluent 2 Plus	<ul style="list-style-type: none"> • AlbaQ-Chek Level1-QC1 • AlbaQ-Chek Level3-QC3
Donor Rh Neg	<ul style="list-style-type: none"> • MTS® A/B/D Monoclonal Grouping Card 	N/A	MTS Diluent 2 Plus	
DAT Poly	<ul style="list-style-type: none"> • MTS® Anti-IgG, -C3d Polyspecific Card 	N/A	MTS Diluent 2	<ul style="list-style-type: none"> • AlbaQ-Chek Level4-QC4 (IgG C₃ NEG) • IgG POS prepared cells • C3 POS prepared cells
DAT IgG	<ul style="list-style-type: none"> • MTS® Anti-IgG Card 	N/A	MTS Diluent 2	<ul style="list-style-type: none"> • AlbaQ-Chek Level4-QC4 (IgG C₃ NEG) • IgG POS prepared cells
Rh Phenotype (performed only on day of use)	<ul style="list-style-type: none"> • MTS® Monoclonal Rh Phenotype Card 	N/A	MTS Diluent 2 Plus	Panel A or B cells with the following specification prepared in Diluent 2 Plus <ul style="list-style-type: none"> • R₁R₁ • R₂R₂ • r'r • r''r

- Resources are stored and used according to manufacture requirements and current Good Manufacturing Practices.
 - Storage and onboard stability requirements are found in [Table 2: Resource Storage Requirements and Onboard Stability](#).
 - General specifications for use are listed in [Table 3: Use and Handling of Resources](#)

Table 2: Resource Storage Requirements and Onboard Stability

Materials	Storage Requirements	Use Requirements	On-Board Stability	Comments
MTS [®] A/B/D Monoclonal and Reverse Grouping Card	2-25°C	Room temperature	Refer to manufacture expiration date on card	<ul style="list-style-type: none"> • Store in an upright position
MTS [®] A/B/D Monoclonal Grouping Card				
MTS [®] A/B Monoclonal Grouping Card				
MTS [®] Anti-IgG,-C3d Card				
MTS [®] Anti-IgG Card				
MTS [®] Monoclonal Rh Phenotype Card				
MTS [®] Diluent 2 Plus	2-8°C	Room temperature	24 hours	<ul style="list-style-type: none"> • Diluents should not be left on the analyzer longer than 24 hours. • Change diluent bottles prior to performing daily QC
MTS [®] Diluent 2				
0.8% AFFIRMAGEN [®] Reagent Red Blood Cells	2-8°C	Room temperature	5 days (120 hours)	<ul style="list-style-type: none"> • Reagent red cells should not be left on the instrument longer than 5 days. Ensure reagent red cells are properly re-suspended prior to use. • Return reagents to refrigerated storage when not in use.
0.8% SURGISCREEN [®] Reagent Red Blood Cells				
IgG Coated Cells	2-8°C	Room temperature	<ul style="list-style-type: none"> • Manufactured at time of quality control testing • Expires 24 hours after manufacture 	<ul style="list-style-type: none"> • Discard after use for quality control testing – May be used for same day automated and manual quality control testing prior to discard
C3 Coated Cells				

Materials	Storage Requirements	Use Requirements	On-Board Stability	Comments
AlbaQ-Chek® QC Kit:	2-8°C	Room temperature	<ul style="list-style-type: none"> Used only during quality control testing Stored in the reagent refrigerator Vials expire 7 days from opening 	Kit contains the following: <ul style="list-style-type: none"> Vial 1 (QC1) – Group A RhD Negative (rr) containing Anti-B, Anti-D Vial 2 (QC2) – Group O RhD Positive (R₁R₁) containing Anti-A, Anti-B, Anti-c Vial 3 (QC3) – Group B RhD Positive (R₁r) containing Anti-A Vial 4 (QC4) – Group A₂B RhD Positive

Table 3: Use and Handling of Resources

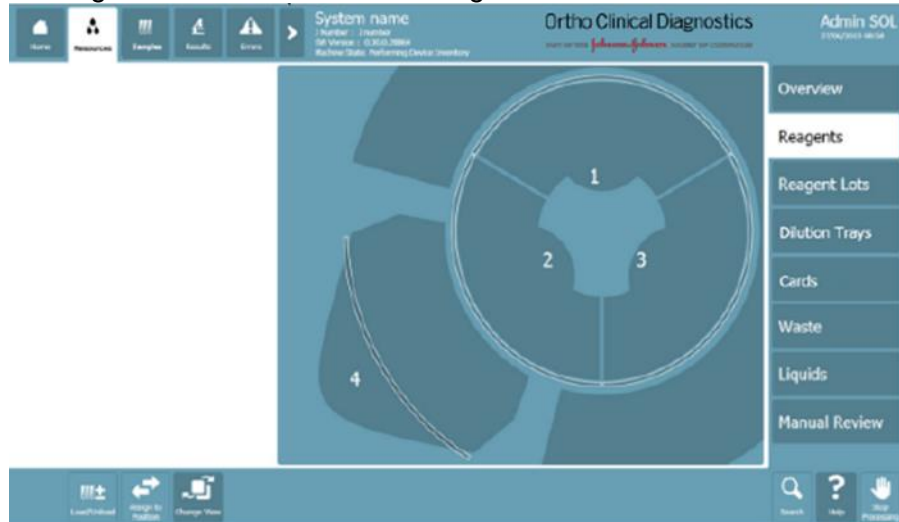
Resource	Specification for Use
Red Blood Cell Reagents, QC Samples, and Diluents	<ul style="list-style-type: none"> Do not use any reagents including quality control samples if evidence of contamination is present.
	<ul style="list-style-type: none"> Do not use if extreme turbidity, precipitation, or hemolysis is present
	<ul style="list-style-type: none"> All reagent red blood cells and diluents must be at room temperature when loaded on the system
	<ul style="list-style-type: none"> Prior to loading on the Vision, reagent red blood cells must be gently inverted to mix the red blood cells until they are completely suspended in the diluent: <ul style="list-style-type: none"> Reagents should not be agitated in a manner that will not cause bubbles in the fluids Remove any bubbles from reagent using an applicator stick or a transfer pipette before loading on the system Care should be used to maintain the concentration and integrity of all reagents
	<ul style="list-style-type: none"> If there is suspicion of testing performed with reagent red blood cells that were not completely suspended: <ul style="list-style-type: none"> Discard the entire set of reagent red cells (ie: A1 and B cells, all 3 screen cells) and Repeat testing with a new set of reagent red cells.
	<ul style="list-style-type: none"> Replace reagent caps on 10mL reagent red blood cell vials with disposable ORTHO VISION® Evaporation Cap prior to loading on the system to prevent evaporation of liquid. In extreme laboratory conditions, such as 15% relative humidity and a temperature of 30° C, excessive evaporation of Ortho Reagent Red Blood Cells reagent may be observed and result in analyzer error. <ul style="list-style-type: none"> ORTHO VISION® Evaporation Cap are intended to be single use

Resource	Specification for Use
Red Blood Cell Reagents, QC Samples, and Diluents	<ul style="list-style-type: none"> and only used on reagent red blood cell vials <ul style="list-style-type: none"> ○ Do not remove ORTHO VISION[®] Evaporation Cap from one vial and place on another vial • Discard used ORTHO VISION[®] Evaporation Cap when the vial is discarded
	<ul style="list-style-type: none"> • Freshly opened 0.8% Affirmagen[®], and 0.8% Surgicreen[®] is validated, by the manufacturer, for 5 days (120 hours) on-board stability when ORTHO VISION[®] Evaporation Cap are used
	<ul style="list-style-type: none"> • Do not store reagent red cells that require agitation on-board the system when the system is in Maintenance Mode or if the system is going to be powered off. <ul style="list-style-type: none"> ○ Reagent red cells requiring agitation will be marked unusable if left on-board when the system is in Maintenance Mode or powered off.
	<ul style="list-style-type: none"> • Ortho Reagent Red Blood Cells 0.8% Affirmagen[®], 0.8% Surgicreen[®], Resolve[®] Panels A, B, C Untreated, and C Ficin should be capped and stored at 2 to 8°C when not in use
	<ul style="list-style-type: none"> • Freshly opened MTS[®] Diluent and MTS[®] Diluent 2 PLUS may be kept on the analyzer for up to 24 hours of continuous use. <ul style="list-style-type: none"> ○ Performance of MTS[®] Diluent 2 PLUS after 24 hours of continuous use on the analyzer has not been validated.
Dilution Trays	<ul style="list-style-type: none"> • Dilutions trays may be used only once. Once all wells are used, the tray should be discarded in a red biohazard container.
MTS[®] Gel Cards	<ul style="list-style-type: none"> • Do not load cards that appear dried out.
	<ul style="list-style-type: none"> • Make sure the foil is sealed properly over all columns.
	<ul style="list-style-type: none"> • Do not manually punch the foil. The system will automatically punch the foil over the correct columns before running the test.
	<ul style="list-style-type: none"> • Before loading cards to the system, make sure the cards do not contain any liquid on the foil. <ul style="list-style-type: none"> ○ Discard cards with liquid on the foil. Cards with liquid on the foil may cause contamination with the gripper or punch tool.
	<ul style="list-style-type: none"> • When loading a sleeve with more than one lot of the same type of gel card, leave an empty slot between lots.
	<ul style="list-style-type: none"> • Only load the same type of card in a single sleeve. The system assumes resources in groups separated by an empty slot are all the same type of test.
	<ul style="list-style-type: none"> • Sleeves must be stored upright with the foil facing up at all times. • Partially used cards are placed in the room temperature incubator by the system and may remain there for up to 4 hour. <ul style="list-style-type: none"> ○ After 4 hours or if there is no available space in the room temperature incubator, the system will discard the card in the waste drawer ○ Do not manually add partially used cards to the system. Partially used cards, may be used for manual gel testing or quality control.
Liquids	<ul style="list-style-type: none"> • Only fill containers with liquids at room temperature.

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Resource	Specification for Use
	<ul style="list-style-type: none"> Liquids should not be agitated in a manner that could cause bubbles in the liquid.

- Resources are tracked, monitored, discarded, and replenished using the Vision Resource Screen and the **Ortho Vision Daily Resource Tracking** form
- On-Board management of resources is managed via the **Resources Screen**



- Use touch buttons located on the right side of the screen to display information about each resource – refer to **Appendix 1: Resource Screen Access and Use**
- Use touch buttons located in the horizontal bar at the bottom of each screen to act or prompt on the data displayed on the screen – refer to **Appendix 2: Resource Action Buttons**
- Ortho Vision Daily Resource Tracking** form is used to track lot number and expiration of reagents that are not captured and monitored by the analyzer.

**SPECIMEN REQUIREMENTS:
N/A**

REAGENTS/SUPPLIES/EQUIPMENT:

Reagents:	Supplies:	Equipment:
<ul style="list-style-type: none"> AlbaQ-Check™ Simulated Whole Blood Controls IgG Sensitized Red Blood Cells ALBAcyte® C3 Coated Red Blood Cells 0.8% AFFIRMAGEN® Reagent Red Blood Cells 0.8% SURGISCREEN® Reagent Red Blood Cells ID-MTS® Diluent 2 ID-MTS® Diluent 2 Plus ID-MTS® Gel Cards 	<ul style="list-style-type: none"> 10 x 75 glass tubes 12 x 75 glass or plastic tubes Evaporation caps Dilution trays Applicator sticks Transfer pipettes Sample racks 	<ul style="list-style-type: none"> Ortho Vision

QUALITY CONTROL: QC is performed daily

INSTRUCTIONS:

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- [Clearing Multiple Profile QC Lines \(Key Operators Only\)](#)
- [Appendix 1: Resource Screen Access and Use](#)
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Printing Barcoded Labels for QC Samples

STEP	ACTION	
1	Log into SmarTerm	
2	Enter the appropriate entry when prompted	
	Prompt	Enter
	Function	BAR
	Which medical center (H or U)?	U
3	Press the down arrow key to move the prompt to “Custom Barcodes” and press <Enter>	
4	Enter the appropriate entry when prompted	
	Prompt	Enter
	Enter device or valid printer #	13
5	Press <Enter> to select the UWMC TSL (686) printer	
6	Enter the text for the appropriate specimen label:	
	If performing QC for MTS® Gel card	Enter the following
	MTS® Anti-IgG,- C3d Polyspecific Card and/or MTS® Anti-IgG Card for DAT IgG	<ul style="list-style-type: none"> • IgG C3 NEG • IgG POS • C3 POS
	MTS® Monoclonal Rh Phenotype Card	<ul style="list-style-type: none"> • R₁R₁ • R₂R₂ • r’r • r”r

STEP	ACTION										
7	Press <Enter> 3 times										
8	Enter the appropriate entry when prompted										
	<table border="1"> <thead> <tr> <th>Prompt</th> <th>Enter</th> </tr> </thead> <tbody> <tr> <td>Do you want sequential labels</td> <td>N</td> </tr> <tr> <td>Which medical center (H or U)?</td> <td>U</td> </tr> <tr> <td>Enter number of labels to print</td> <td>1</td> </tr> <tr> <td>Is this correct</td> <td>Y</td> </tr> </tbody> </table>	Prompt	Enter	Do you want sequential labels	N	Which medical center (H or U)?	U	Enter number of labels to print	1	Is this correct	Y
	Prompt	Enter									
	Do you want sequential labels	N									
	Which medical center (H or U)?	U									
Enter number of labels to print	1										
Is this correct	Y										
9	Repeat steps 2 thru 8 to print additional QC sample labels										
10	Affix the label to the appropriate QC sample container (tube) in a manner allowing scanning by the Vision										


Preparing 0.8% IgG and C3 Positive Controls Samples

STEP	ACTION						
1	Dispense 2 mL of MTS Diluent 2 into a clean test tube NOTE: The 2 mL sample will be used in steps 4 & 8 below						
2	Label 10 x 75 mL test tube with the appropriate label in a manner that allows the barcode to be read by the analyzer – refer to section Printing Barcoded Labels for QC Samples						
	<table border="1"> <thead> <tr> <th>If preparing</th> <th>Then label tube with</th> </tr> </thead> <tbody> <tr> <td>IgG positive red cell suspension</td> <td>IgG POS label</td> </tr> <tr> <td>C3 positive red cell suspension</td> <td>C3 POS label</td> </tr> </tbody> </table>	If preparing	Then label tube with	IgG positive red cell suspension	IgG POS label	C3 positive red cell suspension	C3 POS label
	If preparing	Then label tube with					
IgG positive red cell suspension	IgG POS label						
C3 positive red cell suspension	C3 POS label						
3	Add 200µL of sensitized cells to the labeled tube <table border="1"> <thead> <tr> <th>If preparing</th> <th>Add</th> </tr> </thead> <tbody> <tr> <td>IgG positive red cell suspension</td> <td>IgG sensitized cells</td> </tr> <tr> <td>C3 positive red cell suspension</td> <td>C3 sensitized cells</td> </tr> </tbody> </table>	If preparing	Add	IgG positive red cell suspension	IgG sensitized cells	C3 positive red cell suspension	C3 sensitized cells
If preparing	Add						
IgG positive red cell suspension	IgG sensitized cells						
C3 positive red cell suspension	C3 sensitized cells						
4	Add 100µL of MTS Diluent 2						
5	Mix gently to resuspend						
6	Centrifuge at wash setting						
7	Decant the supernatant						
8	Add 400µL of MTS™ Diluent 2 to each tube						

STEP	ACTION
9	Gently mix to re-suspend avoiding bubbles or foaming
10	Record the following on the label: <ul style="list-style-type: none"> • Date prepared • Time prepared • Tech ID

Loading Controls Samples

STEP	ACTION						
1	<p>Record the lot number and expiration date for the following on the <i>Ortho Vision Daily Resource Tracking</i> form:</p> <ul style="list-style-type: none"> • AlbaQ-Chek Control samples • IgG Check Cells • C3 Check Cells • 0.8% Affirmagen A1 & B • 0.8% Surgiscreen 1, 2, 3 • R1R1, R2R2, r'r, and r'r if performing QC of the MTS[®] Monoclonal Rh Phenotype Card 						
2	<table border="1"> <thead> <tr> <th>If Vials</th> <th>Then</th> </tr> </thead> <tbody> <tr> <td>Were previously opened</td> <td>Go to next step</td> </tr> <tr> <td>Are from a new AlbaQ-Chek kit</td> <td> <ul style="list-style-type: none"> • Place a IgG C3 NEG barcoded label on Vial 4 in a manner that allows the barcode to be read by the analyzer - refer to section Printing Barcoded Labels for QC Samples • Record the following on each vial <ul style="list-style-type: none"> ○ Open date ○ Expiration date: Samples expire within 7 days or the original expiration date, whichever is shorter ○ Tech ID </td> </tr> </tbody> </table>	If Vials	Then	Were previously opened	Go to next step	Are from a new AlbaQ-Chek kit	<ul style="list-style-type: none"> • Place a IgG C3 NEG barcoded label on Vial 4 in a manner that allows the barcode to be read by the analyzer - refer to section Printing Barcoded Labels for QC Samples • Record the following on each vial <ul style="list-style-type: none"> ○ Open date ○ Expiration date: Samples expire within 7 days or the original expiration date, whichever is shorter ○ Tech ID
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3	<table border="1"> <thead> <tr> <th>If</th> <th>Then</th> </tr> </thead> <tbody> <tr> <td>Vials were previously centrifuged and red cells and plasma is separated</td> <td>Go to next step</td> </tr> <tr> <td>Opening a new kit or Red cells and plasma are not separated</td> <td>Centrifuge all 4 vials in the kit to separate the red blood cells and plasma in the same manner as patient samples</td> </tr> </tbody> </table>	If	Then	Vials were previously centrifuged and red cells and plasma is separated	Go to next step	Opening a new kit or Red cells and plasma are not separated	Centrifuge all 4 vials in the kit to separate the red blood cells and plasma in the same manner as patient samples
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Vials were previously centrifuged and red cells and plasma is separated	Go to next step						
Opening a new kit or Red cells and plasma are not separated	Centrifuge all 4 vials in the kit to separate the red blood cells and plasma in the same manner as patient samples						
4	Gently mix IgG POS and C3 POS control samples to resuspend cells						

STEP	ACTION								
5	Ensure contents of all control samples have warmed to room temperature prior to loading on the analyzer								
6	Remove caps from control samples NOTE: Cap colors on AlbaQ-Chek samples match the number color on each vial								
7	<table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <td colspan="2" style="padding: 5px;">Load the QC samples into the appropriate rack</td> </tr> <tr style="background-color: #cccccc;"> <th style="width: 30%;">Control Sample</th> <th style="width: 70%;">Load into</th> </tr> <tr> <td style="padding: 5px;">AlbaQ-Chek sample</td> <td style="padding: 5px;"> <ul style="list-style-type: none"> Blue rack, S13B with barcodes facing out </td> </tr> <tr> <td style="padding: 5px;">IgG POS and C3 POS</td> <td style="padding: 5px;"> <ul style="list-style-type: none"> Red rack, S10B with barcodes facing out Leave about half an inch between the tubes and the bottom of the rack, so that the tubes do not rest all the way down. <p>NOTE: This ensures the probe is able to reach the small volume of sample</p> </td> </tr> </table> 	Load the QC samples into the appropriate rack		Control Sample	Load into	AlbaQ-Chek sample	<ul style="list-style-type: none"> Blue rack, S13B with barcodes facing out 	IgG POS and C3 POS	<ul style="list-style-type: none"> Red rack, S10B with barcodes facing out Leave about half an inch between the tubes and the bottom of the rack, so that the tubes do not rest all the way down. <p>NOTE: This ensures the probe is able to reach the small volume of sample</p>
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Running Quality Control

STEP	ACTION								
1	<table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <th style="width: 50%;">If performing QC on</th> <th style="width: 50%;">Then</th> </tr> <tr> <td style="padding: 5px;">New lot number for any reagent</td> <td style="padding: 5px;">Go to section Programming New QC Sample Lot Numbers</td> </tr> <tr> <td style="padding: 5px;">Previously QC'd lot number</td> <td style="padding: 5px;">Go to next step</td> </tr> </table>	If performing QC on	Then	New lot number for any reagent	Go to section Programming New QC Sample Lot Numbers	Previously QC'd lot number	Go to next step		
If performing QC on	Then								
New lot number for any reagent	Go to section Programming New QC Sample Lot Numbers								
Previously QC'd lot number	Go to next step								
2	<table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <td colspan="2" style="padding: 5px;">Load the following reagents into the appropriate racks</td> </tr> <tr style="background-color: #cccccc;"> <th style="width: 50%;">Reagent</th> <th style="width: 50%;">Rack</th> </tr> <tr> <td style="padding: 5px;"> <ul style="list-style-type: none"> MTS Diluent 2 Plus MTS Diluent 2 </td> <td style="padding: 5px;">Diluent Rack- N02B</td> </tr> <tr> <td style="padding: 5px;"> <ul style="list-style-type: none"> 0.8% AFFIRMAGEN® Reagent Red Blood Cells 0.8% SURGISCREEN® Reagent Red Blood Cells </td> <td style="padding: 5px;">Red Cell Reagent Rack-R10B</td> </tr> </table> <p>NOTE: All reagents must be at room temperature prior to testing</p>	Load the following reagents into the appropriate racks		Reagent	Rack	<ul style="list-style-type: none"> MTS Diluent 2 Plus MTS Diluent 2 	Diluent Rack- N02B	<ul style="list-style-type: none"> 0.8% AFFIRMAGEN® Reagent Red Blood Cells 0.8% SURGISCREEN® Reagent Red Blood Cells 	Red Cell Reagent Rack-R10B
Load the following reagents into the appropriate racks									
Reagent	Rack								
<ul style="list-style-type: none"> MTS Diluent 2 Plus MTS Diluent 2 	Diluent Rack- N02B								
<ul style="list-style-type: none"> 0.8% AFFIRMAGEN® Reagent Red Blood Cells 0.8% SURGISCREEN® Reagent Red Blood Cells 	Red Cell Reagent Rack-R10B								
3	Touch <QC> menu button								
4	Select the QC Test Profile to process – refer to Table 1: MBC QC Test Profiles, Reagents, QC Samples								
5	Touch <Run QC Job> action button								

STEP	ACTION
6	Touch <Save>
7	Touch <Start>
8	Repeat steps 3-7 to select additional test profiles
8	Touch <Samples> then select a ring position into which samples will be loaded
9	Touch <Load/Unload> and open the door
10	Select any additional ring positions into which QC samples will be loaded
11	Place rack(s) in the Load Station and close the door NOTE: The system will automatically start running the QC as STAT samples once programmed and loaded it will also auto accept results and archive them.

Programming New QC Control Sample ID or Lot Numbers

STEP	ACTION
1	Verify no samples are running or results waiting to be archived
2	Touch <QC> menu button
3	Select the QC Test Profile for which the QC Sample is associated - refer to Table 1: MBC QC Test Profiles, Reagents, QC Samples
4	Touch <Run QC Job> action button
5	Touch <Change Ortho QC sample ID>
6	Delete the existing number in the sample ID (lot number) Note: To see what QC samples are run with each profile refer to the QC Profiles Table
7	Scan in the new sample ID (lot number) for each sample used for the QC Test Profile – refer to Table 1: MBC QC Test Profiles, Reagents, QC Samples IMPORTANT: Entering the barcode using the handheld barcode scanner will automatically enter the number twice. If manually typing in the lot number, type the barcode twice.
8	Touch <Save>
9	Repeat steps 2-8 to program additional QC Sample ID (lot number)

QC of different ID-MTS Gel Card lots or Reagent Red Cell Lots

STEP	ACTION	
1	Load all resources including all lots requiring QC	
2	Touch <QC> menu button	
3	Touch <Run QC Job> action button	
4	If new lot of	Then
	Red cell reagent	<ul style="list-style-type: none"> • Touch a reagent lot for the reagent type • Touch twice to view all lot numbers that are loaded the analyzer <p>NOTE: If there is more than one reagent lot loaded on the analyzer, the default selection is the lot that was most recently registered.</p>
	ID-MTS Gel Card	<ul style="list-style-type: none"> • Touch a card lot for each required card type • Touch the card type twice to view all lots that are loaded on the analyzer. <p>NOTE: If there is more than one required card lot loaded on the analyzer, the default selection is the lot that was most recently registered.</p>

Managing Failed QC Results

STEP	ACTION	
2	Select the failed QC sample	
	If	Then
	Failure is due to reason that does not affect the result (ie: dust on the card)	<ul style="list-style-type: none"> • Modify the result and accept the results. Refer to SOP <i>Ortho Vision® Results Management</i>
	QC should be repeated	<ul style="list-style-type: none"> • Reject the failed QC sample profile • Go to Step 3 <p>NOTE: If only one sample of a QC Test Profile fails, only the sample that failed needs to be repeated</p>
3	Touch <Run QC Job>	
	NOTE: The failed QC run will remain on the MBC QC Profile list as failed	
4	Touch <Save>	
5	Touch <Start>	

Printing QC Result Reports

STEP	ACTION
1	Touch <QC> menu button
2	Touch the MBC Test Profile Name NOTE: Each profile needs to be individually printed
3	Touch <Show QC History Report>
4	Touch <Presets>
5	Touch <Today> and <OK> to display the report
6	Touch <Print>
7	Review the report to verify QC is acceptable
8	Record the following at the bottom of the report: <ul style="list-style-type: none"> • Date of Review • Open date and time of red blood cell reagent vials • Tech ID
9	Repeat steps 1 thru 8 for additional Test Profile QC results

Running Rh phenotype Card QC

STEP	ACTION
1	Select QC samples with the following phenotypes from reagent panels cells to run alongside your patient/donor sample: <ul style="list-style-type: none"> • R₁R₁ • R₂R₂ • r'r • r''r
2	Remove Rh Phenotype cards from refrigerator and allow to come to room temperature
3	Wipe with a Kimwipe to remove any dust
4	Label 10x75mm tubes with phenotype selected EXAMPLE: R ₁ R ₁
5	Aliquot 7- 10 drops of corresponding cells into labeled tubes
6	Load Tubes into a red S10B sample rack and load into the analyzer
7	Assign to a ring position after the system alerts of a unreadable barcode
8	Close load station door

STEP	ACTION
9	Touch <Sample> and <Create Order>
10	Touch <1 st sample liquid type>
11	Touch <Assigned Profiles> and <Rh Phenotype>
12	Touch <Save> and <Start>
13	Go to section Printing QC Result Reports when testing is complete

Clearing Multiple Profile QC Lines (Key Operators Only)

STEP	ACTION
1	Verify no samples are running and that there are no results waiting to be archived NOTE: Profiles cannot be cleared while samples are running or results are pending
2	Touch <Stop Processing> twice
3	Touch <Setup> and <Testing>
4	<ul style="list-style-type: none"> • Touch <Grouping Tests into profiles> tab • Touch <Show Details> <p>NOTE: A list of all the testing profiles is displayed; only the tests listed as MBC (Method Based Control) need to be reprogrammed with each new lot.</p>
5	Touch the desired profile (it should be highlighted in white)
6	Touch <Show QC Samples>
7	<ul style="list-style-type: none"> • Touch the QC sample to delete • Touch <Delete Selected> • Repeat until all necessary QC Samples are deleted
8	<ul style="list-style-type: none"> • Touch <Create QC Sample>
9	<ul style="list-style-type: none"> • Touch QC sample type and select either: <ul style="list-style-type: none"> ○ Ortho QC Sample <ul style="list-style-type: none"> ▪ Type and Screen Test Profile ▪ Donor Rh Pos Test Profile ▪ Donor Rh Neg Test Profile ○ User defined QC sample <ul style="list-style-type: none"> ▪ DAT Poly Test Profile ▪ DAT IgGTest Profile

STEP	ACTION																								
10	If using	Then																							
	Ortho QC sample	Touch <Ortho QC Sample ID> and scan barcode, sample information will auto populate <table border="1" data-bbox="526 415 1437 625"> <thead> <tr> <th data-bbox="526 415 784 485">Test</th> <th data-bbox="784 415 1003 485">Sample Type</th> <th data-bbox="1003 415 1222 485">Liquid Type</th> <th data-bbox="1222 415 1437 485">Expected Results</th> </tr> </thead> <tbody> <tr> <td data-bbox="526 485 784 527">Type and Screen</td> <td data-bbox="784 485 1003 625" rowspan="3">Ortho QC Sample</td> <td data-bbox="1003 485 1222 625" rowspan="3">AlbaQ-Chek</td> <td data-bbox="1222 485 1437 625" rowspan="3">Will auto populate</td> </tr> <tr> <td data-bbox="526 527 784 569">Donor Rh Pos</td> </tr> <tr> <td data-bbox="526 569 784 625">Donor Rh Neg</td> </tr> </tbody> </table>			Test	Sample Type	Liquid Type	Expected Results	Type and Screen	Ortho QC Sample	AlbaQ-Chek	Will auto populate	Donor Rh Pos	Donor Rh Neg											
Test	Sample Type	Liquid Type	Expected Results																						
Type and Screen	Ortho QC Sample	AlbaQ-Chek	Will auto populate																						
Donor Rh Pos																									
Donor Rh Neg																									
	User defined QC sample	<ul style="list-style-type: none"> • Touch <1st sample ID> and scan barcode • Touch <1st Sample Liquid Type> and select according to the liquid type listed below • Touch <Expected Results> and select result according expected results listed below <table border="1" data-bbox="526 800 1424 1142"> <thead> <tr> <th data-bbox="526 800 824 842">DAT Poly Test</th> <th data-bbox="824 800 1123 842">Liquid Type</th> <th data-bbox="1123 800 1424 842">Expected Results</th> </tr> </thead> <tbody> <tr> <td data-bbox="526 842 824 894">IgG C3 NEG</td> <td data-bbox="824 842 1123 894">CentBlood</td> <td data-bbox="1123 842 1424 894">NEG</td> </tr> <tr> <td data-bbox="526 894 824 947">IgG POS</td> <td data-bbox="824 894 1123 947">0.8% cells</td> <td data-bbox="1123 894 1424 947">POS</td> </tr> <tr> <td data-bbox="526 947 824 999">C3 POS</td> <td data-bbox="824 947 1123 999">0.8% cells</td> <td data-bbox="1123 947 1424 999">POS</td> </tr> <tr> <th data-bbox="526 999 824 1041">DAT IgG Test</th> <th data-bbox="824 999 1123 1041">Liquid Type</th> <th data-bbox="1123 999 1424 1041">Expected Results</th> </tr> <tr> <td data-bbox="526 1041 824 1094">IgG C3 NEG</td> <td data-bbox="824 1041 1123 1094">CentBlood</td> <td data-bbox="1123 1041 1424 1094">NEG</td> </tr> <tr> <td data-bbox="526 1094 824 1142">IgG POS</td> <td data-bbox="824 1094 1123 1142">0.8% cells</td> <td data-bbox="1123 1094 1424 1142">POS</td> </tr> </tbody> </table>			DAT Poly Test	Liquid Type	Expected Results	IgG C3 NEG	CentBlood	NEG	IgG POS	0.8% cells	POS	C3 POS	0.8% cells	POS	DAT IgG Test	Liquid Type	Expected Results	IgG C3 NEG	CentBlood	NEG	IgG POS	0.8% cells	POS
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11	For Ortho Defined QC Sample Note: For expected results for each AlbaQ-Chek samples go to: Result Interpretation in tests with AlbaQ-Chek and Routine Blood Bank Reagents																								
12	Follow these steps to clear each old lot Profile QC line when a new lot of cards/reagents/diluents or other is introduced to the system																								

CALCULATIONS/INTERPRETATIONS/RESULTS REPORTING/NORMAL VALUES/CRITICAL VALUES

EXPECTED RESULTS:

Type and Screen Test Profile

MTS™ A/B/D Monoclonal and Reverse Grouping Card								MTS™ Anti-IgG Card			
ABO/Rh		Anti-A	Anti-B	Anti-D	Ctrl	A₁ Cells	B Cells	Antibody Detection	SC 1	SC 2	SC 3
QC1	Expected Results	3-4	0	0	0	0	3-4	Expected Results	3-4	3-4	0
QC2	Expected Results	0	0	3-4	0	3-4	3-4	Expected Results	0	3-4	3-4
QC3	Expected Results	0	3-4	3-4	0	3-4	0	Expected Results	0	0	0
QC4*	Expected Results	3-4	3-4	3-4	0	0	0	Expected Results	0	0	0

* QC4 is not routinely used for QC of the Type and Screen Test Profile.

Donor Rh Neg Test Profile

MTS™ A/B/D Monoclonal Grouping Card				
ABORh		Anti-A	Anti-B	Anti-D
QC1	Expected Results	3-4	0	0
QC3	Expected Results	0	3-4	3-4

Donor Rh Pos Test Profile

MTS™ A/B Monoclonal Grouping Card			
ABO		Anti-A	Anti-B
QC1	Expected Results	3-4	0
QC3	Expected Results	0	3-4

DAT Poly Test Profile

MTS™ IgG,-C3d Polyspecific Card		
Poly DAT		IgG,-C3d
IgG POS	Expected Results	3-4
C3 POS	Expected Results	3-4
QC4	Expected Results	0

DAT IgG Test Profile

MTS [™] IgG Card		
IgG DAT		IgG
IgG POS	Expected Results	3-4
QC4	Expected Results	0

Rh Phenotype Test Profile

MTS [™] Monoclonal Rh Phenotype Card							
Rh Phenotype		Anti-D	Anti-C	Anti-E	Anti-c	Anti-e	Ctl
R ₁ R ₁	Expected Results	+	+	-	-	+	-
R ₂ R ₂	Expected Results	+	-	+	+	-	-
r'r	Expected Results	-	+	-	+	+	-
r''r	Expected Results	-	-	+	+	+	-

CALIBRATION:

NA

PROCEDURE NOTES AND LIMITATIONS:

- Biohazard material is disposed of according to local regulations and University of Washington Medical Center guidelines
- The user is responsible for monitoring the length of time the reagents have been on board the analyzer – refer to **Ortho Vision Daily Resource Tracking** form
- When using automated instruments, refer to the limitations contained in the operator's manual provided by the device manufacturer. Strict adherence to the procedures and recommended equipment is essential.
- Proper centrifuge calibration is particularly important to the performance of the ID-MTS[™] Gel Test. The ORTHO[™] Workstation and ORTHO VISION[™] Analyzer have been exclusively designed to provide the correct time, speed and angle.
- Hemolyzed and grossly icteric blood samples may cause difficulty in interpretation, and test results should be used with caution. Occasionally, specimens showing incomplete clotting or excess particulates may need to be washed prior to testing
- Make sure to select the proper sample type before proceeding. Selecting an incorrect sample type may cause incorrect results.
- When a 2nd MLS is not available to perform the review and patient or unit test results must be reported, the MLS tech will independently review the QC results to verify acceptability of result prior to reporting patient testing
- False positive or false negative test results may occur from bacterial or chemical contamination of test materials, aged or hemolyzed blood specimens, inadequate incubation

time or temperature, improper centrifugation, improper storage of materials, or omission of test samples.

- Antibodies to preservatives, medications, disease states, Wharton's jelly, and/or cross-contamination of reaction microtubes may cause false positive reactions.
- False positive results may occur if a card that shows signs of drying is used in testing.
- Red blood cells must be diluted to 4% ± 1% in MTS™ Diluent 2 *PLUS* before addition to the microtubes of MTS Gel cards for AB, ABD or ABO/Rh testing.
- Red blood cells must be suspended in MTS™ Diluent 2 or be a commercial 0.8% red blood cell in a low ionic strength diluent specifically approved for use in the ID-Micro Typing System™ to be used with MTS IgG or MTS Anti-IgG, -C3d gel cards.
- Some weak subgroups of the A and B antigen may not be detected by these MTS™ Anti-A and Anti-B reagents. The use of the MTS™ Monoclonal Anti-A,B Card may better detect these weak antigens.
- Variations in red blood cell concentration can markedly affect the sensitivity of test results. If red blood cell suspensions are too concentrated, they can give weaker results due to the increase in the antigen/antibody ratio. In addition, cells may fail to completely migrate to the bottom of the microtube and could cause a false positive interpretation. When red blood cells are too low in concentration, they become difficult to visualize and, in extreme cases, a weak positive can fail to be detected.
- Rouleaux caused by serum or plasma with abnormally high concentrations of protein (such as in patients with multiple myeloma or Waldenstrom's macroglobulinemia or from patients who have received plasma expanders of high molecular weight) may infrequently cause difficulties in ID-MTS™ Gel Test interpretation, false positive results or hazy reactions may occur with these samples but are rare. If false positive reactions (e.g., rouleaux, cells coated with immunoglobulins, etc.) occur in the control gel, the blood group cannot be established. Additional testing will be necessary to resolve this false positive reaction. Refer to SOPs- *ABO/Rh Discrepancy* and *Saline Replacement*.
- Suppressed or diminished expression of certain blood group antigens may give rise to false negative reactions. For this reason, caution should always be exercised when assigning the ABO phenotype. The results of forward grouping (red blood cell) testing should be confirmed by reverse grouping (serum) testing.
- In some patients (e.g., newborns, elderly or immunocompromised patients) the expected ABO antibodies may be weak or missing. For any recipient whose ABO group cannot be accurately determined, group O red blood cells should be considered as a transfusion alternative. The interpretation of reactions obtained when testing infant blood may be complicated by the fact that the infant's serum does not necessarily contain antibody for any antigen absent from the cells, and passive anti-A and/or Anti-B from the mother's circulation may yield conflicting reactions when tests are performed on cord blood specimens. Cord blood specimens may also give weaker than normal reactions in the cell grouping test. Imperfect development of the ABH antigens at birth may lead to false negative results, particularly with Anti-A reagents.
- When using automated instruments, refer to the limitations contained in the operator's manual provided by the device manufacturer
- Anomalous results may be caused by fibrin or other particulate matter in blood samples that could stick to the sides of the microtube during ABO/Rh testing. .
- Anomalous results may be caused by fresh serum, fibrin, or particulate matter in serum or plasma, or red blood cells that stick to the sides of the microtube of MTS IgG or MTS Anti-IgG, -C3d gel cards. Anomalous results (i.e., a line of red blood cells on the top of the gel) may be observed with serum samples and can be minimized with the use of EDTA plasma

- The MTS™ Anti-IgG Card is not manufactured to detect Anti-C3 cell sensitizations. It may be used in the compatibility test; however, some literature reports indicate that the Anti-IgG may occasionally fail to detect antibodies that are demonstrable by the use of antiglobulin reagents containing Anti-C3.
- There is the potential for IgM antibodies to react to anti-IgG testing. Some patient antibodies that are IgM in nature may react with corresponding antigens in the upper portion of the microtube and be trapped in the top portion of the gel at the time of centrifugation resulting in a positive reaction.
- Negative direct antiglobulin test results do not necessarily rule out hemolytic disease of the newborn (HDN), especially if ABO incompatibility is suspected.

REFERENCES:

- ORTHO VISION® Analyzer Procedure Pub. No. J56102
- ORTHO VISION® Analyzer General Operator Training Manual
- AlbaQ-Chek™ Simulated Whole Blood Controls Package Insert
- Antibody Identification Method (with Auto Control) Using MTS™ Anti-IgG Cards

RELATED DOCUMENTS:

FORM Ortho Vision Daily Resource Tracking














APPENDICES:

Appendix 1: Resource Screen Access and Use

Resource	To access	Use
Reagents	Reagents Screen <ul style="list-style-type: none"> • Touch <Resources> • Touch <Reagents> 	To evaluate current inventory of reagents loaded on the system and manage reagent lots
	Load/Unload button <ul style="list-style-type: none"> • Touch <Show Details> while in Table View 	Loading and unloading reagents as needed
Reagents Lots	Reagent Lot information <ul style="list-style-type: none"> • Touch <Resources> • Touch <Reagent Lots> 	To register lots and view information about reagent kits. Information displayed may include <ul style="list-style-type: none"> • Lot # • Expiration Date • Onboard reagents • QC status if applicable • Reagent Kit – The name of the reagent kit or reagent family associated with the lot
Dilution Trays	Dilution Trays screen <ul style="list-style-type: none"> • Touch <Resources> • Touch <Dilution trays> 	To view information about the availability and position of dilution wells
Cards	Cards screen <ul style="list-style-type: none"> • Touch <Resources> • Touch <Cards> 	To view: <ul style="list-style-type: none"> • Card carton position and other associated information, such as location and lot expiration dates • Card type, errors and warnings
Waste	Waste screen <ul style="list-style-type: none"> • Touch <Resources> • Touch <Waste> 	To view the current status of the waste drawer, card waste drawer and liquid waste bottle. Information displayed includes: <ul style="list-style-type: none"> • Total Capacity – Available fill level • Current Capacity – Free fill level • Time Until Full – An expected fill level and estimated time until the fill level is reached.
Liquids	Liquid screen <ul style="list-style-type: none"> • Touch <Resources> • Touch <Liquids> 	To monitor the availability of deionized water and saline on the system.

Resource	To access	Use
Manual Load/Review	Manual Load/Review <ul style="list-style-type: none"> • Touch <Resources> • Touch < Manual Load/Review 	To view the status of tests on the system requiring manual review. NOTE: This screen only provides access to the manual review function. It does not allow use of the LOAD AREA in the DUAL PURPOSE DRAWER.

Appendix 2: Resource Action Buttons

Action Button	Name	Description
	Assign to Position	Displays the Assign to Position wizard.
	Change View	Switches the display between a diagram view and a table view.
	Show Details	Displays additional information for the selected item.
	Empty Liquid	Starts a wizard which guides you through the process of emptying the liquids container.
	Empty Cards	Starts a wizard which guides the user through the process of emptying the Cards container.
	Load/Unload	Starts a wizard which guides the user through the process of loading/unloading reagents.
	Pause Auto Refresh	Pauses the automatic update of table information and changes the text of the button to Start Auto Refresh. Touch the Start Auto Refresh button to resume the automatic update.
	Refill	Starts a wizard which guides the user through the process of refilling the Deionized Water and Saline.
	Register OCD Lot	Starts a wizard which guides the user through the process of adding an OCD Lot.
	Register User Defined	Starts a wizard which guides the user through the process of adding a User Defined Lot.
	Show Inventory Report	Displays information about the quantity of consumables on-board the system.
	Show Lot Switch Log	Displays information about the product name, Lot ID, and first use.
	Show Usage Statistics	Displays information about used/unused consumables. Number of orders failed/finished. Number of tests started/reportable/finished/failed. Number of test result started/indeterminate/finished/failed

UWMC SOP Approval:

UWMC CLIA

Medical Director

Mark H. Wener, MD

Date

Transfusion

Service Manager

Nina Sen

Date

Compliance

Analyst

Christine Clark

Date

Transfusion

Service

Medical Director

Monica Pagano, MD

Date

UWMC Biennial Review:

Date

Date