

Operation

- 1 Spin only balanced loads. Use only Mylar Coated or plastic capillary tubes. Make sure that capillary tubes are placed opposite each other. Proper sample balancing will improve sample separation and will extend the life of the centrifuge. Out-of-balance loads may damage the centrifuge.
- 2 Be certain to follow the instructions on the included vial of capillary tubes when drawing the specimen and sealing the tube. **IMPORTANT:** The tubes must be sealed on the dry end – the end opposite from the one in which the specimen was drawn. Improper technique can result in the sealant and specimen leaking out into the tube shields during centrifugation. Should this happens, the tube shields need to be replaced or removed from the rotor and thoroughly cleaned with hot soapy water and then allowed to dry completely. Failure to do this can cause leakage and/or tube breakage during future runs.
- 3 Set timer for 5 minutes. Unit will begin spinning at full speed (10,000 rpm).
- 4 After unit has stopped, carefully remove the capillary tubes and hold vertically.
- 5 Place the reader card directly behind the tube, and align the top and bottom of the fluid with the 0 and 100 lines.
- 6 The separation line should be read on the graph at the point of the separation.

Note: The separation line will be at an angle, therefore you must read the midpoint of the angle for an accurate percentage.

Specifications

Nominal speed:	10,000 RPM (+/- 500)*
Max. volume:	4 - 75mm capillary tubes
Max. power:	12 volt DC
Height:	4.5 inches
Width:	6 inches
Depth:	8 inches
Weight:	3.3 lbs.
Boxed dimensions:	11.5 x 10 x 7 inches
Boxed weight:	5.5 lbs.

* These are average speeds which vary slightly between units, and with load size. Please use a LW Scientific tachometer for actual RPMs on your unit, and for calibration purposes.

CULTURE-PADDLES® should be immediately replaced in the protective vial, closing the screw cap. The inoculated CULTURE-PADDLE® may be incubated immediately or stored or transported to a laboratory for incubation and/or interpretation. If stored or transported to a laboratory, the URICULT® cap should be tightened. Storage or transportation should not exceed 48 hours at 45...77°F (7...25°C). Stored or transported URicuit should be incubated at 97°F ± 4°F (36°C ± 2°C) for 18–24 hours. URICULT® paddles which have been stored or transported up to 48 hours before incubation can only be used for growth and/or colony count. Transportation of URICULT® Urine CULTURE-PADDLES® with tight caps may result in inconclusive agar color reactions and atypical colony morphology, making presumptive identification impossible.

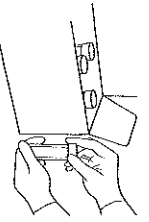
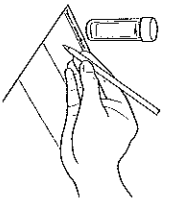

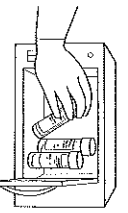
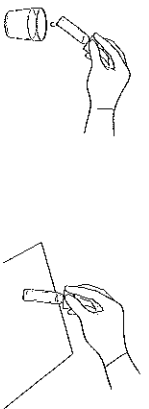
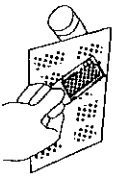
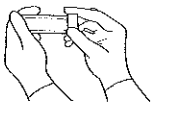
(39°F/4°C) in a closed sterile container. Storage should not exceed 24 hours.

Note
URICULT® test results may be affected if the patient has been receiving antibiotics or anti-infective treatment. Should URICULT® test results show the presence of bacterial growth during the course of therapy, the physician may wish to reassess the dosage or organism susceptibility to the anti-infective being used.

URICULT® testing may be used to assess the effectiveness of antibiotic or anti-infective therapy. In this instance, it is recommended that a test be performed no sooner than 48 hours (2 days) following the administration of the final dose of medication.

Test procedure

Compliance with the following directions is required to achieve reliable test results.

<p>1</p>  <p>Remove the URICULT® Urine CULTURE-PADDLES® from the protective vial by unscrewing the vial cap.</p>	<p>5</p>  <p>Complete patient label indicating patient's name, date and time of inoculation. Attach label to URICULT® vial.</p>
<p>2</p>  <p>Handling the URICULT® Urine CULTURE-PADDLE® by the cap, dip the CULTURE-PADDLE® into the urine specimen to fully immerse the agar surfaces. If the urine volume is not adequate to fully immerse the agar surfaces, as is sometimes the case with infants or small children, the urine may be poured over the agar surfaces.</p>	<p>6</p>  <p>Place inoculated URICULT® vial upright in incubator 97°F ± 4°F (36°C ± 2°C) for 18 to 24 hours. Incubation should not exceed 24 hours. Incubation exceeding 24 hours may cause bacterial overgrowth resulting in difficult interpretation of colony counts and possibly misleading biochemical reactions.</p>
<p>3</p>  <p>Allow the excess urine to drain from the URICULT® Urine CULTURE-PADDLE®. The base of the culture-paddle may be blotted on absorbent paper if desired.</p>	<p>7</p>  <p>Remove URICULT® vial from incubator following incubation period. Compare colony count density on the agar surfaces with the Colony Density Chart provided to obtain a semiquantitative colony count in CFU/ml of urine. Compare only the number of colonies present, not the size of the colonies or the agar surface area they cover. The colonies on the agar surface may also be observed at this time for morphology and agar color reactions which may be used for presumptive identification of the bacterial growth.</p>
<p>4</p>  <p>Replace the inoculated URICULT® Urine CULTURE-PADDLE® in its protective vial.</p>	<p>8</p> <p>Negative cultures may be incubated for an additional 24 hour period, if desired. This will allow for the detection of slow growing bacteria.</p>

Culture disposal

Because bacterial colonies on inoculated URICULT® Urine CULTURE-PADDLES® are actual or potential pathogens, they should not be touched and should not be unduly exposed to other office personnel or patients. It is advised that the procedure to dispose of inoculated culture media be in accordance with existing state or local laws.

To avoid any risk of contamination after a culture has been interpreted, it is also recommended that used URICULT® Urine CULTURE-PADDLES® be promptly and completely immersed in a cup of bactericidal "biocide" solution such as 3% phenol, Staphene® (Vestal Labs) or Cidec® (Surgikos, Inc.).

Bactericidal-treated paddles and protective vials can then be placed in a large wide-mouthed jar or other suitable disposal container filled about 1/3 of its capacity with additional biocide. Keep the container tightly capped and discard when filled.

