**UW Medicine - Pathology**

400-04-01-07

De-Staining of G-Banded Slides for Fluorescence in Situ Hybridization (FISH)

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| Adopted Date: 08/20/02Review Date: 06/12/09Revision Date: 04/26/11 |

PURPOSE

This is a method that allows FISH to be done on slides that have already been G-banded. Once de-stained, the appropriate FISH protocol is to be used with the probe of choice.

PROCEDURE

### Materials and Solutions

### Xylene subsititute

### Coplin jars

### Methanol

### Acetic acid

### 3% buffered formalin (Add 15 ml of 10% formalin to 35 ml PBS). Discard after 3 months.

### 0.2% HCl. Discard after 3months.

### 2X SSC *Note:*  For SSC recipes see Uncultured Amniocytes procedure.

### De-staining Procedure

### Remove oil from the slide by soaking in Xylene substitute for 10 min and dry.

### De-stain the slide by soaking it in 40 ml of 3:1 fixative for 5 min and dry.

### Put the slide into a Coplin jar containing 3% buffered formalin for 2.5 min.

### Put the slide into a Coplin jar containing 0.2% HCl for 5 min.

### Put the slide into a Coplin jar containing 2X SSC at room temperature for at least 5 mins.

### Dehydrate slide 1 min each in 70%, 80%, 95%, 100% EtOH jars.

### Probe slide as per the appropriate FISH protocol.

REFERENCES

Wilbur Harrison, Applied Cytogenetics, vol. 21, number 6, 1995.

Written By: Director Approval:

(Signature and Date) (Signature and Date)

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 Cytogenetics Supervisor

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**SIGNATURE PAGE FOR POLICIES AND PROCEDURES**

Procedure / Policy Title: De-Staining of G-Banded Slides for Fluorescence in Situ Hyridizaton (FISH)

Procedure / Policy Number: 400-04-01-07

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