

UPMC HANOVER

MICROBIOLOGY SPECIMEN PROCESSING CHART

| SPECIMEN | GRAM STAIN | AEROBIC MEDIA | ANAEROBIC MEDIA | COMMENTS |
|------------------------------------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------|-----------------------------------------------------------------------|-----------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Abscess | X | BAP, CHOC, MAC | BRUC, LKV, BBE, and Cooked meat | Rectal, perirectal, decubitus ARE NOT acceptable for Anaerobic cultures. |
| Sterile Body Fluid- Synovial, Pleural, Pericardial. Set Blood cultures Bottles. If inadequate specimen for bottles, set up directly on media. | X For thick specimens add 2 drops to 1mL sterile saline and cytospin. | BAP, CHOC, MAC Add Thayer Martin to synovial fluid if GC suspected | BMB, LKV, BBE, and Cooked meat (set anaerobic media directly from specimen) | For <3mL fluid spin at 3400 rpm for 15 minutes. Plate sediment to aerobic media. Minimum volume for blood culture bottles is 5 ml per bottle. Maximum 10 ml per bottle. |
| Non-Sterile Body Fluid- Abdominal Ascites, Peritoneal, Aspirates. | X Cytospin if not too thick/bloody, otherwise, make direct slides. | BAP, CHOC, MAC, | BMB, LKV, BBE, and Cooked meat (set anaerobic media directly from specimen) | |
| Bone Marrow | X | <5mL - BAP, CHOC >5mL AER bottle | <5mL - BRUC, Chopped meat >5mL- ANAE bottle | Colony count streak (urine streak) |
| Bronchial Lavage/ Bronchial Wash | X | BAP, CHOC, MAC | | 4 quadrant streak |
| Bronch Brush | X | BAP, CHOC, MAC | | Use green loop (1 uL) and do Colony count streak. Use 1 loop/plate |
| Cath Tip | | BAP | | Aseptically roll across surface of BAP |
| CSF | X Cytospin | BAP, CHOC, Cooked meat (set direct) | | Do not spin |
| Dialysis Fluid – Set Blood cultures bottles | X | BAP, CHOC if <5 mL | BMB, LKV, BBE, Cooked meat (set anaerobic media directly from specimen) | |

UPMC HANOVER

| SPECIMEN | GRAM STAIN | AEROBIC MEDIA | ANAEROBIC MEDIA | COMMENTS |
|--------------------------------------------------|------------|-----------------------------------------------------------------------------------------------------------------------------------------------|-----------------|----------------------------------------------------------------|
| Ear | X | BAP, CHOC, MAC | | If one swab, no gram stain |
| Eye | X | BAP, CHOC, MAC | | If one swab, no gram stain |
| Upper Respiratory (Sinus, Nasal) | | BAP, CHOC, MAC | | |
| Group A Strep (Throat Culture) | | SSA w/Taxo A (Bacitracin disk) in primary streak | | |
| Group B Strep (Vag/Rectal) | | Swab in to LIM Broth and snap | | Incubate 18 to 24 hours |
| Gonorrhoeae Culture (throat, rectal, genital) | | Choc, Thayer Martin | | E. swab or Amies |
| Gonorrhoeae/ Chlamydia PCR (Cepheid) | | Females: Pink Cepheid collection tube (vaginal/cervical) or first stream urine (<50mL) Males: First stream urine (<50mL) | | Add “.UR50” in “Comments” box when >50mL urine received. |
| Genital Culture (Cervical, Vaginal, Penis) | X | BAP, CHOC, MAC, THAYER MARTIN | | |
| Bacterial Vaginosis/Wet Prep | X | Add 1 drop specimen to slide and coverslip-read @ 40X | | Specimen must be received w/in 30 minutes of collection. |

UPMC HANOVER

| SPECIMEN | GRAM STAIN | AEROBIC MEDIA | ANAEROBIC MEDIA | COMMENTS |
|-------------------------------------------|------------|--------------------------------------------------------------------------------------------------------------|----------------------------|------------------------------------------------------------------------------------------------------------|
| IUD (Intra-uterine device) | | BAP, CHOC, MAC | BMB, LKV, BBE, Cooked meat | Add broth to specimen container, incubate overnight. Set media the next day. |
| Medical Device | | BAP, CHOC, MAC | BMB, LKV, BBE, Cooked meat | Add broth to specimen container, incubate overnight. Set media the next day. |
| Lower Respiratory (Sputum/Trach aspirate) | X | BAP, CHOC, MAC | | |
| Stool/ Shiga toxin | | Preserve in Orange top Cary Blair | | Sent to QUEST |
| Parasites (Stool) | | Preserve in Black Top Para-Pak-Fill to line | | Sent to QUEST. |
| Cryptosporidia/ Giardia | | Preserve in Black Top Para-Pak-Fill to line | | Sent to QUEST |
| C. difficile | | Stool with no preservatives. Consistency watery, liquid or soft. Stool must take the shape of the container. | | Run on Cepheid w/in 24 hours of collection. Test refrigerated specimens w/in 5 days. Reject formed stools. |
| Tissue/Bone/ Surgical Wound | X | BAP, CHOC, MAC | BMB, LKV, BBE, Cooked meat | Add PEA plate to placenta cultures. |
| Wound | X | BAP, CHOC, MAC | BMB, LKV, BBE, Cooked meat | |

Document Information

Document Title

Microbiology Specimen Processing Chart

Document Description

N/A

Approval Information

Approved On: 06/25/2024

Approved By: Cindy Sturtz (Hanover - Laboratory Medical Director: sturci00) on 06/25/2024

Approval Expires: 06/20/2026

Approval Type: Process

Document Location: / Laboratory - Hanover / Hanover - Microbiology

Keywords: Micro., Setup,

Printed By: Sherilyn Solanick

Standard References: N/A

Note: This copy will expire in 24 hours