Department of LABORATORY MEDICINE

University of Washington Medical Center 1959 NE Pacific Street. Seattle, WA 98195 Transfusion Services Laboratory Policies and Procedures Manual

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TITLE: ABO/Rh Discrepancy Resolution

PURPOSE:

To provide guidance for resolving ABO and Rh testing discrepancies. A discrepancy exists when:

- Results of the forward group do not agree with the reverse group
- Unexplained weak or mixed field reactivity
- Current testing is not in agreement with previous records

PRINCIPLE & CLINICAL SIGNIFICANCE:

The test methods employed depend on the type of discrepancy detected. Failure to resolve a blood typing discrepancy accurately can lead to transfusion of incompatible blood components which can result in hemolytic transfusion reactions.

- Discrepancies may arise from patient identification or labeling errors, intrinsic problems with red cell (forward type), serum (back type), patient disease, treatment, or technical errors in test performance.
- Mixed field (MF) reaction and other discrepancies may arise due to transfusion of ABO compatible but non-identical blood components and or allogeneic BMT
- Weak D positive donors considered Rh positive can lead to Rh discrepancies between the patient and transfused circulating red blood cells.

POLICIES:

- Difficulty in typing and resolution must be documented in the LIS as a BBCS comment.
- Mixed field reactions seen with cord blood samples may be due to contamination from maternal blood, Warton's jelly and weak expression of RBC antigens. Follow the instructions in section <u>Cord Blood Mixed Field Reaction</u> for resolution and action to take if not resolved.

SPECIMEN REQUIREMENTS:

EDTA is preferred and if not tested soon after collection, should be stored at 1-6°C. Red top tubes are also acceptable. See SOP *Specimen Acceptability and Order Receipt*

REAGENTS/SUPPLIES/EQUIPMENT:

Refer to the SOP specific for the test being performed.

QUALITY CONTROL:

Refer to the SOP specific for the test being performed.

INSTRUCTIONS:

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All Discrepancies

STEP	ACTION		
	Review the patient's history in the Laboratory Information System (LIS) and/or Electronic Medical Record (eMR) for indications of the cause of the discrepancy NOTE: It may be necessary to contact other facilities where the patient may have been treated to obtain transfusion and/or transplant history.		
	If Then		
	Patient has received a BMT or other type of HSCT	 If the discrepancy is explained by a mixed population of donor and recipient cells and/or antibodies -report as No Type Determined (NTD). Go to section <u>Reporting Results</u> If the discrepancy is explained by a mixed population including the donor RBCs, report as the patient's original blood type Go to section Reporting Results 	
	Patient received out of group blood		
	Current does not match historical	Request a new sample and repeat testing.	
1		If new sample	Then
		Agrees with historical	 Proceed to interpretation and reporting results. Reject first sample as MISLABEL Look for a reciprocal mislabeled specimen Notify a manager or lead
		Disagrees with historical	 Write a QI Examine historical for misinterpretation Ask RN to verify photo-ID Notify department management and
			obtain Manager/Lead approval before issuing any products to patient if current and new samples both disagree with historical type
	Discrepancy not resolved	Go to next step	

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STEP	ACTION		
	Prepare a new 3-4% the manual test met	patient cell suspension and repeat the ABO/Rh testing using hod	
	If discrepancy is	Then	
2	Resolved	Go to section Results Reporting	
	NOT resolved	 Save reaction results, but do not enter an interpretation in Sunquest Go to next step 	
3	 Review patient's transfusion/transplant/age history for cause of the discrepancy. The following sources may be used to obtain relevant patient information: Sunquest BBI Epic Contact the patient, patient's physician or caregiver Contact other medical facilities where the patient was provided care 		
4	 Contact other medical facilities where the patient was provided care Select the type of discrepancy below and follow the suggested technique as appropriate to resolve the discrepancy Unexpected positive reactions in the forward type (patient cells) Weak or unexpected negative reaction in the forward type (patient cells) Unexpected positive reactions in reverse type (patient serum) Weak or unexpected negative reaction in the reverse type (patient serum) Rh Discrepancy Cord Blood Mixed Field Reaction 		

Unexpected positive reactions in the forward type (patient cells)

STEP	ACTION		
Perform an IgG DAT			
	lf	Then	
1	Positive	Treat with EGA and repeat testing with EGA treated cells	
	Negative	Wash patient cells with warm saline 4X and repeat testing	
	If resolved	Then	
2	Yes	Go to section Results Reporting	
	No	Consult with TSL management or submit to IRL	

Weak or unexpected negative reaction in the forward type (patient cells)

STEP	ACTION		
	lf	Then	
	Weak subgroup of A or B or depression	Try the following in order and discontinue additional testing once the discrepancy is resolved:	
1	of antigen expression	 Incubate patient's cells with anti-A, anti-B, 7% albumin and anti-A,B (if applicable) for 15-20 minutes at RT and repeat testing 	
		 Incubate the tubes from repeat ABO/Rh testing (along with 1 drop group O screen cells and two drops of patient's plasma as a control) at 4°C for 10 min 	
	Overset	• Try using a different manufacturer of antisera if available	
	Suspect neutralization of blood group antisera	testing	
	If resolved	Then	
2	Yes	Go to section Results Reporting	
	No	Consult with TSL management or submit to IRL	

Unexpected positive reactions in reverse type (patient serum)

STEP	ACTION		
Perform a microscopic evaluation of the reaction to look for rouleaux		aluation of the reaction to look for rouleaux	
	If rouleaux is	Then	
1	Observed (retractile stacking of red cells resembling a stack of coins)	Perform Saline Replacement and repeat testing	
	Not Observed	Go to next step	
	Perform an antibody scree	en at IS phase	
	If Antibody screen is	Then	
2	Positive	 Suspect possible cold antibody: Perform a cold panel to detect cold-reacting antibodies If identified repeat testing with antigen negative reverse cells Prewarm reverse typing for cold agglutinin 	
	Negative	 Suspect isoantibodies such as anti-A₁ Type patient cells with A₁ lectin and test patient plasma with A₂ cells 	
	If resolved	Then	
3	Yes	Go to section Results Reporting	
	No	Consult with TSL management or submit to IRL	

Weak or unexpected negative reaction in the reverse type (patient serum)

STEP		ACTION
1	 Suspect depressed antibody production and try the following in the order listed and stop once discrepancy is resolved: Incubate patient's plasma with A1 and B cells, screen cells and an auto control for 15-30 minutes at room temperature (RT) Incubate at 4°C for at least 10 minutes Increase the serum to cell ratio by using 3-4 drops of patient's plasma and one drop of reagent A1 and P cells and repeat texting. 	
	If resolved	Then
2	Yes	Go to section Results Reporting
	No	Consult with TSL management or submit to IRL

Rh Discrepancy

STEP		
	Repeat ABO/Rh using alternate method - refer to SOPs ABO/Rh Manual Tube Method or Ortho Vision Patient and Donor Testing and the appropriate package insert	
1	lf	Then
	Resolved	Go to section Results Reporting
	Not Resolved	Go to next step
Perform Weak D testing per SOP Weak D Tube Testing Pro		P Weak D Tube Testing Procedure
	lf	Then
2	Resolved	Report results according to SOP Weak D Tube
		Testing Procedure
	Not Resolved	Consult with TSL management

Cord Blood Mixed Field Reactions

STEP	ACTION		
1	Wash cord blood specimens 3 times with saline		
2	Repeat ABO/Rh front type		
	If mixed field is resolved	Then	
3	Yes	Go to section Results Reporting below	
	No	Go to next step	
	If cord blood ABO/Rh is Then		
4	For maternal Rh immunoglobulin evaluation (mother is Rh negative)	Reject specimenNotify the RN a new specimen is required	

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STEP	ACTION		
	Cancel the BBCORD test in SQ and create a QI		CORD test in SQ and create a
		lf	Then
		First collected specimen	 Reject specimen Notify the RN a new specimen is required Cancel the BBCORD test in SQ and create a QI
		Repeat specimen	Go to section <u>Results</u> <u>Reporting</u> below
	NOT for Maternal Rh immunoglobulin evaluation	Go to section Res	ults Reporting below

CALCULATIONS/INTERPRETATIONS/RESULTS REPORTING/NORMAL VALUES/CRITICAL VALUES

Results Reporting

STEP	ACTION		
	Record all reactions immediately in Sunquest or the appropriate manual result		
	If discrepancy is	Then	
	Resolved	Refer to interpretation and reporting results according to SOP ABO/Rh Testing by Tube Method	
	Due to a bone marrow transplant (BMT) with	Report the ABO/Rh interpretation as NTD	
1	discrepant forward/reverse reactions or in between typing	NOTE: In some cases, the patient may not make the new isoantibody following conversion. Testing may still be reported with the new blood group with TSL MD approval on file.	
	Due to recent transfusion of blood components that were not identical to the patient's own ABO/Rh	Result the patient's actual blood type and add a BBCS comment explaining the cause of the mixed cell population	
	Resolved by an Immunohematology Reference Lab (IRL)	TSL manager or designee will enter the results	

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STEP	ACTION	
	Unresolved cord blood specimen mixed field where the source is not contaminated	Report the ABO/Rh interpretation as NTD Document the following on the cord blood log o ABO: as "NTD" o Rh: record as POS or NEG based on anti-D reaction results – refer to SOP ABO/Rh Manual Tube Method
2	Add a PB comment indicating the cause of the discrepancy in SQ for future reference NOTE: If the cause of the discrepancy is already noted in the BAD file, the PB comment is not required	
Determine if the new blood type is discrepant with the type of		s discrepant with the type of record in the BAD file Then
	Does match the historical type or the historical type is NTD	No action needed
3	Does NOT match the historical type	Refer the discrepancy to TSL management for evaluation for record correction.
		NOTE: BMT patient's historical record should not be updated following type conversion until the conversion is verified to be complete and approved by the TSL MD

CALIBRATION:

NA

PROCEDURE NOTES AND LIMITATIONS:

Refer to reagent manufacturer's package insert for instructions and limitations for use of reagent

REFERENCES:

- Standards for Blood Banks and Transfusion Services, Bethesda, MD; AABB, current edition
- Technical Manual, Bethesda, MD; AABB, current edition

RELATED DOCUMENTS:

FORM Extended Workup FORM Cord Blood Log SOP Specimen Acceptability and Order Receipt SOP Quality Control for Manual Testing Reagents SOP Labelling for Manual Testing SOP ABO/Rh Manual Tube Method SOP Antibody Identification SOP Grading Reactions SOP Grading Reactions SOP Saline Replacement SOP Weak D Manual Tube Testing SOP EGA Treatment of Red Blood Cells SOP DAT (Direct Antiglobulin Test) by Tube Method SOP Cold Panel

TYPE OF DISCREPANCY	POSSIBLE CAUSES
Mixed-field forward type	 Recent transfusion Transplantation Fetomaternal hemorrhage Twin or dispermic (tetragametic)
	chimerism
Extra reaction in the forward type (includes positive control)	 Autoagglutinins/excess protein coating red cells Unwashed red cells: plasma proteins antibody in patient's serum to reagent constituent Transplantation Acquired B antigen
	 B(A) phenomenon
	Out-of-group transfusion
Negative and/or weak forward type	 ABO subgroup Leukemia/malignancy Recent transfusion Intrauterine fetal transfusion Transplantation Excessive soluble blood group substance
Extra reaction in the reverse type	 Cold autoantibody Cold alloantibody Serum antibody to reagent constituent Excess serum protein Recent transfusion of plasma components Transplantation Infusion of intravenous immune globulin
Negative, weak reverse type	 Age related (<4-6 months old, elderly) ABO subgroup Hypogammaglobulinemia Transplantation
Rh does not match historical Type	 Weak D antigen Differences in antisera specificity Recent transfusion Transplantation

APPENDIX: Possible Causes of ABO Typing Discrepancies

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REVISIONS:

07/21/2020: Add instructions for resolution of mixed field reactions from cord blood and neonatal specimens