Reducing Hemolysis in Specimen Collection and Processing

This presentation was prepared by the ACL Specimen Collection, Processing, and Exception Handling Technical Advisory Team

Last reviewed 8/12/2022 SJP/SDV



Objectives:

This presentation applies to new and experienced phlebotomists, laboratory assistants, and all team members involved in collection or processing of blood specimens.

- Improve patient safety by reducing specimen rejection due to hemolysis to ensure accurate results
- Define hemolysis and its causes
- Tips for prevention and reduction of hemolysis
- Tips for specimen processing
- Best practices when using butterfly/wing-set collection devices



Possible Outcomes if Best Practices Are Not Followed

- Patient safety is compromised
- Testing delay
- Delay in treatment / diagnosis / discharge
- Redraws / additional patient sticks
- Rework
- Patient Dissatisfaction
- Physician Dissatisfaction



Venipuncture Best Practices

- Recommended needle size is 21 gauge. Smaller gauges may be used in difficult or pediatric venipuncture situations, but have higher chance of hemolysis.
- Cleanse site with alcohol and allow site to air dry
- Never leave tourniquet on for more than 60 seconds
- Patient should be asked to close their hand <u>without</u> clenching or pumping
- Instruct patient to open hand once blood starts flowing into vacutainer tube(s)
- Avoid slow draws/poor blood flow from improperly positioned needles
- In the event a syringe needs to be used for the collection, the plunger must be pulled back slowly and gently to prevent hemolysis



Venipuncture Best Practices (continued)

- Tubes must be collected via venipuncture in the correct order of draw to avoid contamination from preceding tube:
- Blood Cultures
- Blue top tubes (sodium citrate)
- Gold top tubes (serum tube with or without gel)
- Red top tubes (serum tube without gel)
- Green tubes (heparin gel)
- Green tubes (heparin no gel)
- Lavender top tubes (EDTA)
- Pink top tubes (EDTA)
- Gray top tubes (glycolytic inhibitor)
- Royal blue top tubes(with or without additive)



TUBES NOT LISTED ABOVE SHOULD BE DRAWN AT THE END OF THE COLLECTION SEQUENCE If you have any questions, contact your site laboratory.



Venipuncture Best Practices (continued)

- Any additional tubes, for example (ACD Yellow, Quantiferon Kit, etc..)
- Note: If a winged butterfly set is used and a coagulation tube needs to be drawn and a blood culture sample is not required, use a no-additive or sodium citrate discard tube first.
- Note: If a capillary collection is completed, the order of draw is: EDTA > Lithium Heparin (no gel) > Lithium Heparin (with gel) > Sodium Fluoride > Clot Activator and gel for serum separation > No Additive.





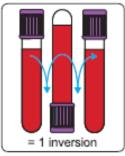
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Venipuncture Best Practices (continued)

- Immediately after each tube is filled, invert tubes <u>gently</u> the appropriate # of times to ensure thorough mixing:
 - Blue 3-4 times each
 - Gold/Red 5 times each
 - All other tube types 8 times each
- *DO NOT SHAKE TUBES!*
 - Red cells can be damaged and release potassium and other cell contents that can impact results
 - Quantiferon Collection Kit is an exception

Note: If not properly mixed, fine clots may form in tubes containing anticoagulants that may seriously interfere with all testing.





Processing Best Practices

- Allow gel top tubes to clot in an upright position for 30 minutes at room temperature; no longer than 2 hours before centrifuging
- Allow red top tubes to clot in an upright position for 60 minutes at room temperature; no longer than 2 hours before centrifuging
- Specimens must be centrifuged for the appropriate time at the appropriate speed
- Do not re-spin blood in a gel tube after it has been centrifuged to recover additional serum
- Do not refrigerate specimens prior to centrifugation
 - Refrigeration will elevate potassium levels quickly, significantly, and irreversibly
- Do not freeze blood in a gel tube or freeze a serum aliquot if red cells are present.



Freezing will release potassium from red cells

Butterflies (also known as: winged infusion sets)

- Butterfly usage should be kept to a minimum and used with discretio they are inherently higher risk for hemolysis
 - When collecting blood cultures
 - When it is a difficult venipuncture
 - Small/difficult veins
 - Hand veins
 - When a patient is insistent
 - Pediatric patient
- 21 Gauge butterfly is the preferred size to prevent hemolysis
- Reasons <u>not</u> to use butterflies:
 - Butterflies with a gauge of 22 or higher are not indicated for certain types of draws (i.e. potassium) because the smaller diameter of the needle may damage red cells
 - Increased risk of needle stick incident
 - Higher cost/expense









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Butterflies (continued)

Misconceptions:

- Drawing multiple tubes is easier with a butterfly.
 - Truth: with proper technique, drawing multiple tubes with a standard, straight needle is the same
- Using a butterfly causes less discomfort.
 - Truth: discomfort is caused by poor technique and certainly not by the length of the needle
 - Butterflies are easier to use and control
 - Truth: more accidental needle sticks occur while performing venipunctures with a butterfly (source: *Medical Data International*)



Positive Patient Outcomes

- Increased Patient Safety by:
 - \checkmark Providing timely test results and treatment
 - ✓ Providing successful venipunctures
 - ✓ Following best practices
- Increased Patient Satisfaction
- Increased Physician Satisfaction



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