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**CANNABINOIDS**

**URINE**

**ABBOTT ARCHITECT**

**Intended Use**

The MULTIGENT Cannabinoids assay is intended for the qualitative and semiquantitative determination of cannabinoids (THC) in human urine on the ARCHITECT *c* Systems. The cutoff for the qualitative application is 50 ng/mL.

The assay is intended for use in clinical laboratories. This assay provides only a preliminary analytical test result. A more specific alternate chemical method must be used to obtain a confirmed analytical result. Gas Chromatography/Mass Spectrometry (GC/MS) is the preferred confirmatory method. Clinical consideration and professional judgment should be applied to any drug of abuse test result, particularly when preliminary positive results are used.

**Clinical Significance**

The principal active agent in marijuana and/or hashish that produces hallucinogenic and other biological effects is generally accepted to be Δ9-tetrahydrocannabinol (Δ9-THC). Δ9-THC is rapidly absorbed and almost completely metabolized by inhalation or through the gastrointestinal tract. Peak plasma levels of Δ9-THC occur within 10 minutes of inhalation and approximately 1 hour after ingestion.

About 70% of a THC dose is excreted within 72 hours with 30% in urine and 40% in feces.3 Passive inhalation of marijuana smoke can result in an elevation of urine THC concentration as high as 10 to 40 ng/mL. In chronic users, THC may accumulate in fatty tissue faster than it can be excreted. This leads to longer detection times in urine for chronic users than for occasional users.

**Principle**

The MULTIGENT Cannabinoids assay is a homogeneous enzyme immunoassay using ready-to-use liquid reagents. The assay uses monoclonal antibodies that detect the major metabolite of Δ9-THC in urine. The assay is based on the competition between an enzyme labeled drug and the drug from the urine for a fixed number of specific antibody binding sites. In the absence of drug from the sample, the specific antibody binds to the drug labeled with glucose-6-phosphate dehydrogenase (G6PDH) and the enzyme activity is inhibited. This phenomenon creates a direct relationship between the drug concentration in the urine and the enzyme activity. The G6PDH enzyme activity is determined spectrophotometrically at 340/412 nm (416 nm for *c*4000 and *c*16000) by measuring its ability to convert nicotinamide adenine dinucleotide (NAD) to NADH.

**Methodology:** Enzyme Immunoassay

**Specimen Collection and Handling**

Collect urine specimens in clean plastic or glass containers. Testing of fresh urine specimens is suggested. Samples within a pH range of 3 to 11 are suitable for testing with this assay.

The Clinical and Laboratory Standards Institute (formerly NCCLS) Urine Drug Testing in the Clinical Laboratory; Approved Guideline states that prior to analysis, urine specimens may be stored at 2 to 8°C for five working days. For longer storage prior to analysis, or for sample retention after analysis, this document recommends freezing at –20°C or less. Laboratories following the Substance Abuse and Mental Health Services Administration (SAMHSA) mandatory guidelines should refer to SAMHSA “Short-Term Refrigerated Storage” and “Long-Term Storage” requirements. Thaw and mix frozen specimens prior to analysis.

**NOTE:** Stored specimens must be inspected for particulates. If present, mix and centrifuge the specimen to remove particulates prior to testing. An effort should be made to keep pipetted samples free of gross debris. It is recommended that highly turbid specimens be centrifuged before analysis. Adulteration of the urine sample may cause erroneous results. If adulteration is suspected, obtain another sample and forward both specimens to the laboratory for testing.

**Materials and Equipment Required**

**TEST INSTRUMENT**: Abbott ARCHITECT System

**MATERIALS PROVIDED**

3L41-20 MULTIGENT Cannabinoids

**MATERIALS REQUIRED BUT NOT PROVIDED**

* 3L43-05 MULTIGENT DOA MC Neg Cal
* 3L41-02 MULTIGENT Cannabinoids 50 Cal

**Reagent Handling and Storage:**

***CAUTION*:**

1. For in vitro diagnostic use.

2. Do not use components beyond the expiration date.

3. Do not mix materials from different kit lot numbers.

**CAUTION:** This product requires the handling of human specimens.

It is recommended that all human sourced materials be considered

potentially infectious and be handled in accordance with the OSHA

Standard on Bloodborne Pathogens. Biosafety Level 2 or other appropriate biosafety practices should be used for materials that contain or are suspected of containing infectious agents.

• The following warning and precaution apply to R1 and R2:

Contains sodium azide.

EUH032 Contact with acids liberates very toxic gas.

These materials and their containers must be disposed of in a safe way.

**Reagent Handling**

• R1 Ready for use. Before use, invert several times, avoiding the formation of bubbles.

• R2 Ready for use. Before use, invert several times, avoiding the formation of bubbles.

• Remove air bubbles, if present in the reagent cartridge, with a new applicator stick. Alternatively, allow the reagent to sit at the appropriate storage temperature to allow the bubbles to dissipate.

To minimize volume depletion, do not use a transfer pipette to remove the bubbles.

**CAUTION:** Reagent bubbles may interfere with proper detection of reagent level in the cartridge, causing insufficient reagent aspiration that could impact results.

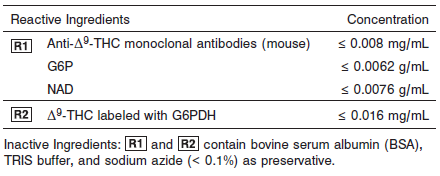
**Reagent Storage**

• Reagent stability is 30 days if the reagent is uncapped and onboard.

• Unopened reagents are stable until the expiration date when stored at 2 to 8°C.

Reagent Preparation:

3L41-20 MULTIGENT Cannabinoids is supplied as a liquid, ready-to-use, two-reagent kit which contains: **R1 & R2**



**Calibrator:**

3L43-05 MULTIGENT DOA MC Neg Cal

3L41-02 MULTIGENT Cannabinoids 50 Cal

**Quality Control:** Minimum 2 levels of DOA UrineControl run every 24 hours

Abbott recommends:

3L41-10 MULTIGENT Cannabinoids 40 Control

3L41-12 MULTIGENT Cannabinoids 60 Control

3L41-14 MULTIGENT Cannabinoids 75 Control

3L41-16 MULTIGENT Cannabinoids 125 Control

**Calibration**

**Frequency:**

Calibration is stable for 13 days (312 hours) for any one lot. Calibration is required with each change in reagent lot number.

**A new calibration is required:**

1. If quality control results do not meet acceptance criteria defined by your laboratory, patient values may be suspect. Follow the established quality control procedures for your laboratory. Recalibration may be necessary.
2. Review quality control results and acceptance criteria following a change of reagent or calibrator lot.

**Calibrator Required:**

* 3L43-05 MULTIGENT DOA MC Neg Cal
* 3L41-02 MULTIGENT Cannabinoids 50 Cal

**Reagents:**

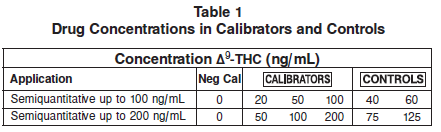
The MULTIGENT Cannabinoids Calibrators and Controls are liquid and ready-to-use. They are prepared by spiking negative human urine with known quantities of 11-nor-Δ9-THC-9-carboxylic acid (Δ9-THC). Stabilizer is also present.

For qualitative analysis, the MULTIGENT DOA MC Neg Cal and the MULTIGENT Cannabinoids 50 Cal are used for system calibration. The concentration of the MULTIGENT Cannabinoids 50 Cal determines the cutoff between “positive” and “negative” samples. The MULTIGENT

Cannabinoids 40 and 60 Controls are used to validate qualitative assay performance.

When an estimate of drug concentration is required, a calibration curve can be established up to 100 ng/mL or 200 ng/mL. The MULTIGENT Cannabinoids 40, 60, 75, and 125 Controls validate semiquantitative assay performance as indicated in Table 1.

The MULTIGENT DOA MC Neg Cal is a drug-free human urine pool that is used as zero drug calibrator and can also be used for dilution of samples exceeding the highest calibrator.



**Calibrator Preparation:**

The MULTIGENT Cannabinoids Calibrators and Controls are ready-to-use.

**Calibration Procedure:**

Before performing the assay, refer to the ASSAY PARAMETERS which are included in the MULTIGENT Cannabinoids reagent package insert.

The parameters contain additional instructions for using qualitative and semiquantitative protocols. For further instructions refer to the CALIBRATION and QUALITY CONTROL sections of the MULTIGENT Cannabinoids reagent package insert.

1. Verify that the calibrator values are correct in the instrument parameter files.

2. Mix bottles by gentle inversion several times.

3. Open the bottles, place appropriate amounts of the required calibrators and/or controls in separate sample cups, and place in the assigned positions.

4. Calibrate as outlined in *Section 6* of the **ARCHITECT System Operations Manual**.

5. Follow the established quality control procedures for your laboratory and the instructions found in *Section 5* of the **ARCHITECT System** **Operations Manual**.

6. Cap bottles tightly and return to refrigerated storage after use.

7. Verify control results are within acceptable limits before reporting patient results.

**RESULTS**

The MULTIGENT Cannabinoids 50 Cal is used as a qualitative cutoff reference for distinguishing between “positive” and “negative” samples. A sample that gives a change in absorbance rate (ΔmAU/min) equal to or greater than the MULTIGENT Cannabinoids 50 Cal is considered positive.

A sample that gives a change in absorbance rate (ΔmAU/min) less than that obtained with the MULTIGENT Cannabinoids 50 Cal is considered negative.

When an estimate of drug concentration is required, a calibration curve can be established up to 100 ng/mL or 200 ng/mL. See Table 1 for calibrators and controls required for each range. The concentration of the sample can be obtained by quantitation from the calibration curve. When the sample concentration is greater than the highest calibrator, it may be diluted with the MULTIGENT DOA MC Neg Cal and retested.

**Troubleshooting and Overall Acceptance Criteria Failure**

See ARCHITECT Operations Manual for further calibration troubleshooting.

**Quality Control:**

Abbott recommends, refer to your laboratory standard operating procedure(s) and/or quality assurance plan for additional quality control requirements and potential corrective actions:

• Two levels of controls (normal and abnormal) are to be run every 24 hours.

• If more frequent control monitoring is required, follow the established quality control procedures for your laboratory.

• If quality control results do not meet the acceptance criteria defined by your laboratory, patient values may be suspect. Follow the established quality control procedures for your laboratory.

Recalibration may be necessary.

• Review quality control results and acceptance criteria following a change of reagent or calibrator lot.

**Procedure**

For a detailed description of how to run an assay, refer to *Section 5* of the **ARCHITECT System Operations Manual**.

**Qualitative Analysis**

For qualitative analysis, the MULTIGENT DOA MC Neg Cal and the 3L41-02 MULTIGENT Cannabinoids 50 Cal (50 ng/mL 11-nor-Δ9-THC-9-carboxylic acid) are used for system calibration. The 3L41-02 MULTIGENT Cannabinoids 50 Cal determines the cutoff between “positive” and “negative” samples. Suggested negative and positive controls are the MULTIGENT Cannabinoids 40 Control and the MULTIGENT Cannabinoids 60 Control.

**Calculations**

Refer to *Appendix C* of the **ARCHITECT System Operations Manual** for information on results calculations.

**Reporting Results**

**Qualitative Results:** Cutoff = 50 ng/mL

A sample that exhibits a change in absorbance rate (ΔmAU/min) equal to or greater than the value obtained with the cutoff calibrator is considered positive.

A sample that exhibits a change in absorbance rate (ΔmAU/min) lower than the value obtained with the cutoff calibrator is considered negative.

**Specific Performance Characteristics**

**Performance Characteristics**

**Sensitivity**

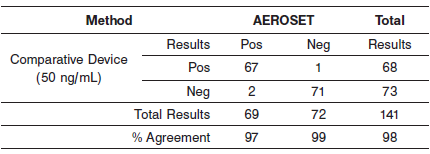
Using the parameters for the semiquantitative range of up to 100 ng/mL or cutoff of 50 ng/mL, the sensitivity is 10.0 ng/mL.

**Accuracy**

A 15 ng/mL cutoff was used for GC/MS. The Cannabinoids assay showed a 100% correlation with GC/MS technique when a 50 ng/mL cutoff calibrator was used..

**Qualitative**

There was 98% overall agreement between the two methods.



**Dilution:**

Samples with semiquantitative results that exceed the highest calibrator may be manually diluted and rerun. Acceptable sample diluents are the MULTIGENT DOA MC Neg Cal or drug-free human urine.

The dilution must be performed so the diluted test results read greater than 10.0 ng/mL.

The operator must enter the manual dilution factor in the patient or control order screen. The system uses this dilution factor to automatically correct the concentration by multiplying the result by the entered factor. The printed result is the reportable result if no errors are present.

**NOTE:** If the operator does not enter the manual dilution factor, the

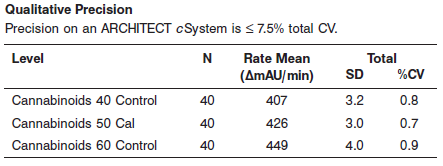
printed result must be multiplied by the manual dilution factor before

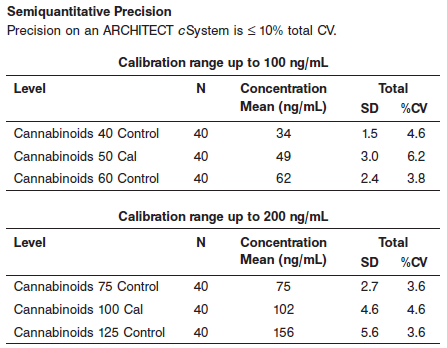
reporting the result..



For detailed information on ordering dilutions, refer to *Section 5* of the **ARCHITECT System Operations Manual**.

**Precision:**





#### Limitations of Procedure

• The assay is designed for use with human urine only.

• A positive result indicates only the presence of cannabinoids and does not necessarily correlate with the extent of physiological and psychological effects.

• A positive result by this assay should be confirmed by a chemical method such as GC, Thin Layer Chromatography (TLC), or GC/MS.

• It is possible that other substances and/or factors (e.g., technical or procedural) not listed in the specificity table may interfere with the test and cause false results.

• When dispensing samples containing THC:

– Do not use disposable polyethylene transfer pipettes.

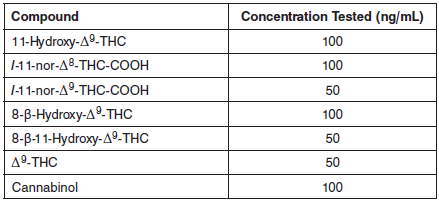
– Use a minimum of 600 μL of calibrator, control, or sample in sample cups.

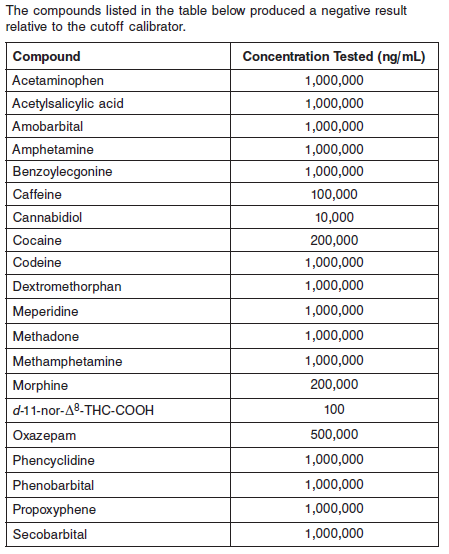
– Do not redispense solutions from a pipette back into a container (includes redispensing after wetting a pipette tip).

– In order to minimize the contact of the pipette tip, immerse pipette tips below the surface of the liquid only far enough to aspirate the needed volume.

**Specificity**

Various potentially interfering substances were tested for cross-reactivity with the assay. The compounds listed in the table below produced a result approximately equivalent to the cutoff calibrator.





**References:**

1. ABBOTT ARCHITECT Cannabinoids package insert

Abbott Laboratories

Diagnostics Division

Abbott Park, IL 60064

May 2014 306376/R08

1. ABBOTT ARCHITECT Cannabinoids Calibrator package insert

Abbott Laboratories

Diagnostics Division

Abbott Park, IL 60064

1. Abbott ARCHITECT Operator’s Guide

**Related Documents:**

**Attachments:**