Department Meeting Notes 2/15/22

Wins:

CAP surveys (MRSA/MSSA, bacteriology, BacT Alert, Covid PCR, India Ink, Crypto/Giardia, Mono, RA) all correct! One of our BCID's was incorrect-we reported "no organism detected" and it was supposed to contain Listeria monocytogenes. I re-ran the sample and got the correct answer. We must make sure we thoroughly mix all samples and fill the pipet to the correct line with sample. Also, make sure the sample pouch is filling appropriately. Sometimes they lose vacuum and don't fill correctly.

Technical:

Please confirm ESBL and CRE calls from the Vitek in addition to setting discs, MCIM. If Vitek calls ESBL but marker drugs (Cephalosporins, Aztreonam) look o.k., flip interps back to susceptible. We had issues in the past with Proteus mirabilis being called an ESBL due to Amoxicillin/Clavulanate or Ampicillin being resistant and 10-disc confirmations were always negative on these isolates.

We are going to start eliminating the OD tubes from Proteus identifications. If we have a swarmer, preliminarily sign it out as "Proteus species". No spot testing is required. Order a definitive I.D. on the Vitek. We are prematurely signing these isolates out as mirabilis before we even have the OD result. Pseudomonas can be treated the same. Spready, fruity hemolytic GNR's can be signed out as Pseudomonas species and a definitive I.D. ordered. We can only call a Pseudomonas aeruginosa if it is grapey and has a metallic sheen.

Do not report "rare normal flora" in stool cultures. If present, normal flora does not need to be mentioned. A smart phrase will be built to report "no normal enteric flora present", as this could be significant to clinicians. Only report yeast and Staph aureus if in pure culture.

We are going to reserve Isolate 1 for normal respiratory flora in respiratory specimens. Organism name is already built to quantitate and report "normal respiratory flora present, including....." so we can list organisms we are not working up in addition to noting the quantity of normal flora.

Blood culture anaerobic bottle sub plates are to be read at 24 hours and 48 hours and these reads documented. Other anaerobic plates (body fluids, wounds, etc.) will be read/documented at 48 and 72 hours. This way the anaerobic plates can be compared to the thio subs. We will also read/document the Campy plates at 48 and 72 hours.

Thio broths are to be subbed to aerobic bap and anaerobic bap. Any growth we recover on any anaerobic media must be subbed to both aerobic and anaerobic media to prove a true anaerobe.

We are going to start subbing the following QC bugs daily so we have fresh organisms to set along with patients:

Strpne 49619, Pseaer 27853, Klepne BAA 1705 and 1706, Esccol 25922.

Doing this will save a day in reporting sensitivities when we have a patient that needs a Kirby Bauer set.

Please begin using wire loops and bacticinerator for streaking out specimen plates. Better isolation is achieved when the loop is flamed in between plates and there is less of a chance of introducing contaminants from an opened pack of disposable loops. This is extremely important when streaking out sterile specimens.

Schedule:

Email me your thoughts on having Christmas as the only holiday worked in a year and dividing the rest of the holidays (New Year's Day, MLK Day, Memorial Day, July 4th, Labor Day, Thanksgiving). We need to add Trish to the holiday rotation and need a consensus on how we're going to divide the holidays up.