**Purpose:**

To describe the process of setting up Blood Bank Instruments for results to be transferred to SQ from the Tango. This also describes the process of LIS transfer utilizing the Blood Bank Instruments application of Sunquest from the Tango instrument to the LIS and provides guidelines for performing a file cleanup in SmarTerm (Function OFC), which clears the cups in the Blood Bank Instruments application.

**Limitations and Precautions:**

* Failure to verify patient identifiers prior to release of test data may result in incorrect reporting of test results.
* Failure to clear the Blood Bank Instrument application may result in failure of results to transfer from the TANGO to Sunquest.
* Applying Function OFC prior to completing LIS transfer will result in the loss of transferred test results.

**Procedure:**

|  |  |  |
| --- | --- | --- |
| **Step** | **Action** | **Related Documents** |
| **Setting up Blood Bank Instruments in Sunquest** |
| 1 | Open Blood Bank Method Configuration |  |
| 2 | Configuration for patient results:* Type PATIENT in configuration field
* Patient/Product: Patient Specimen
* View: Batch
* Cups: Unreviewed Only
* Specimen ID Display: CID Only
* Add HTANGO to Selected Methods
* Add: %ABR and %AS to Selected Test Codes
* Click on Save
 |  |
| 3 | Configuration for donor unit results:* Type UNIT in configuration field
* Patient/Product: Product
* View: Batch
* Cups: Unreviewed only
* Add HTANGO to Selected Methods
* Add: %ARC to Selected Test Code
* Click on Save
 |  |
| **Step** | **Action** | **Related Documents** |
| **Reviewing and Submitting Transferred Results to LIS Tests** |
| 1 | Open Blood Bank Instruments application in Sunquest. |  |
| 2 | Select the configuration desired for test result entry by utilizing the drop down menu located in the ‘Methods Configuration’ box:* Patient Testing
* Unit Testing
 |  |
| 3 | Select the ‘OK’ button to access the BBI Results screen. |  |
| 4 | Test results transmitted from the Tango can be verified and released by Batch Specimen Mode.* The Batch Specimen Mode will allow for a list of specimens to be verified and released one at a time.
 | TANGO Validation and Export of Results |

|  |  |  |
| --- | --- | --- |
| 5 | Verify that the reaction results correlate with the interpretation. | Table ATable BTable C |
| 6 | **No Discrepancies:*** Check mark in the Release column
* Select the ‘Release Batch’ button

**Discrepancies Found:*** Deselect the discrepant results in the Release column
* Checkmark removed
 |  |
| 7 | Once released, a box appears that says, ‘All released instrument data filed successfully.’ Select the ‘OK’ button.Note: Only one patient’s ABO result will be available at a time. If you run both the TSCR sample and the ABRH2 sample, you must validate the first result that crosses close then re-open the BBI screen to result the second |  |

|  |
| --- |
| **Clear Function OFC in SmarTerm** |
| 1 | Ensure that all results transmitted from the TANGO are reviewed and verified prior to performing this function. |  |
| 2 | Go to **SMARTTERM**. |  |
| 3 | At the Function prompt, enter **OFC**. |  |
| 4 | At the Method Code prompt, enter **HTANGO**. |  |
| 5 | Return |  |
| 6 | At the **START AT CUP NUMBER** prompt, return. |  |
| 7 | At the **STOP WITH CUP NUMBER** prompt, return. |  |
| 8 | At the Method Code Prompt, return to Exit. |  |
| 9 | Function OFC is scheduled to be cleared once daily. When workload is high, more frequent clearing will ensure successful transfer of results.  |  |

**Table A: Patient ABO/Rh Result Grid:**

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **A** | **B** | **A,B** | **D1** | **D2** | **NC** | **A1C\*** | **BC\*** | **Blood Type** |
| 0 | 0 | 0 | ≥2 | ≥2 | 0 | ≥2 | ≥2 | O Pos |
| 0 | 0 | 0 | 0 | 0 | 0 | ≥2 | ≥2 | O Neg |
| ≥2 | 0 | ≥2 | ≥2 | ≥2 | 0 | 0 | ≥2 | A Pos |
| ≥2 | 0 | ≥2 | 0 | 0 | 0 | 0 | ≥2 | A Neg |
| 0 | ≥2 | ≥2 | ≥2 | ≥2 | 0 | ≥2 | 0 | B Pos |
| 0 | ≥2 | ≥2 | 0 | 0 | 0 | ≥2 | 0 | B Neg |
| ≥2 | ≥2 | ≥2 | ≥2 | ≥2 | 0 | 0 | 0 | AB Pos |
| ≥2 | ≥2 | ≥2 | 0 | 0 | 0 | 0 | 0 | AB Neg |

**\*** Initial reactivity of 1+ is acceptable for A1 and B cells if discrepancy workup has previously been performed. Subsequent testing matching the 1+ reactivity need not be repeated

**Table B: Patient Solidscreen Result Grid:**

|  |  |  |
| --- | --- | --- |
|  | **AHG** | **Interpretation** |
| **SC1/SC2/SC3** | **≥ +/-** | **Positive** |
| **SC1/SC2/SC3** | **0** | **Negative** |

**Table C: Donor Unit Result Grid:**

**ND = Not Done**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Anti-A** | **Anti-B** | **Anti-D\*** | **Negative control\*** | **Blood Type** |
| 0 | 0 | ND | ND | O Pos |
| 0 | 0 | ≥3 | 0 | O Pos |
| 0 | 0 | 0 | 0 | O Neg |
| ≥3 | 0 | ND | ND | A Pos |
| ≥3 | 0 | ≥3 | 0 | A Pos |
| ≥3 | 0 | 0 | 0 | A Neg |
| 0 | ≥3 | ND | ND | B Pos |
| 0 | ≥3 | ≥3 | 0 | B Pos |
| 0 | ≥3 | 0 | 0 | B Neg |
| ≥3 | ≥3 | ND | ND | AB Pos |
| ≥3 | ≥3 | ≥3 | 0 | AB Pos |
| ≥3 | ≥3 | 0 | 0 | AB Neg |

**\*Rh confirm strip not required for Rh positive units**

**References:**

Blood Bank User Guide, Sunquest Laboratory version 8.1