



University of Washington Medical Center 1959 NE Pacific Street, Seattle, WA, 98195 Transfusion Services Laboratory Policies and Procedures Manual	Original Effective Date: 02-11-2016	Number: PC-0038.032
	Revision Effective Date: 03-05-2023	
TITLE: Weak D Manual Tube Testing		

PURPOSE:

To provide instructions for performing Weak D testing using a manual tube technique

PRINCIPLE & CLINICAL SIGNIFICANCE:

Principle

Red blood cells from patients are tested with commercial anti-D reagent to determine if the D antigen is present on the patient's red blood cells. In the presence of D antigen, Anti-D reagents bind to the antigen on sample red blood cells and cause an antigen-antibody reaction visible as red blood cell agglutination. RBCs with weakened D antigen may require incubation with the anti-D reagent or testing through the antiglobulin phase for detection.

Clinical Significance

In some recipients determined to be weak D, the possibility exists that pregnancy or transfusion with D+ RBC containing components could result in the production of an allo anti-D.

POLICY

- Weak D testing is performed on the following specimens when the initial Rh result is negative:
 - Cord blood specimens for determination of maternal Rh immune globulin eligibility of a Rh-negative mother
 - Specimens from donors or potential donors of blood or stem cell components (does not include red blood cell component type confirmation testing)
- Weak D testing may also be utilized for ABO/Rh type discrepancy resolution.
- Complete history check regarding prior Rh D typing and transfusion history must be performed in LIS, blood supplier and other hospitals prior to resolution of discrepancy
- For newly identified weak D positive perinatal females:
 - Send the sample to reference lab for weak D type 1,2,3, genotype evaluation
 - Notify the TSL MD on-call 8 am to 5 pm to determine the need for Rh immune globulin.

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SPECIMEN REQUIREMENTS:

EDTA is preferred and if not tested soon after collection, should be stored at 1-6°C
 Red top clotted blood samples are also acceptable.
 See SOP *Specimen Acceptability and Order Receipt*

REAGENTS/SUPPLIES/EQUIPMENT:

Reagents:	Supplies:	Equipment:
<ul style="list-style-type: none"> • Anti-D • ABO + Rh Control • Blood Bank Saline 	<ul style="list-style-type: none"> • 12 x 75 glass tubes 	<ul style="list-style-type: none"> • Calibrated Serologic Centrifuge • Calibrated Cell washer

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<ul style="list-style-type: none"> • Anti-IgG • IgG coated control cells 	<ul style="list-style-type: none"> • Blood bank transfer pipettes 	<ul style="list-style-type: none"> • 37 °C Heat block • Agglutination viewer
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QUALITY CONTROL:

Quality Control is performed each day of use.

INSTRUCTIONS:

STEP	ACTION	
1	Label tube(s) per SOP <i>Labeling for Manual Testing</i>	
	If initial ABO/Rh test tubes are	Then
	Available	Use the D tube and patient cell suspension from ABO/Rh typing to continue testing <ul style="list-style-type: none"> • Label one tube for control • Go to next step
	Not available	<ul style="list-style-type: none"> • Label 3 tubes: one for patient red cell suspension, one for control and one for anti-D • Prepare an approximate 3-4% patient cell suspension • Add the following to the anti-D tube <ul style="list-style-type: none"> ○ 1 drop of Anti-D ○ Add 1 drop of the patient's cell suspension
2	Add 1 drop of the ABO/Rh Control reagent to the control tube	
3	Add 1 drop of the 3-4% patient cell suspension to the control tube	
4	Mix and incubate the anti-D and control tubes at 37°C ± 1 for 15-30 minutes	
5	Wash tubes 3 times with blood bank saline	
6	Add 2 drops of anti-IgG to each tube	
7	Mix and centrifuge	
8	Resuspend the cell buttons gently, and examine for agglutination	
9	Read, grade, and record the reactions	
10	Add 1 drop of IgG Coated Control Cells to all negative tests	
11	Mix and centrifuge	
12	Read, grade, and record the results	
13	Go to section "Interpreting & Reporting Results"	

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CALCULATIONS/INTERPRETATIONS/RESULTS REPORTING/NORMAL VALUES/CRITICAL VALUES

Results Reporting in Sunquest

Step	Action																											
1	Select the appropriate patient specimen in "Blood Order Processing" (BOP)																											
2	Add; DU (Weak D test) in the <u>Add Spec.</u> Test window, if not previously ordered <u>Note: %DU Weak D will be automatically ordered in LIS on patients ≤ 3 days old when Rh status is reported as negative. This can be cancelled if not needed for neonatal specimens per policy</u>																											
3	Enter the results and interpretation																											
	<table border="1"> <thead> <tr> <th colspan="3">If</th> <th rowspan="2">Interpretation</th> <th rowspan="2">Sunquest Key</th> </tr> <tr> <th>Field</th> <th>DAHG</th> <th>DUCC</th> </tr> </thead> <tbody> <tr> <td>PT</td> <td>0</td> <td>+</td> <td rowspan="2">Negative</td> <td rowspan="2">N</td> </tr> <tr> <td>C</td> <td>0</td> <td>+</td> </tr> <tr> <th>Field</th> <th>DAHG</th> <th>DUCC</th> <td rowspan="2">Positive</td> <td rowspan="2">P</td> </tr> <tr> <td>PT</td> <td>+</td> <td>ND</td> </tr> <tr> <td>C</td> <td>0</td> <td>+</td> </tr> </tbody> </table>	If			Interpretation	Sunquest Key	Field	DAHG	DUCC	PT	0	+	Negative	N	C	0	+	Field	DAHG	DUCC	Positive	P	PT	+	ND	C	0	+
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4	Add the Antigen/Antibody code corresponding to Du test interpretation to flag the patient's historical record: <ul style="list-style-type: none"> Enter ;PB (Patient Problem Info) in the <u>Add Spec.</u> Test window Enter the code below: <table border="1"> <thead> <tr> <th>If</th> <th>Enter</th> </tr> </thead> <tbody> <tr> <td>Weak D Positive</td> <td>;WKDP</td> </tr> <tr> <td>Weak D Negative</td> <td>;WKDN</td> </tr> </tbody> </table>	If	Enter	Weak D Positive	;WKDP	Weak D Negative	;WKDN																					
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Step	Action		
			<p>is weak D positive</p> <ul style="list-style-type: none"> • Add a BBC comment: RHREC = Further D antigen characterization should be done through Rh D genotyping to determine the need for Rhogam. If genotyping is not done or not available, patient is a candidate for Rh Immune globulin. • Send the sample to reference lab for weak D type 1,2,3, genotype evaluation • Notify the TSL MD on-call 8 am to 5 pm (to determine the need for Rh immune globulin. <ul style="list-style-type: none"> ○ Document the following in SQ BOP as a BBCS comment: <ul style="list-style-type: none"> ▪ Date and time of notification ▪ Name of TSL MD notified
	Positive	Cord Blood - male	<ul style="list-style-type: none"> • Rh Positive
	Positive	Cord Blood - female	<ul style="list-style-type: none"> • Rh Positive • Add a PB comment indicating the following "Consider Rh negative for transfusion purposes • Notify the TSL MD on call 8 am to 5 pm (to determine the need for Rh immune globulin. • Document the following in SQ BOP as a BBCS comment: <ul style="list-style-type: none"> • Date and time of notification • Name of TSL MD notified

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CALIBRATION:
NA

PROCEDURE NOTES AND LIMITATIONS:

- Calls from patient providers regarding Rh genotyping should be referred to the TSL MD on-call.
- Resulting the Rh type as positive in a weak D positive patient may result in a QA failure that can be overridden using BBR along with a free text comment indicating the "Patient is weak D positive".
- Mixed-field agglutination should be investigated for possible cause – refer to SOP ABO/Rh

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Discrepancy Resolution

- 3-4% suspension of RBCs may be prepared by one of the following:
 - Using volume estimation with comparison to the reagent red cells for visual verification
 - By adding one drop of packed RBCs to approximately 1-1.5 mL of blood bank saline
- Results are considered invalid and must be repeated if negative reactions do not produce agglutination following the addition of IgG Coated Control Cells

REFERENCES:

- Technical Manual. Bethesda, MD: AABB Press, current edition
- Standards for Blood Banks and Transfusion Services. Bethesda, MD: AABB Press, current edition

RELATED DOCUMENTS:

SOP *Specimen Acceptability and Order Receipt*
SOP *Quality Control for Manual Testing Reagents*
SOP *Labeling Tubes and Gel Cards for Testing*
SOP *Grading Reactions*
SOP *ABO/Rh Manual Tube Testing*
SOP *ABO/Rh Discrepancy Resolution*

APPENDICES:

TITLE: **Weak D Manual Tube Testing**

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UWMC SOP Approval:

UWMC CLIA

Medical Director

Andrew Bryan, MD

Date

Transfusion

Service Manager

Nina Sen

Date

QA Manager

Taylor Reeves

Date

Transfusion

Service

Medical Director

Monica Pagano, MD

Date

UWMC Biennial Review:

Date

Date

REVISIONS:

07/21/2021: Clarification when to notify the TSL MD for consideration of genotyping, Rhogam eligibility, and adding "consider genotyping" comment to test results. Updated related SOPs.

[3/1/2023: Updated to clarify weak D policy for non prenatal patients, %DU added as reflex on Rh negative neonatal samples ≤ 3 days old](#)